

DISTRIBUTION AND FEEDING PREFERENCES OF
FREE-LIVING NEMATODES ASSOCIATED WITH
THE KELP, Macrocystis integrifolia (LAMINARIALES)

by

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THE REQUIREMENTS FOR THE DEGREE OF
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Distribution and feeding preferences of free-living nematodes
associated with the kelp, Macrocystis integrifolia (Laminariales)

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ABSTRACT

Free-living nematodes are a major component of the fauna of marine interstitial environments and are frequently associated with intertidal and subtidal algae. A study was done to categorize the nematodes of a kelp, Macrocystis integrifolia Bory, in the Bamfield region of Barkley Sound, British Columbia, Canada. The feeding preferences of the predominant free-living nematode species found on the M. integrifolia blades were evaluated.

Nine species (belonging to six families) of nematodes were found on the blades and three of them comprised 91-99% of the nematode fauna. Two monhysterids (Monhystera disjuncta and M. refringens) and a chromadorid (Prochromadorella neapolitana) occurred in all monthly samples. About 50% of the nematodes on the blades from July to October was M. refringens, which was at its peak population level during these months. During the fall, the M. refringens population declined to low levels prior to a progressive increase in population commencing in February. M. disjuncta was relatively abundant throughout the year and, in particular, during the late winter and early spring. P. neapolitana occurred in relatively large numbers in July and subsequently declined to a very low population in the winter and spring.

Nematode distribution on M. integrifolia appeared to be related to blade age and the associated food sources on the blade. Both monhysterids occurred in greatest abundance on the bottom blades of Macrocystis plants and on the middle

blades of plants in deep water. Low numbers of P. neapolitana occurred throughout the entire depth gradient on both bottom and middle blades. Few of these three species occurred on the top blades. Nematode species other than these three contributed little to the abundance and distribution patterns observed on the Macrocystis blades.

All three predominant nematode species showed specific responses to available bacterial and diatom food sources in feeding experiments. P. neapolitana exhibited a significant preference for diatoms, particularly Cocconeis scutellum and Gramatophora marina, over the bacteria species under both simulated summer and winter conditions. M. refringens showed no preference towards bacteria or diatoms but exhibited some preference for particular diatom or bacteria species. Under both environmental simulations M. disjuncta showed a marked preference for rod-shaped bacteria.

The probability that the seasonal abundance and distribution of these marine nematodes is correlated with the seasonality of their food sources, bacteria and diatoms, is suggested.

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TABLE OF CONTENTS

| | Page |
|---|------|
| Title Page | i |
| Approval Page | ii |
| Abstract | iii |
| Acknowledgements | v |
| Table of Contents | vi |
| List of Tables | viii |
| List of Figures | xi |
| List of Appendices | xiv |
| INTRODUCTION | 1 |
| DESCRIPTION OF STUDY SITE AND ASSOCIATED KELP | 8 |
| MATERIALS AND METHODS | 12 |
| I. Field Survey | 12 |
| a) Sampling Procedure | 12 |
| b) Treatment of Samples | 13 |
| c) Statistical Treatment | 14 |
| d) Environmental Parameters | 14 |
| e) Taxonomic notes | 14 |
| II. Feeding Preference Experiments | 16 |
| a) Culture of Nematodes | 16 |
| b) Culture of Food Sources | 17 |
| c) Experimental Design | 19 |
| d) Statistical Treatment | 21 |

| | Page |
|---|------|
| RESULTS I | 22 |
| a) Observations on Physical Parameters | 22 |
| b) Nematode Species Composition | 22 |
| c) Seasonal Species Changes | 22 |
| d) Species Distribution on Kelp | 34 |
| DISCUSSION I | 56 |
| RESULTS II | 63 |
| a) <u>Prochromadorella neapolitana</u> | 63 |
| b) <u>Monhystera refringens</u> | 68 |
| c) <u>Monhystera disjuncta</u> | 72 |
| d) Life Cycle Observations on <u>Monhystera</u> <u>disjuncta</u> | 78 |
| DISCUSSION II | 80 |
| GENERAL DISCUSSION | 84 |
| APPENDICES | 89 |
| REFERENCES | 113 |

LIST OF TABLES

| Table | | Page |
|-------|--|------|
| 1 | Diatom (D1-D8) and bacteria (B1-B4) species isolated from <u>Macrocystis integrifolia</u> blades and used in the feeding preference experiments. | 18 |
| 2 | Checklist of free-living nematodes found in blade samples of <u>Macrocystis integrifolia</u> . | 24 |
| 3 | Total number/0.5 m ² and per cent abundance of males, females and juveniles of <u>Prochromadorella neapolitana</u> over sampling period. | 27 |
| 4 | Total number/0.5 m ² and per cent abundance of males, females and juveniles of <u>Monhystera refringens</u> over sampling period. | 29 |
| 5 | Total number/0.5 m ² and per cent abundance of males, females and juveniles of <u>Monhystera disjuncta</u> over sampling period. | 32 |
| 6 | Distribution of <u>Prochromadorella neapolitana</u> with reference to <u>Macrocystis integrifolia</u> blade position and depth F values for a grouped two-level ANOVA calculated on the number of nematodes/0.5 m ² for each sample month. | 51 |
| 7 | Distribution of <u>Monhystera refringens</u> with reference to <u>Macrocystis integrifolia</u> blade position and depth F values for a grouped two-level ANOVA calculated on the number of nematodes/0.5 m ² for each sample month. | 52 |
| 8 | Distribution of <u>Monhystera disjuncta</u> with reference to <u>Macrocystis integrifolia</u> blade position and depth F values for a grouped two-level ANOVA calculated on the number of nematodes/0.5 m ² for each sample month. | 53 |
| 9 | Distribution of the three predominant nematode species with reference to <u>Macrocystis integrifolia</u> blade position and depth F values for an unreplicated three-level ANOVA calculated on the number of nematodes/0.5 m ² for each sample month. | 54 |

| Table | | Page |
|-------|--|------|
| 10 | Total male and female mean accumulation of <u>Prochromadorella neapolitana</u> to diatom and bacteria species (see Table 1) under simulated summer conditions. | 64 |
| 11 | Analysis of attractiveness and preference of <u>Prochromadorella neapolitana</u> to diatom and bacteria species (see Table 1) using Chi-square two-level ANOVA and Student-Newman-Kuels testing under simulated summer and water conditions. | 65 |
| 12 | Total male and female mean accumulation of <u>Prochromadorella neapolitana</u> to diatom and bacteria species (see Table 1) under simulated winter conditions. | 67 |
| 13 | Total male and female mean accumulation of <u>Monhystera refringens</u> to diatom and bacteria species (see Table 1) under simulated summer conditions. | 69 |
| 14 | Analysis of attractiveness and preference of <u>Monhystera refringens</u> to diatom and bacteria species (see Table 1) using Chi-square, two-level ANOVA and Student-Newman-Kuels testing under simulated summer and winter conditions. | 70 |
| 15 | Total male and female mean accumulation of <u>Monhystera refringens</u> to diatom and bacteria species (see Table 1) under simulated winter conditions. | 71 |
| 16 | Total male and female mean accumulation of <u>Monhystera disjuncta</u> to diatom and bacteria species (see Table 1) under simulated summer conditions. | 73 |
| 17 | Analysis of attractiveness and preference of <u>Monhystera disjuncta</u> to diatom and bacteria species (see Table 1) using Chi-square, two-level ANOVA and Student-Newman-Kuels testing under simulated summer and winter conditions. | 74 |
| 18 | Total male and female mean accumulation of <u>Monhystera disjuncta</u> to rod and coccoid bacteria species (see Table 1) under simulated summer conditions. | 76 |

| Table | | Page |
|-------|--|------|
| 19 | Total male and female mean accumulation of <u>Monhystera disjuncta</u> to diatom and bacteria species (see Table 1) under simulated winter conditions. | 77 |
| 20 | Duration of development and generation time of <u>Monhystera disjuncta</u> in cultures maintained at simulated summer conditions. | 79 |

LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 1 | Location of Dodger Channel study site in the Barkley Sound region of Vancouver Island, B.C. | 9 |
| 2 | Semi-diagrammatic sketch of <u>Macrocystis</u> sp. | 10 |
| 3 | Temperature and salinity in Dodger Channel over the sampling period 1978 - 1979. | 23 |
| 4 | Per cent distribution and age structure of <u>Prochromadorella neapolitana</u> over the sampling period 1978 - 1979. | 25 |
| 5 | Per cent distribution and age structure of <u>Monhystera refringens</u> over the sampling period 1978 - 1979. | 28 |
| 6 | Per cent distribution and age structure of <u>Monhystera disjuncta</u> over the sampling period 1978 - 1979. | 31 |
| 7 | Per cent distribution and age structure of other non-predominant nematode species over the sampling period 1978 - 1979. | 33 |
| 8 | Density of <u>Prochromadorella neapolitana</u> (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect gradient from July 1978 to October 1978 (from deep to shallow depth). | 35 |
| 9 | Density of <u>Prochromadorella neapolitana</u> (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect from November 1978 to March 1979 (from deep to shallow depth). | 36 |
| 10 | Density of <u>Prochromadorella neapolitana</u> (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect from April 1979 to July 1979 (from deep to shallow depth). | 37 |
| 11 | Density of <u>Prochromadorella neapolitana</u> (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect for September and November 1979 (from deep to shallow depth). | 38 |

- 12 Density of Monhystera refringens (nematodes/
0.5 m²) on top, middle and bottom blade
samples of Macrocystis integrifolia along a
depth transect from July 1978 to October
1978 (from deep to shallow depth).
- 13 Density of Monhystera refringens (nematodes/ 40
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect from November 1978 to March 1979
(from deep to shallow depth).
- 14 Density of Monhystera refringens (nematodes/ 41
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect from April 1979 to July 1979 (from
deep to shallow depth).
- 15 Density of Monhystera refringens (nematodes/ 42
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect for September and November 1979
(from deep to shallow depth).
- 16 Density of Monhystera disjuncta (nematodes/ 43
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect from July 1978 to October 1978.
- 17 Density of Monhystera disjuncta (nematodes/ 44
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect from November 1978 to March 1979
(from deep to shallow depth).
- 18 Density of Monhystera disjuncta (nematodes/ 45
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect from April 1979 to July 1979 (from
deep to shallow depth).
- 19 Density of Monhystera disjuncta (nematodes/ 46
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth
transect for September and November 1979 (from
deep to shallow depth).
- 20 Density of non-predominant nematode species 47
(nematodes/0.5 m²) on top, middle and bottom
blade samples of Macrocystis integrifolia along
a depth transect from July 1978 to October 1978
(from deep to shallow depth).

| Figure | | Page |
|--------|--|------|
| 21 | Density of non-predominant nematode species (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect from November 1978 to March 1979 (from deep to shallow depth). | 48 |
| 22 | Density of non-predominant nematode species (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect from April 1979 to July 1979 (from deep to shallow depth). | 49 |
| 23 | Density of non-predominant nematode species (nematodes/0.5 m ²) on top, middle and bottom blade samples of <u>Macrocystis integrifolia</u> along a depth transect for September and November 1979 (from deep to shallow depth). | 50 |

LIST OF APPENDICES

| Appendix | | Page |
|----------|---|------|
| 1 | Selective accumulation of <u>Prochromadorella neapolitana</u> to alternating wells of diatom species D1-D4 (see Table 1). Diagram records four replicates for each condition. | 90 |
| 2 | Selective accumulation of <u>Prochromadorella neapolitana</u> to control wells. | 91 |
| 3 | Selective accumulation of <u>Prochromadorella neapolitana</u> to alternating wells of diatom species D5-D8 (see Table 1). Diagram records four replicates for each condition. | 92 |
| 4 | Selective accumulation of <u>Prochromadorella neapolitana</u> to control wells. | 93 |
| 5 | Selective accumulation of <u>Prochromadorella neapolitana</u> to alternating wells of bacteria species B1-B4 (see Table 1). Diagram records four replicates for each condition. | 94 |
| 6 | Selective accumulation of <u>Prochromadorella neapolitana</u> to control wells. | 95 |
| 7 | Selective accumulation of <u>Monhystera refringens</u> to alternating wells of diatom species D1-D4 (see Table 1). Diagram records four replicates for each condition. | 96 |
| 8 | Selective accumulation of <u>Monhystera refringens</u> to control wells. | 97 |
| 9 | Selective accumulation of <u>Monhystera refringens</u> to alternating wells of diatom species D5-D8 (see Table 1). Diagram records four replicates for each condition. | 98 |
| 10 | Selective accumulation of <u>Monhystera refringens</u> to control wells. | 99 |
| 11 | Selective accumulation of <u>Monhystera refringens</u> to alternating wells of bacteria species B1-B4 (see Table 1). Diagram records four replicates for each condition. | 100 |
| 12 | Selective accumulation of <u>Monhystera refringens</u> to control wells. | 101 |

| Appendix | | Page |
|----------|---|------|
| 13 | Selective accumulation of <u>Monhystera disjuncta</u> to alternating wells of diatom species D1-D4 (see Table 1). Diagram records four replicates for each condition. | 102 |
| 14 | Selective accumulation of <u>Monhystera disjuncta</u> to control wells. | 103 |
| 15 | Selective accumulation of <u>Monhystera disjuncta</u> to alternating wells of diatom species D5-D8 (see Table 1). Diagram records four replicates for each condition. | 104 |
| 16 | Selective accumulation of <u>Monhystera disjuncta</u> to control wells. | 105 |
| 17 | Selective accumulation of <u>Monhystera disjuncta</u> to alternating wells of bacteria species B1-B4 (see Table 1). Diagram records four replicates for each condition. | 106 |
| 18 | Selective accumulation of <u>Monhystera disjuncta</u> to control wells. | 107 |
| 19 | Selective accumulation of <u>Monhystera disjuncta</u> to alternating wells of (i) rod or (ii) coccoid bacteria species (see Table 1). Diagram records four replicates for each condition. | 108 |
| 20 | Selective accumulation of <u>Monhystera disjuncta</u> to control wells. | 109 |
| 21 | Two-level ANOVA calculated on age structure (males, females and juveniles/0.5 m ²) densities of <u>Prochromadorella neapolitana</u> . | 110 |
| 22 | Two-level ANOVA calculated on age structure (males, females and juveniles/0.5 m ²) densities of <u>Monhystera refringens</u> . | 111 |
| 23 | Two-level ANOVA calculated on age structure (males, females and juveniles/0.5 m ²) densities of <u>Monhystera disjuncta</u> . | 112 |

INTRODUCTION

Free-living nematodes are the most abundant component of the meiofauna in most coastal and oceanic sediments. They occur from above the highwater mark into the deepest oceanic trenches. Abundance and distribution studies of nematodes have been primarily in the intertidal and subtidal interstitial environments (Nicholas, 1975). Observers on sandy beaches in East Cornwall (U.K.) found that the nematode population constituted 50% of the total number of meiofauna inhabitants (Harris, 1972a). In a subsequent study, Harris (1972b) observed seasonal variations in the relative proportion of nematodes in the meiofaunal population ranging from 35.8% to 67.5% of the total number. A study of core samples from New York City Beach (U.S.A.) indicated that marine nematodes numerically made up 85% of the total interstitial fauna of the area (Martinez, 1975). In the subtidal meiofauna of two New England (U.S.A.) estuaries, nematodes were the dominant group, averaging 83% of the total numbers and 64% of the total biomass (Tietjen, 1969). Further observations by Tietjen (1969) indicated that the nematode population had marked seasonal changes in species composition and this was possibly associated with feeding behaviour. Epigrowth feeders reached maximum population density in the spring and summer (a time of high microflora production), whereas the deposit feeding and omnivorous species had maximum population densities in the fall and winter when detritus was most

abundant.

The bottom sediments are not the sole marine environment inhabited by free-living nematodes. One habitat that has been insufficiently studied, in spite of nematode abundance, is the microhabitat immediately surrounding marine macroalgae. The most notable contributions to this area have been those of Wieser who did an extensive taxonomic survey of marine nematodes associated with seaweeds in Chile (1953, 1959a) and in Plymouth (U.K.) (1952). In the Plymouth study, eight species of intertidal algae were sampled and he found that nematode species distribution was dependent on the algal species and the silt content of the area. Colman (1940) found nematodes to be widely distributed in all samples of the brown algae Ascophyllum and Laminaria and the red alga, Gigartina. Relative numbers as high as 247.8 individuals for every 10.0 g wet weight of seaweed were recorded. A study of Fucus serratus Linnaeus, in Sweden showed relatively low numbers of associated nematodes and the highest concentration occurred in June (6.05 individuals/g wet weight of algae) (Hagerman, 1966).

More recent studies of nematode-algae associations have considered the abundance and distribution of particular feeding types, and the parameters affecting this ecological pattern. Nematode populations on the brown alga, Sargassum, in the Adriatic Sea, are greatly affected by the thickness of the epigrowth on the algae (Ott, 1967). Epigrowth feeders are predominant on the thalli of these plants.

Studies on the nematode fauna associated with turtle grass beds, Thalassia testudinum Konig, in Florida, with Laminaria hyperborea Gunn, holdfasts off Britain, and Sargassum species off Japan have shown similar results (Hopper and Meyers, 1967a, 1967b; Moore, 1971; Mukai, 1971). Nematode distribution and abundance on Macrocystis pyrifera (Linnaeus) C. Ag., in California was shown to be closely related to blade age and position (Wing and Clendenning, 1971). The degree of shelter and the accumulation of detritus afforded by seaweeds permits particular patterns of nematode abundance and feeding strategies (Moore, 1971; Wieser, 1952b).

Investigations of trophic relationships in the marine environment have shown that meiofauna food webs encompass a wide variety of complexities and interactions. Each major taxonomic group has been split into distinct and different feeding types, with each species feeding on as varied a diet as can be found in their environment. Recent comprehensive studies have shown a great diversity of feeding within these taxa. Marine nematodes are without exception to these generalized observations.

A large number of marine ecology studies include reference to marine nematodes which are notoriously difficult to identify. This resulted in the classification of nematode species based on their respective feeding habits. Although it is incomplete, it provides a basis for analysis of their food preferences. The four categories, as

postulated by Wieser (1952a), are based on a correlation of stomal morphology with observations on feeding habits.

Wieser's four groups are:

- 1A. Selective deposit feeders: with or without very reduced stomal cavity. Food, which must be soft and in suspension, is ingested by oesophageal suction. Large and hard particles are not ingested.
- 1B. Unselective deposit feeders: stoma with unarmored cup-shaped or cylindrical cavity. Oesophageal suction supplemented by the movements of the lips and stoma in ingesting food. Food, in suspension, includes relatively large hard objects, such as diatoms, as well as finer material.
- 2A. Epigrowth feeders: stoma with teeth, rods or plates. Food may be scraped from surfaces for ingestion, or cells may be pierced and the contents sucked out.
- 2B. Predators and omnivores: stoma with powerful armature of teeth and plates. Prey may be swallowed whole, or small animals or algal cells may be pierced and the contents sucked out (Nicholas, 1975).

A study on the nematode fauna associated with kelp hold-fasts showed 52% of the species belonging to the omnivore category, 24% epigrowth feeders, 18% selective deposit feeders and 6% unselective deposit feeders (Moore, 1971).

These results indicated that the most important sources of food were the epibiota growing on the algae and that the sparse occurrence of the deposit feeders reflected the scarcity of suitable deposits within the holdfast habitat.

Detritus is an important factor limiting the distribution and abundance of nematode faunas (Tietjen, 1966). Teal and Wieser (1966), investigating the ecology of nematodes in a Georgia salt March (U.S.A.), observed that areas subject to the highest organic detritus levels constituted the most productive habitats for nematode abundance. From these studies, Tenore et al. (1977) have suggested that it is not the detritus which supplies the energy but rather the associated microbiota particularly bacteria. Bacteria have been suggested as the source of much of the deposit-feeders nourishment (Zobell and Feltham 1937, 1942). These bacteria for the most part, along with blue-green algae, diatoms, yeasts, fungi and microscopic stages of some brown algae are present on the surfaces of detrital and granular surfaces (McIntyre and Munson, 1973; Meadows and Anderson, 1966). Investigation on the distribution of these microscopic populations have suggested that local and geographical differences are likely paralleled by the productivity of the environment and these observed differences may account for the localized distribution of various nematode species.

Observations from an experimental sand ecosystem showed that a marine nematode population varied directly with

the relative bacterial numbers and that the vertical zonations of the bacteria and nematode were strikingly similar (Boucher and Chamroux, 1976).

Nematodes have a variety of nutritional associations with certain species of filamentous fungi (Meyers, 1963). An Aphelechooides sp. develops and reproduces on viable mycelia of Dendryphiella arenaria. Studies by Meyers and Hopper (1966, 1967), with fungal substrates, found a dominant nematode fauna (Metonchalaimus sp.) colonizing the fungal mats. Whereas a number of marine nematode investigations have probed trophic relationships in terms of species abundance and levels of attraction; a number of studies have tried to characterize nematode biotic associations by way of feeding experiments and gut contents. Jennings and Colman (1970) found large amounts of mud and silt in the guts of a marine nematode, Pontonema vulgaris Bastian, suggesting the digestion of bacteria and any other organic materials present in the substrate. The marine Monhystera species have been shown to have detritus throughout the gut and feeding experiments successfully maintained Monhystera species on the bacteria: Arthrobacter, sp., Pseudomonas sp., Vibria sp. and Flavobacterium sp. (Chitwood and Murphy, 1964). Observations on the marine nematode, Chromadora macrolaimoides Steiner, a species equipped with a long stoma armed with three movable teeth used to scrape food from substrate, indicated it to be very selective in its ingestion and digestion of algae. Twenty

species of algae, mostly diatom species, were tested, with the result that only eleven were effectively utilized and of the eleven, only five could sustain growth to ten generations (Tietjen and Lee, 1973). A related study by Tietjen and Lee (1977) comparing feeding habits of two epigrowth feeders and two non-selective feeders suggested that not only was selective ingestion a significant factor in nematode feeding habits, but so also was selective digestion.

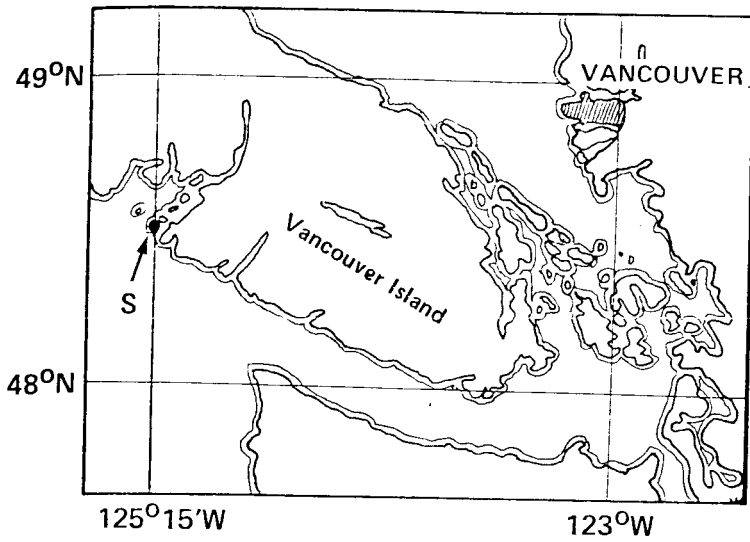
The objectives of this study were to characterize the abundance, seasonality and distribution of the predominant marine nematode species inhabiting M. integrifolia beds. Then, using diatoms and bacteria isolated from M. integrifolia blades, determine food preference and relate these findings to their seasonality and distribution on M. integrifolia blades.

DESCRIPTION OF STUDY SITE AND ASSOCIATED KELP

This study of marine nematodes associated with the large brown kelp, Macrocystis integrifolia Bory, was located in Dodger Channel (48 50' 04" N, 125 11' 55" W) in the Barkley Sound region of Vancouver Island, British Columbia, Canada (Fig. 1). The kelp bed extended 16.0 m out from the shoreline and had a depth range from -1.2 to -5.5 m below mean zero tide. The study site's substrate was characterized by sand, rock rubble and small rock outcroppings. In summer, the kelp bed was fairly sheltered from wave action but during the winter was semi-wave exposed due to the direction of the prevailing winds.

Macrocystis integrifolia, (Fig. 2), is the largest member of the Phaeophyceae found on the British Columbia coast (Scagel, 1971). It is characterized by having a large flattened rhizome-like holdfast from which arise many dichotomously branched haptera which are used to secure the plant to the substrate. From this holdfast extend erect stipes ranging up to 0.5 cm in diameter and 10.0 m in height. Large blades arise at intervals along the stipe. The blades may range from 15.0 cm to 1.5 m in length and 5.0 cm to 25.0 cm in width. The blades are flattened, deeply furrowed, denticulate along the edges and have at their point of attachment a pneumatocyst. The main intercalary meristem is located near the top of a stipe, with a secondary meristematic region located at the distal end of the

Fig. 1: Location of Dodger Channel study site in the Barkley Sound region of Vancouver Island, B.C.



S - Bamfield Study Site

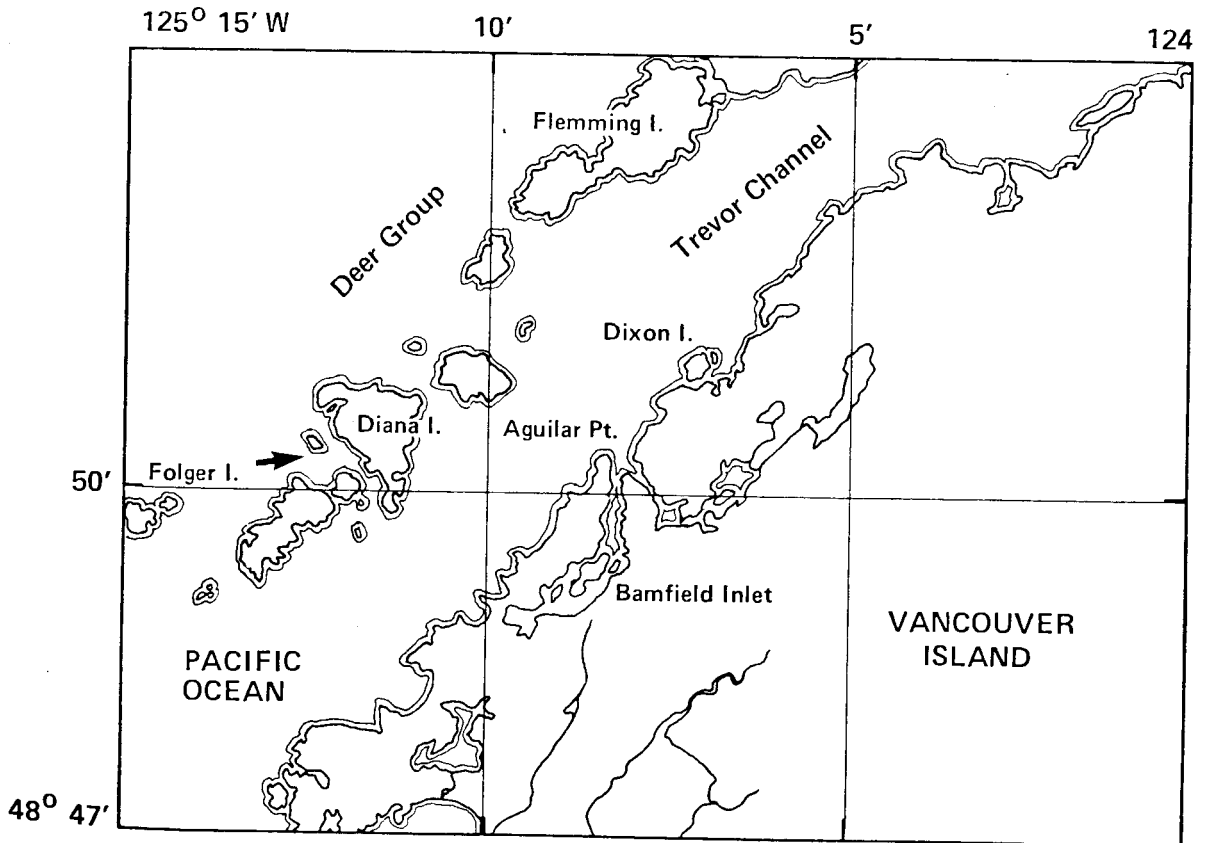
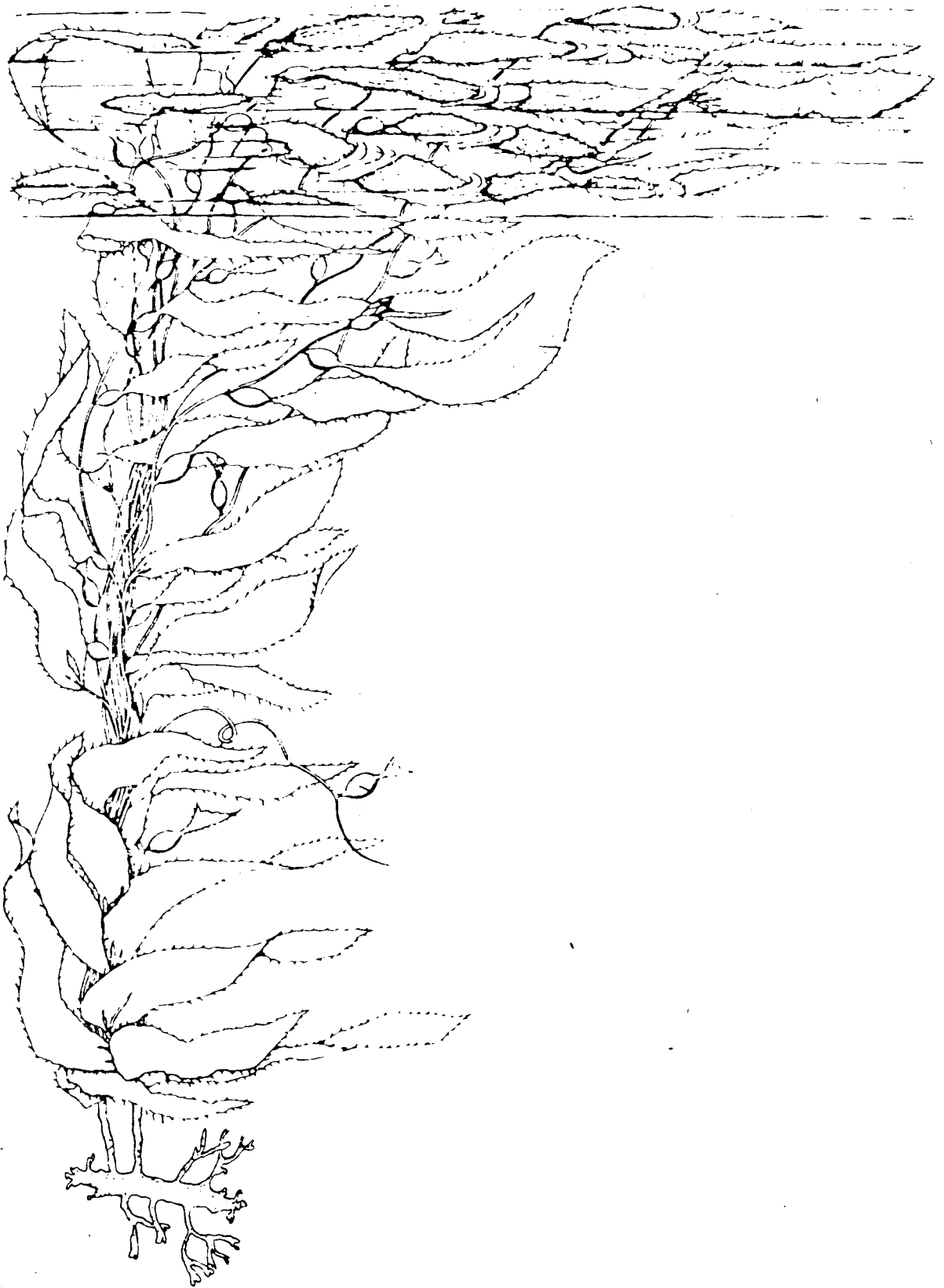


Fig. 2: Semi-diagrammatic sketch of Macrocystis sp.



pneumatocyst.

The holdfast of M. integrifolia is perennial and new stipe and blades are produced throughout the year with production rates and distribution varying seasonally (Druehl, 1978; Lobban, 1978). With respect to blade age, three stages are recognized; growing tissue, mature tissue and senescent tissue (Lobban, 1978).

MATERIALS AND METHODS

I. Field Survey

a) Sampling Procedure

To ascertain the distribution of free-living marine nematodes on M. integrifolia, a single, weighted transect line was randomly established in a kelp bed in Dodger Channel. The line was secured approximately 2 m inshore from the edge of the kelp bed and extended through the kelp, perpendicular to the shore line and secured 33 m out from the inshore point by cement blocks. Prior to its placement in the kelp bed, the line was marked with coded surveyor's tape at 2 m intervals. Blades of M. integrifolia were collected monthly for one year from July 1978 to July 1979 and subsequently bi-monthly until November 1979, using SCUBA equipment. No data was collected for December 1978, August and October 1979. The closest plant within 1.0 m radius of each marker on the transect line was selected for sampling. At each sampling, three blades were removed; one from a height of 1 m from the plant holdfast, one at 1 m from the distal end of the plant and the third at a point between the first two sampled blades. Specific care was taken to sample only vegetative blades and not sporophylls. Twelve plants at 2 m intervals were sampled on each sampling date.

The blades were removed by carefully slipping a 38.5 x 13.0 x 7.5 cm plastic bag over the sample blade which was then detached from the stipe and the bag was immediately closed and secured with a twist tie. Care was taken to

disturb the blade as little as possible. Each sampling bag was coded and registered on an underwater slate. The coded bags were numbered as to date, transect line marker and depth. This procedure was repeated at each of the other eleven sample points and then the samples were taken to the surface and transported in buckets of seawater to the Bamfield Marine Station laboratory. Time for transport never exceeded 45 minutes.

b) Treatment of Samples

On arrival at the Bamfield Marine Station, the contents of each bag were fixed with 250 ml of a solution of 10% formalin in filtered seawater and left at room temperature for 24-36 h. Each sample was processed as follows: the bag was agitated so as to thoroughly mix the debris and detritus in the solution and on the blade surface. The suspension was carefully poured through a 44 μ brass sieve. The material remaining in the sieve was then washed into a 100 x 15 mm Petri dish. The blade was then carefully removed from the bag and placed into a small bath of 10% formalin in filtered seawater. The plastic bag was washed out with filtered seawater and the suspension filtered as described above. The blade was agitated for approximately one minute and then whilst being suspended by the pneumato-cyst, washed down twice with a 10% formalin solution from a plastic squeeze bottle. The bath solution was filtered twice through a 44 μ sieve and the collected residue placed in a 100 x 15 mm Petri dish. Every third blade was examined

under the dissecting microscope (120x) to evaluate the efficiency of this processing procedure for removing nematodes. The cleaned blade was photographed to scale and discarded. These photographs were used subsequently to determine the total surface area of the blade (both sides) using a compensating polar planimeter.

The Petri dish samples were examined under a dissecting microscope and all nematodes were transferred by hand to 1.5 cm diam. watchglasses where they were processed as outlined by Sharma (1978). The nematodes were individually mounted in glycerine on permanent slides for identification and sexing and for enumeration with respect to blade sample.

c) Statistical Treatment

The collected data were analyzed using a two-level and three-level analysis of variance using programs developed by Davies (1971).

d) Environmental Parameters

Measurements of temperature and salinity for each of the sampling months were obtained from data collected by Dr. L. Druehl of Simon Fraser University and the Bamfield Marine Station.

e) Taxonomic Notes

All nematodes found in the samples were identified at least to genus and the most abundant individuals were classified to species with the help of the major taxonomic keys and checklists of Chitwood (1960), Gerlach and Rieman

(1973) and Wieser (1953, 1959b). Those species identified to species had their identity confirmed by Mr. Bruce Hopper, Agriculture Canada, Ottawa.

II. Feeding Preference Experiments

a) Culture of Nematodes

Living nematodes, collected at random from the middle and bottom samples of M. integrifolia blades from January to July 1980, were extracted by the sieving process described above and then transferred to 105 cm² Corning Tissue Culture Flasks, containing 400 ml of filtered seawater and several small pieces of blade of M. integrifolia. The flasks were incubated at 7 C with a 12/12 hour day (250 μ W/cm²)/night cycle. Within 24 h, the culture flasks were transported in an ice chest from Bamfield to the Simon Fraser University laboratory, where they were incubated at 10 C with a 16/8 hour day (250 μ W/cm²)/night cycle.

The nematodes were maintained in such an agnotobiotic culture in which every day for the first week 300 ml of the culture seawater was replaced by an equal amount of autoclaved, filtered seawater. Thereafter, this procedure was performed only once per week. The larger competitive and/or predatory invertebrates were removed from the cultures.

The three most abundant nematode species were isolated by hand from these cultures and placed into separate culture flasks containing only detritus and pieces of M. integrifolia in autoclaved filtered seawater. Once each week, 5 ml of monoxenic diatom and bacterial cultures isolated from M. integrifolia blade samples were added to the flasks to supplement the nematodes diet. When the next generation of nematode larvae appeared, additional nematode cultures were initiated,

using the techniques described above. Hence, a large population of each of the three species was maintained in culture and available for the food preference experiments. Observations on the life cycles of these nematode species were made during the development of these cultures.

One half of these nematode cultures were maintained under simulated winter conditions at 5 C with an 8/16 hour (250 μ W/cm²) /night cycle and the remaining half were maintained under simulated summer conditions (10 C, 16/8 h day/night). It was possible, subsequently to make comparisons with respect to feeding preferences under two environmental conditions. Under both conditions, salinity was maintained at 31‰.

b) Culture of Food Sources

Bacteri and diatom species used in the food preference experiments were isolated from the detritus and from blade scrapings. Bacterial species were isolated in standard Petri dishes containing Difco Marine Broth and 1.0% Difco Bacto Agar. The cultures were maintained at room temperature on a 12/12 hour day/night light cycle. Four bacterial species were isolated from these blades by standard microbiological techniques and then maintained on liquid and solid media at 10 C on a 16/8 hour day/night cycle (Table 1).

Eight marine distom species were isolated by hand from blade scrapings using elongated Pasteur Pipettes (Table 1). These species were incubated in sterile "S" medium (Lee et al. 1970) at 22 C on a 16/8 hour day/night cycle. Every

Table 1: Diatom (D1-D8) and bacteria (B1-B4) species isolated from Macrocystis integrifolia blades and used in the feeding preference experiments.

| <u>Experiment I</u> | | | | <u>Size (µm)</u> | |
|---------------------|---|--|--|------------------|--------------|
| | | | | <u>Length</u> | <u>Width</u> |
| D1 | - | * <u>Gramatophora marina</u> (Lyngb.) Kutz | | 35 - 40 | 6 - 7 |
| D2 | - | * <u>Rhoicosphenia curvata</u> (Kutz) Grunow | | 35 - 40 | 8 - 10 |
| D3 | - | <u>Navicula jamalinensis</u> Cleve | | 25 - 50 | 15 - 25 |
| D4 | - | * <u>Synedra tabulata</u> (Ag.) Kutz | | 25 - 140 | 3 - 6 |

Experiment II

| | | | | | |
|----|---|---|--|---------|---------|
| D5 | - | <u>Gomphanema olivaceum</u> (Lyngb.) Kutz | | 35 - 40 | 5 - 7 |
| D6 | - | * <u>Cocconeis scutellum</u> Ehr. | | 25 - 50 | 10 - 40 |
| D7 | - | * <u>Achnanthes longissipes</u> Ag. | | 15 - 25 | 6 - 9 |
| D8 | - | <u>Amphora acutiuscula</u> Kutz | | 25 | 10 |

Experiment III, IV and V

| | | | |
|----|---|------------------------|---------------------------|
| B1 | - | rod, Gram negative | round, white colony |
| B2 | - | coccoid, Gram positive | round, yellow colony |
| B3 | - | rod, Gram neagtive | round, orange colony |
| B4 | - | coccoid, Gram positive | round, dark yellow colony |

* species associated with M. integrifolia from Roland (1980).

week these cultures of diatom species were checked for the presence of bacteria and marine fungi in the culture tubes by subculturing on Petri dishes containing Difco Marine Broth. Cultures with high populations of bacteria and/or marine fungi (> 5 colonies) were re-isolated and new cultures were initiated. Under this checking system, diatom cultures containing low bacterial and/or fungal populations were maintained.

c) Experimental Design

Experiments were designed to demonstrate the feeding preferences of the three most abundant nematode species found on M. integrifolia. The "cafeteria" design as described by Lee et al. (1977) was employed with the following modifications. Twenty-four hours prior to the feeding experiment, adult male and female nematodes of each species were isolated by hand from the cultures. They were washed in autoclaved, filtered seawater containing 100 units of the bactericidal Polymyxin B, for 6 hours. The individual species were then separated by sex and placed in culture flasks containing only autoclaved filtered seawater for 18 hours.

The nematodes were transferred one at a time into wells, made by a sterilized #13 cork borer (2 cm diam), in a Petri plate (150 x 25 mm) containing 40 ml of 0.4% seawater agar. The well pattern was such that a central and eight equidistant peripheral wells were located on the plate. Subsequently, 0.7 ml of bacteria or diatom

suspension were placed in each of four alternate peripheral wells and the remaining peripheral wells were filled with 0.7 ml of sterile culture media. Large coverslips (29 x 50 mm) were then placed over each well. Control plates had eight wells filled with sterile culture media.

Diatom populations of $1.0 \times 10^5 \pm 1.0 \times 10^3$ individuals/ml were inoculated into the designated food wells. Bacterial food wells were inoculated with 1.0×10^6 /ml \pm 1.0×10^4 /ml (Hobbie et al., 1977).

One hundred adult nematodes (1:1 sex ratio) of one of the three species were placed in seawater in the central well and covered with a coverslip. The agar plate was then carefully covered with a 0.5 cm deep layer of sterile seawater after which all the coverslips were carefully removed. Four replicate plates and one control plate for each of the diatom or bacteria feeding preference experiments were established for each nematode species. Two additional experiments were done using Monhystera disjuncta Bastian under simulated summer conditions. They involved exposing this nematode species to either all rod or coccoid bacteria species under the same "cafeteria" design.

Each of the four experimental plates were oriented at right angles to the plate to its left or right and the control plate was placed in the center of the group of four experimental plates. The experiments were conducted under simulated summer (10 C, 16/8 h day/night cycle) and winter (5 C, 8/16 h day/night cycle) conditions. The

nematode inoculated plates were left under these conditions for 72 hours and then the number of nematodes present in each well was counted and sexed.

Detailed observations on the life cycle were collected only for M. disjuncta.

d) Statistical Treatment

The data from the nematode feeding preference experiments were analyzed by the X^2 test, two-level ANOVA and the Student-Newman-Keuls test (Sokal and Rohlf, 1969).

RESULTS I

a) Observations on Physical Parameters

Surface water temperature (Fig. 3) ranged from 6.1 C to 14 C and surface water salinity from 29.4‰ to 32.0‰. The lowest temperature occurred in January and the highest in July. The highest salinity occurred during the summer months.

b) Nematode Species Composition

Nine nematode species belonging to six families were found associated with M. integrifolia (see Table 2). They were identified and classified according to the system of Chitwood and Chitwood (1950) as modified by Gerlach and Rieman (1973). Three of these species constituted 91.0 - 99.1% of the total nematode population in the M. integrifolia samples. The most abundant species were Monhystera disjuncta, M. refringens and Prochromadorella neapolitana.

c) Seasonal Species Changes

i) Prochromadorella neapolitana

Most P. neapolitana were found in early and mid-summer (Fig. 4). The largest percent abundance levels occurred in July 1978 and 1979 (40.3% and 37.6% respectively). Low numbers of this species were recorded for the remainder of the year except for November 1978 and 1979, when there was an increased frequency to 24.3% and 22.4% respectively. The age structure for P. neapolitana, represented as the percentage of males, females and juveniles, is given in Fig. 4

Fig. 3: Temperature and salinity in Dodger Channel over the sampling period 1978 - 1979.

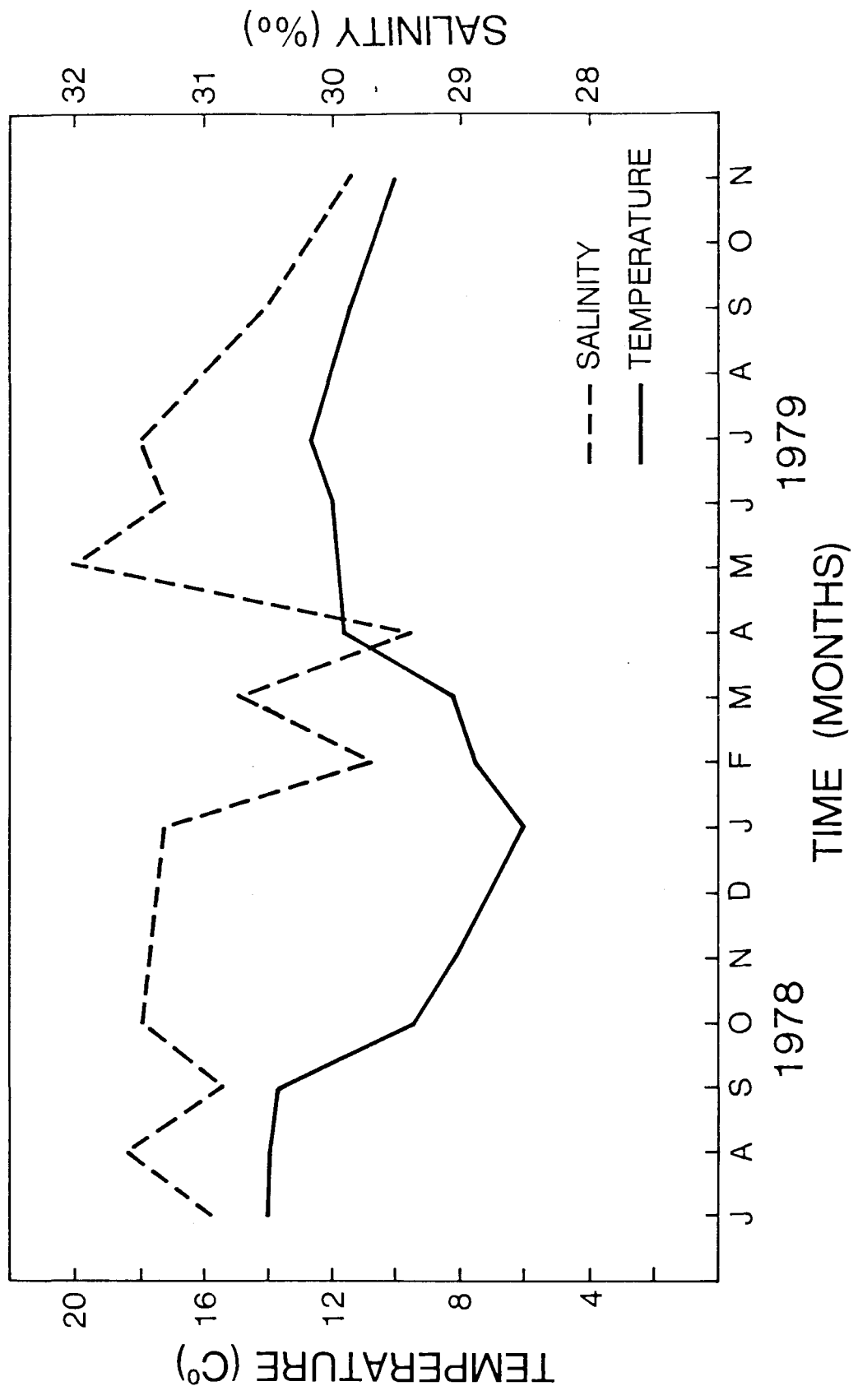


Table 2: Checklist of free-living nematodes found in blade samples of Macrocystis integrifolia.

Class Adenophorea

Order Araeolaimida

Family Axonlaimidae

Subfamily Diplopeltinae

Araeolaimus

A. elegans, DeMan, 1888

Order Monhysterida

Family Monhysteridae

Subfamily Monhysterinae

Monhystera

M. disjuncta, Bastian, 1865

M. refringens, Bresslau & Stekhoven, 1935

Order Chromadorida

Family Chromadoridae

Subfamily Chromadorinae

Chromadora

C. nudicapitata, Bastian, 1865

Chromadorina

C. laeta, Micoletzky, 1924

Prochromadorella

P. neapolitana, Micoletzky, 1924

Family Cyatholaimidae

Subfamily Paracanthonchinae

Parocanthonchus, Micoletzky, 1924

Paracanthonchus sp.

Order Enoplida

Family Anticomidae

Anticoma

A. acuminata, Bastian, 1865

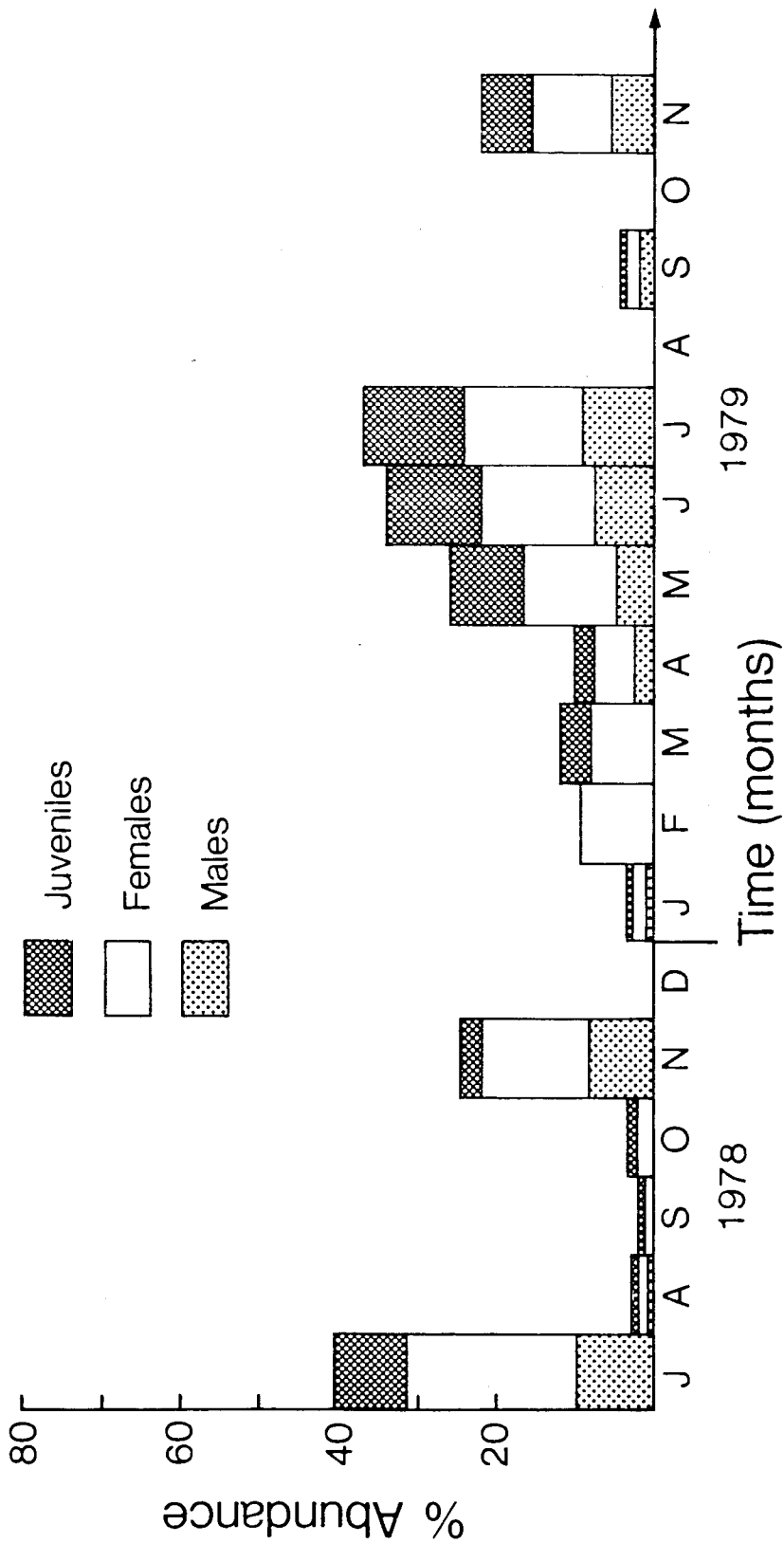
Family Oncholaimidae

Subfamily Oncholaiminae

Oncholaimus

O. dujardini, DeMan, 1876

Fig. 4: Per cent distribution and age structure of
Prochromadorella neapolitana over the sampling
period 1978 - 1979.



and Table 3. Their densities were found to be significant across the sampling period (see Appendix 21). Males were found in low frequencies except in June, July and November. In the months of September 1978, February and March 1979, none were found in the samples. Females were found in every month sampled and were very frequent except in September 1978, 1979, October 1978 and January 1979. There were low frequencies of juveniles in all the sampled months except those in the summer. No juveniles were recorded from the February 1979 samples.

ii) Monhystera refringens

Monhystera refringens occurred throughout the sampling years of 1978 and 1979 (Fig. 5). It had the highest frequency of occurrence during the early fall. The lowest frequency of occurrence was mid-winter and, thereafter, there was a gradual increase in population from January (11.0%) to September 1979 (44.1%). Densities of males, females and juveniles were found to be significant in all the sampled months (see Appendix 22). The age structure of this species, expressed as the percentage of males, females and juveniles, is given in figure 5 and Table 4. Males were less frequent in the population samples than either females or juveniles and exhibited little intersample variation with the exception of slight increases in the fall months. In particular, juveniles occurred most frequently during the early fall and contributed greatly to the increased overall fall frequencies for this species.

Table 3: Total number/0.5 m² and per cent abundance of males, females and juveniles of Prochromadorella neapolitana over sampling period 1978 - 1979.

| Month | Total No. | Males | | Females | | Juveniles | |
|----------|-----------|-------|-------|---------|-------|-----------|-------|
| | | # | % | # | % | # | % |
| July '78 | 108 | 26 | 24.1 | 59 | 54.6 | 23 | 21.3 |
| Aug. | 11 | 3 | 27.3 | 5 | 45.4 | 3 | 27.3 |
| Sept. | 6 | 0 | 0.0 | 4 | 66.7 | 2 | 33.3 |
| Oct. | 11 | 1 | 9.1 | 6 | 54.5 | 4 | 36.4 |
| Nov. | 69 | 23 | 33.3 | 38 | 55.1 | 8 | 11.6 |
| Dec. | -- | -- | -- | -- | -- | -- | -- |
| Jan. '79 | 11 | 2 | 18.2 | 7 | 63.6 | 2 | 18.2 |
| Feb. | 22 | 0 | 0.0 | 22 | 100.0 | 0 | 0.0 |
| March | 30 | 0 | 0.0 | 20 | 66.7 | 10 | 33.3 |
| April | 30 | 8 | 26.7 | 15 | 50.0 | 7 | 23.2 |
| May | 98 | 18 | 18.4 | 45 | 45.9 | 35 | 35.7 |
| June | 161 | 35 | 21.7 | 69 | 42.9 | 57 | 35.4 |
| July | 152 | 37 | 24.4 | 61 | 40.1 | 54 | 35.5 |
| Aug. | -- | -- | -- | -- | -- | -- | -- |
| Sept. | 13 | 6 | 46.1 | 4 | 30.8 | 3 | 23.1 |
| Oct. | -- | -- | -- | -- | -- | -- | -- |
| Nov. | 62 | 15 | 24.2 | 29 | 46.8 | 18 | 29.0 |
| Mean | 55.9 | 12.4 | 19.5 | 27.4 | 54.5 | 16.1 | 26.0 |
| S.D. | +54.0 | +13.3 | +13.4 | +23.2 | +16.6 | +19.3 | +10.7 |

Fig. 5: Per cent distribution and age structure of Monhystera refringens over the sampling period 1978 - 1979.

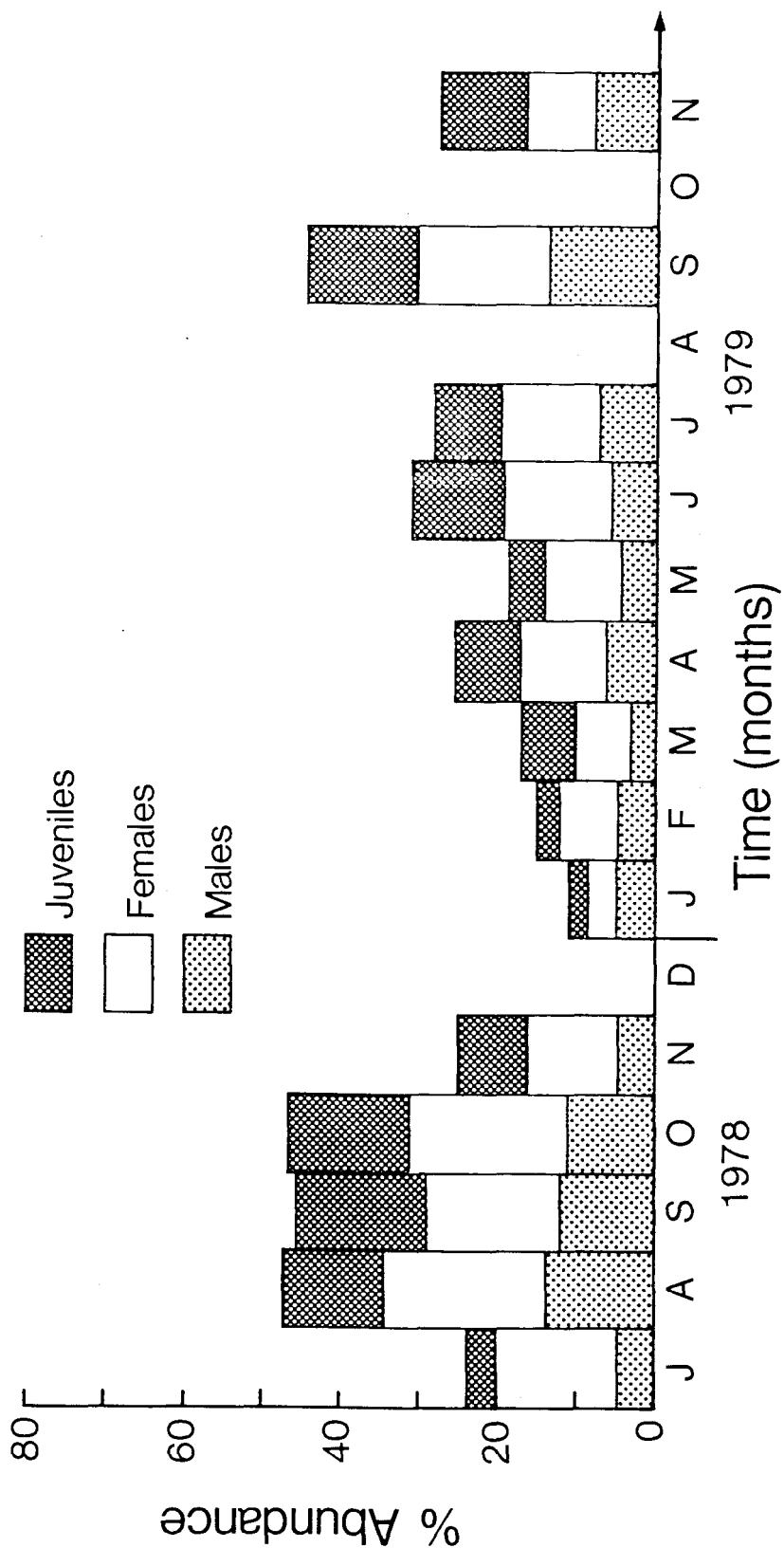


Table 4: Total number/0.5 m² and per cent abundance of males, females and juveniles of Monhystera refringens over sampling period 1978 - 1979.

| Month | Total No. | Males | | Females | | Juveniles | |
|----------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| | | # | % | # | % | # | % |
| July '78 | 64 | 12 | 18.8 | 43 | 67.2 | 9 | 14.0 |
| Aug. | 160 | 47 | 29.4 | 70 | 43.7 | 43 | 26.9 |
| Sept. | 137 | 36 | 26.3 | 51 | 37.2 | 50 | 36.5 |
| Oct. | 168 | 42 | 25.0 | 71 | 42.3 | 55 | 32.7 |
| Nov. | 69 | 13 | 18.9 | 31 | 44.9 | 25 | 36.2 |
| Dec. | -- | -- | -- | -- | -- | -- | -- |
| Jan. '79 | 34 | 15 | 44.1 | 11 | 32.4 | 8 | 23.5 |
| Feb. | 34 | 10 | 29.4 | 17 | 50.0 | 7 | 20.6 |
| March | 42 | 7 | 16.7 | 18 | 42.8 | 17 | 40.5 |
| April | 75 | 17 | 22.7 | 33 | 44.0 | 25 | 33.3 |
| May | 67 | 16 | 23.9 | 36 | 53.7 | 15 | 22.4 |
| June | 147 | 27 | 18.4 | 63 | 42.8 | 57 | 38.8 |
| July | 112 | 28 | 25.0 | 49 | 43.8 | 35 | 31.2 |
| Aug. | -- | -- | -- | -- | -- | -- | -- |
| Sept. | 134 | 40 | 29.8 | 51 | 38.1 | 43 | 32.1 |
| Oct. | -- | -- | -- | -- | -- | -- | -- |
| Nov. | 74 | 21 | 28.4 | 23 | 31.1 | 30 | 40.5 |
| Mean | 94.0 | 23.6 | 25.5 | 40.5 | 43.8 | 29.9 | 30.7 |
| S.D. | <u>+47.5</u> | <u>+13.1</u> | <u>+ 7.0</u> | <u>+19.6</u> | <u>+ 9.0</u> | <u>+17.6</u> | <u>+ 8.1</u> |

iii) Monhystera disjuncta

Seasonal variations in the relative abundance of males, females and juveniles of this species are shown in figure 6. M. disjuncta was present throughout the year but was most abundant in the winter months (January to March). The population of this species was lowest in the summer months, and in particular in June 1979 and July 1978, 1979 when it showed 30.0% and 28.7% relative abundance respectively. The relative abundance in successive fall periods for 1978 and 1979 were closely similar with a range of 44.0% to 50.0% of the total population. Densities of males, females, and juveniles exhibited no significant monthly variation (see Appendix 23). Examination of the age structure of M. disjuncta from July 1978 to November 1979 showed that females were the predominant component of the population (Fig. 6, Table 5). With the exception of the mid-summer months, (June and July), the male and female populations exhibited little relative variation between months. The percentage frequency of the juvenile component in the population was less, particularly in July 1978, 1979, in comparison to the fall.

iv) Other species

Seasonal variations in the relative abundance of all other species found on M. integrifolia are shown in Fig. 7. These species comprised no more than 9.0% of the total population in any one month sampled. Fluctuations in the age structure of the population were not usually determined

Fig. 6: Per cent distribution and age structure of Monhystera disjuncta over the sampling period 1978 - 1979.

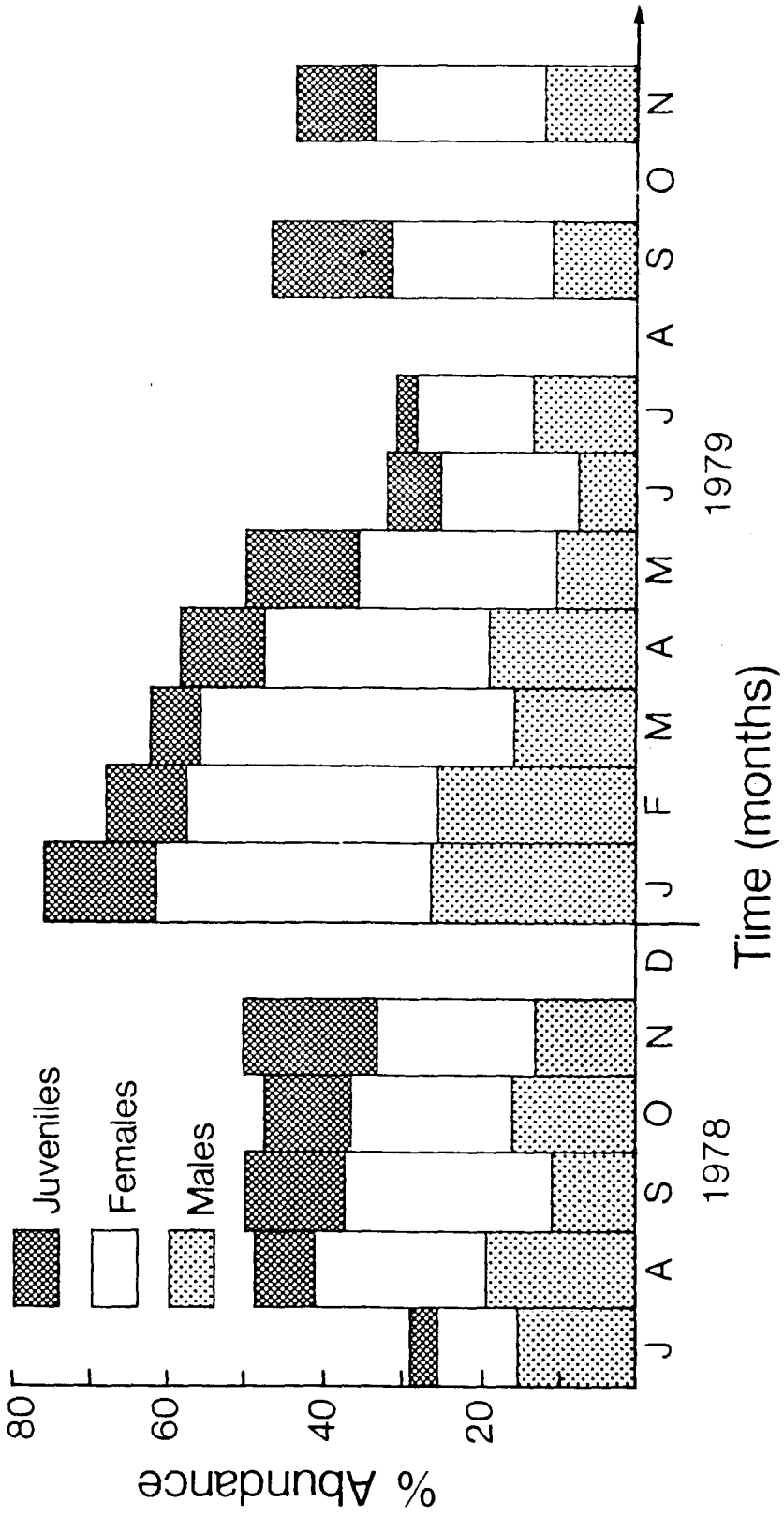
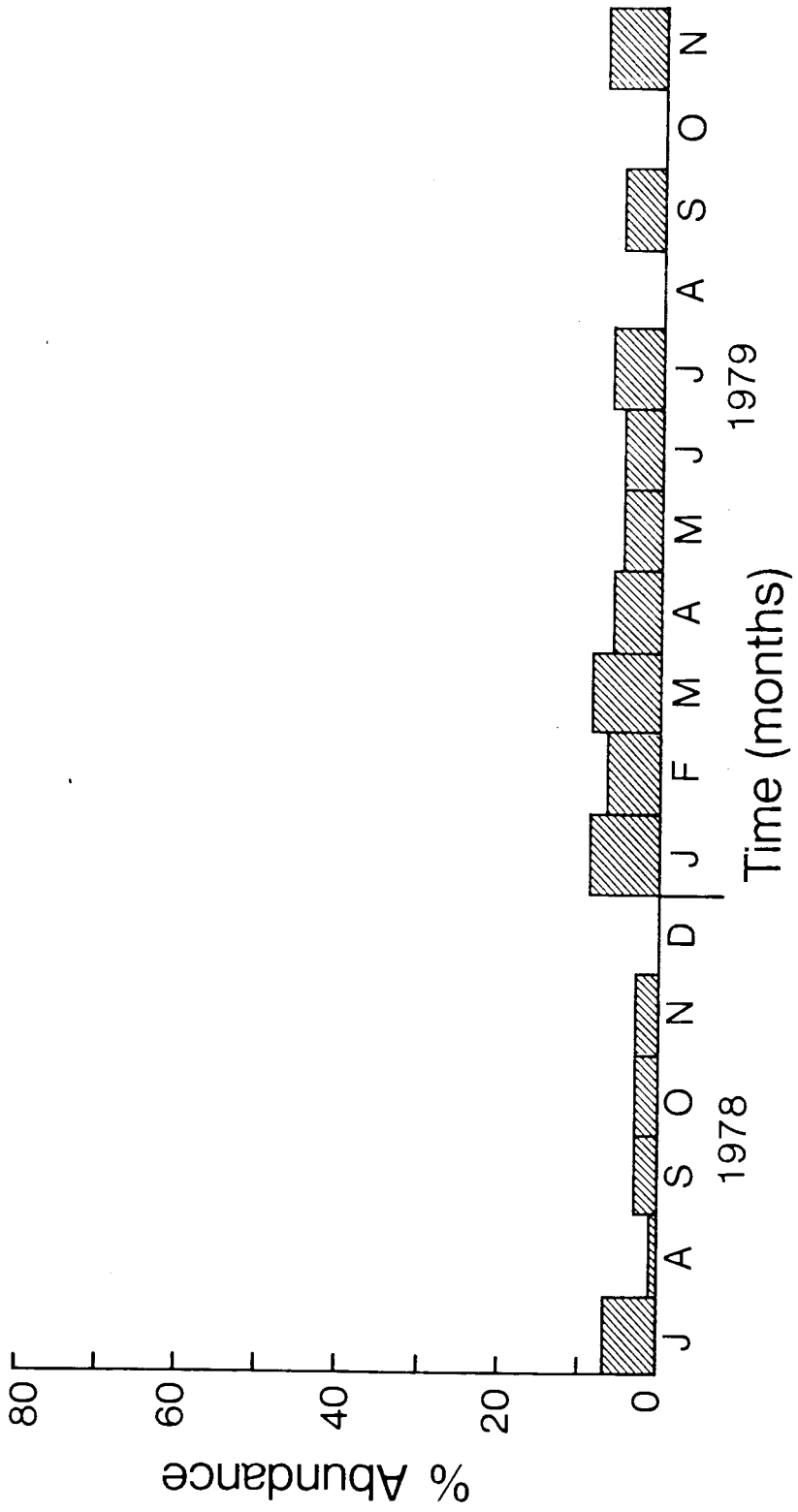


Table 5: Total number/0.5 m² and per cent abundance of males, females and juveniles of Monhystera disjuncta over sampling period.

| Month | Total No. | Males | | Females | | Juveniles | |
|----------|-----------|-------|------|---------|------|-----------|------|
| | | # | % | # | % | # | % |
| July '78 | 77 | 41 | 53.2 | 27 | 35.1 | 9 | 11.7 |
| Aug. | 163 | 66 | 40.5 | 72 | 44.2 | 25 | 15.3 |
| Sept. | 152 | 33 | 21.7 | 81 | 53.3 | 38 | 25.0 |
| Oct. | 171 | 58 | 33.9 | 74 | 43.3 | 39 | 22.8 |
| Nov. | 138 | 36 | 26.1 | 56 | 40.6 | 46 | 33.3 |
| Dec. | -- | -- | -- | -- | -- | -- | -- |
| Jan. '79 | 237 | 82 | 34.6 | 110 | 46.4 | 45 | 19.0 |
| Feb. | 155 | 59 | 38.1 | 73 | 47.1 | 23 | 14.8 |
| March | 155 | 40 | 25.8 | 100 | 64.5 | 15 | 9.7 |
| April | 174 | 58 | 33.3 | 86 | 49.4 | 30 | 17.3 |
| May | 189 | 39 | 20.6 | 97 | 51.3 | 53 | 28.1 |
| June | 142 | 33 | 23.3 | 79 | 55.6 | 30 | 21.1 |
| July | 116 | 50 | 43.1 | 56 | 48.3 | 10 | 8.6 |
| Aug. | -- | -- | -- | -- | -- | -- | -- |
| Sept. | 142 | 32 | 22.5 | 63 | 44.4 | 47 | 33.1 |
| Oct. | -- | -- | -- | -- | -- | -- | -- |
| Nov. | 122 | 33 | 27.0 | 60 | 49.2 | 29 | 23.8 |
| Mean | 152.4 | 47.1 | 31.7 | 73.9 | 48.1 | 31.4 | 20.2 |
| S.D. | +37.0 | +15.4 | +9.6 | +21.3 | +7.1 | +14.0 | +8.0 |

Fig. 7: Per cent distribution of the non-predominant nematode species over the sampling period 1978 - 1979.



because of the low total numbers of these species.

d) Species Distribution on Kelp

The distribution over the surface of M. integrifolia of the three most abundant species of marine nematodes is shown in figures 8-19 and of the remaining species in the population in figures 20-23. Most individuals of each of the three predominant species were located on the lower blades. Significantly fewer nematodes of each species occurred on the mid-blades and very few on the upper ones. Spot samples and ad hoc observations showed that the nematodes were distributed evenly along the length and only on the top surface of the blade.

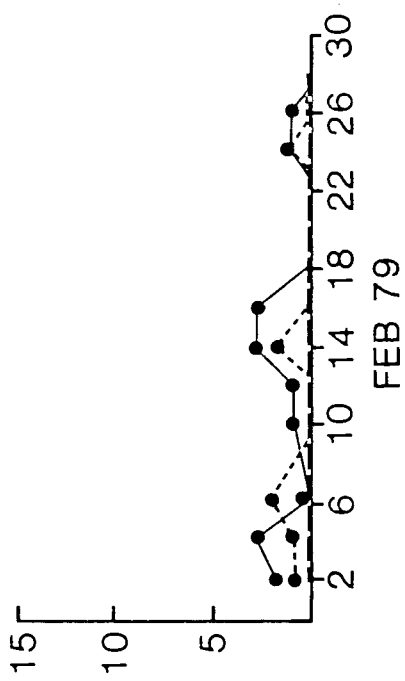
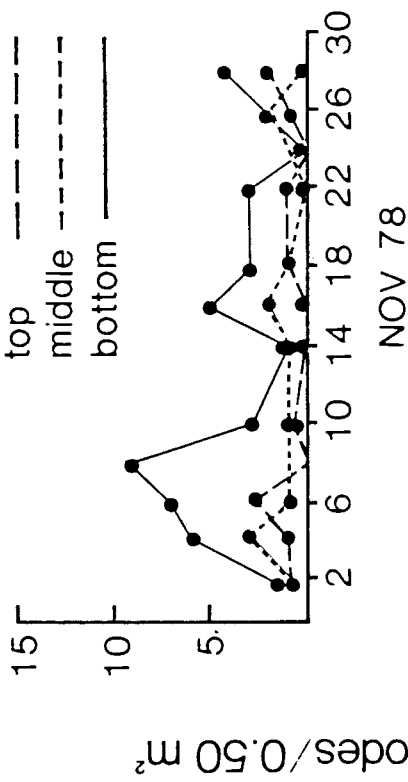
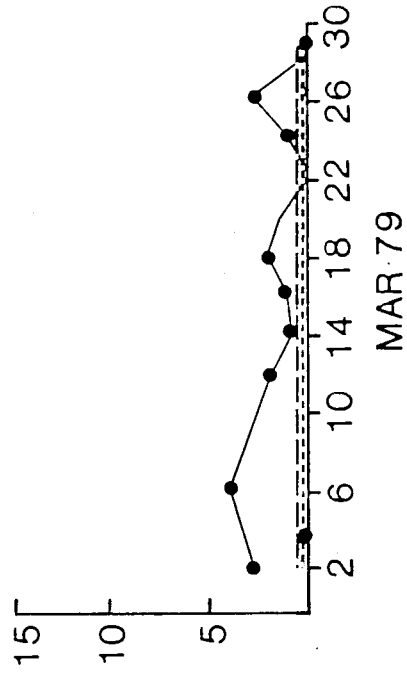
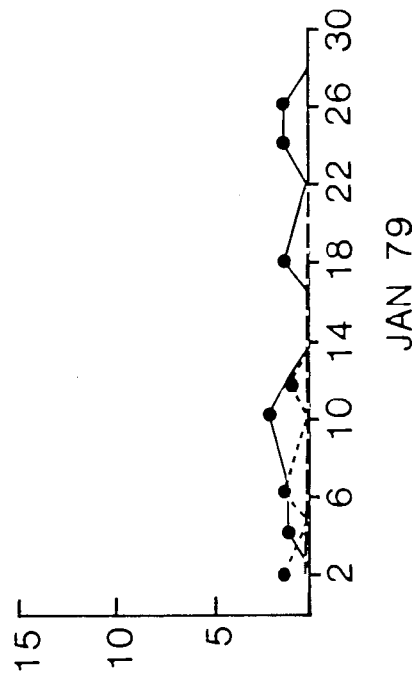
The number of individuals on M. integrifolia increased with depth of water and this was especially so with the number on the mid and lower blades. A two-way ANOVA was done of the data from each of the twelve grid points along the transect line arranged into three groups, each of four points. This showed that for the three predominant nematode species, blade position was a significant factor ($P < 0.005$) in their distribution (Tables 6-8). Further, for most of the sampling times, the distribution of most of the nematodes was related to blade position on the stipe ($P < 0.005$) and only rarely significantly to depth (Table 9).

These generalizations hold true for M. disjuncta with only July 1978, 1979 exhibiting a deviation from these above trends (Figs. 16 and 18).

M. refringens exhibited these general distribution patterns primarily in the late summer and early fall months.

Fig. 8: Density of Prochromadorella neapolitana (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
gradient from July 1978 to October 1978 (from deep
to shallow depth).

Fig. 9: Density of Prochromadorella neapolitana (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
from November 1978 to March 1979 (from deep to
shallow depth).



Distance (m)

No. of Nematodes/0.50 m²

Fig. 10: Density of Prochromadorella neapolitana (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
from April 1979 to July 1979 (from deep to shallow
depth).

Fig. 11: Density of Prochromadorella neapolitana (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
for September and November 1979 (from deep to
shallow depth).

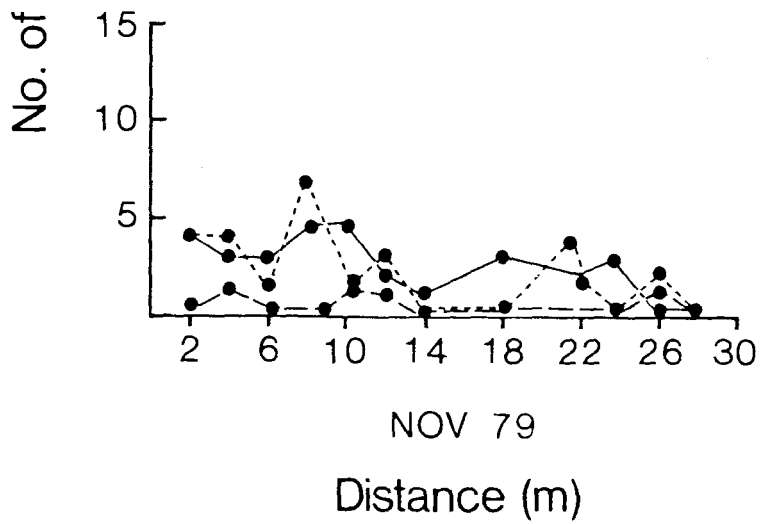
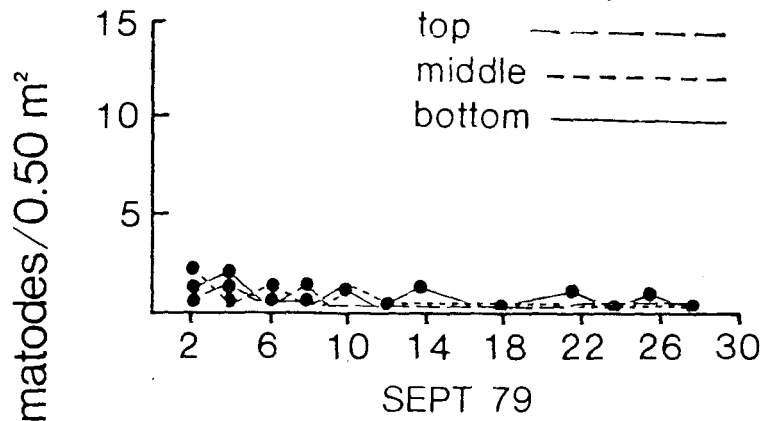
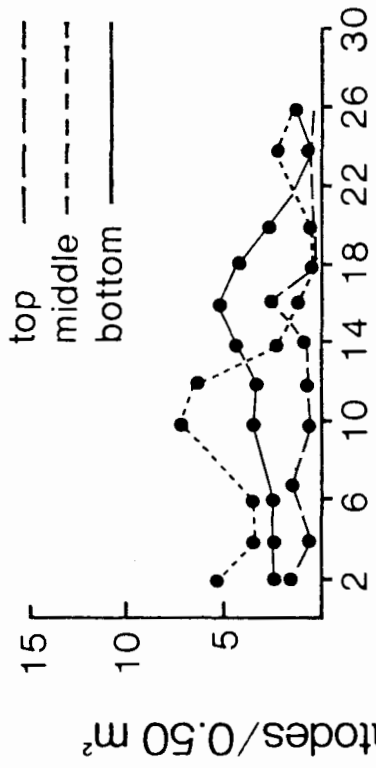
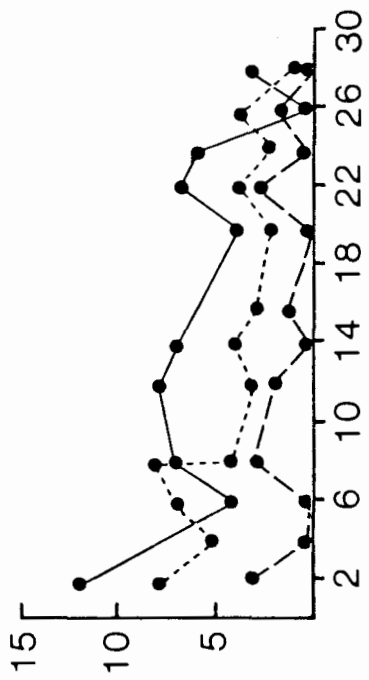
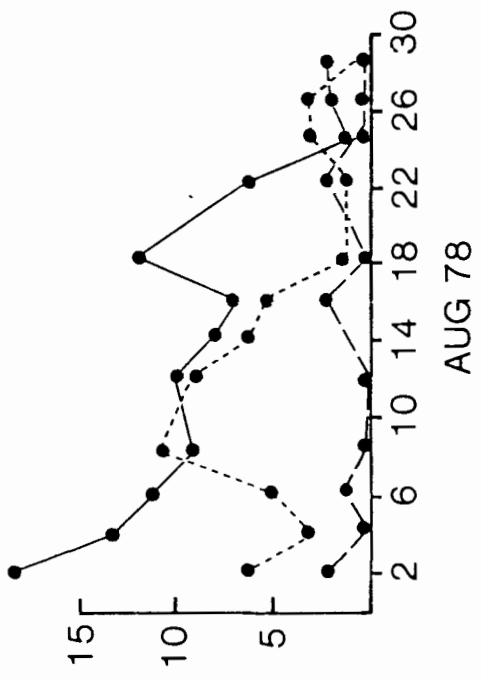


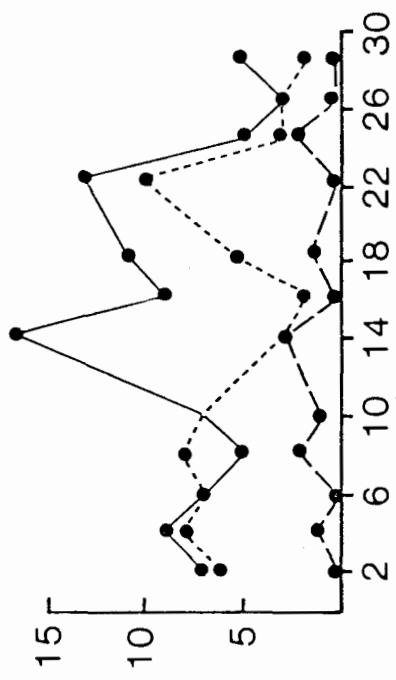
Fig. 12: Density of Monhystera refringens (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
from July 1978 to October 1978 (from deep to
shallow depth).



JULY 78

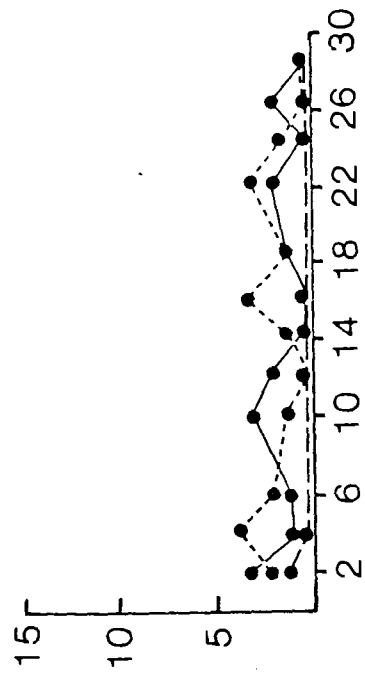


SEPT 78

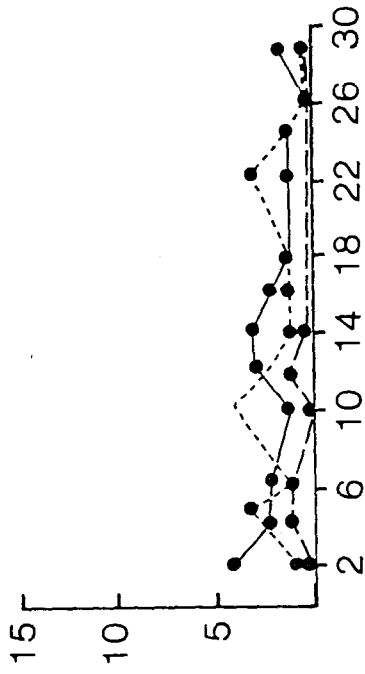


OCT 78

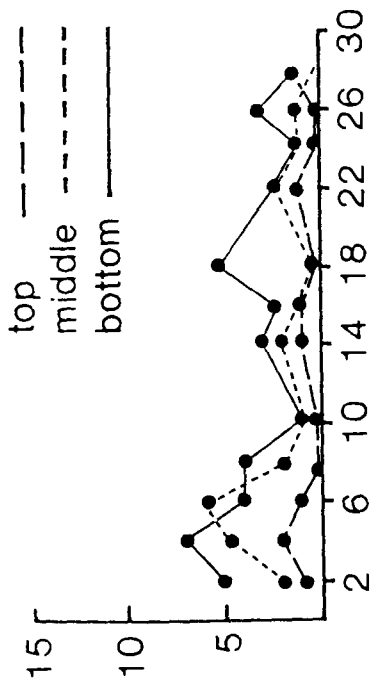
Fig. 13: Density of Monhystera refringens (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
from November 1978 to March 1979. (from deep to
shallow depth).



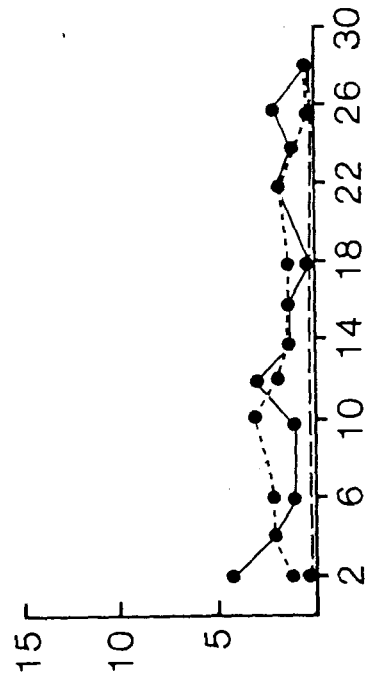
JAN 79



MAR 79



NOV 78

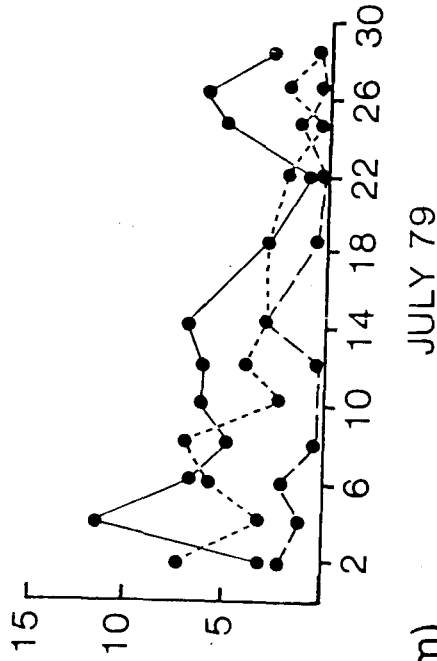
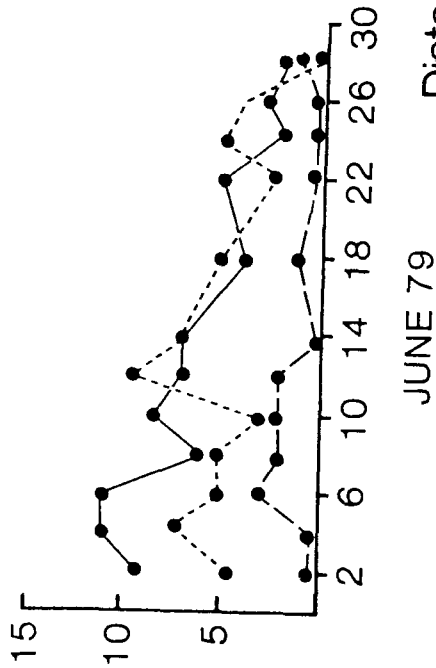
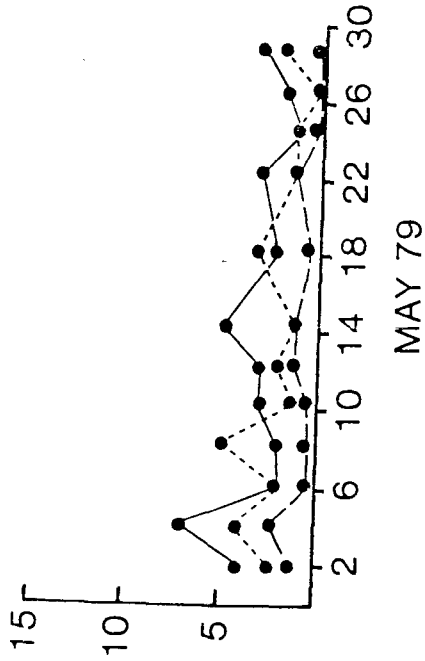
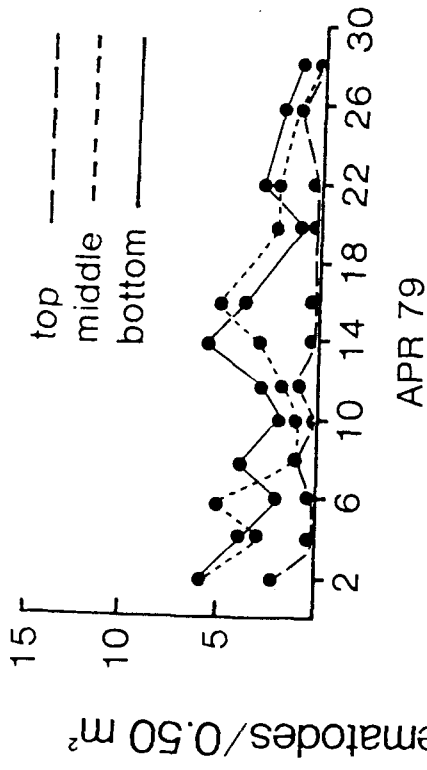


FEB 79

No. of Nematodes/0.50 m²

Distance (m)

Fig. 14: Density of Monhystera refringens (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
from April 1979 to July 1979. (from deep to shallow
depth).



Distance (m)

No. of Nematodes/0.50 m²

Fig. 15: Density of Monhystera refringens (nematodes/
0.5 m²) on top, middle and bottom blade samples
of Macrocystis integrifolia along a depth transect
for September and November 1979. (from deep to
shallow depth).

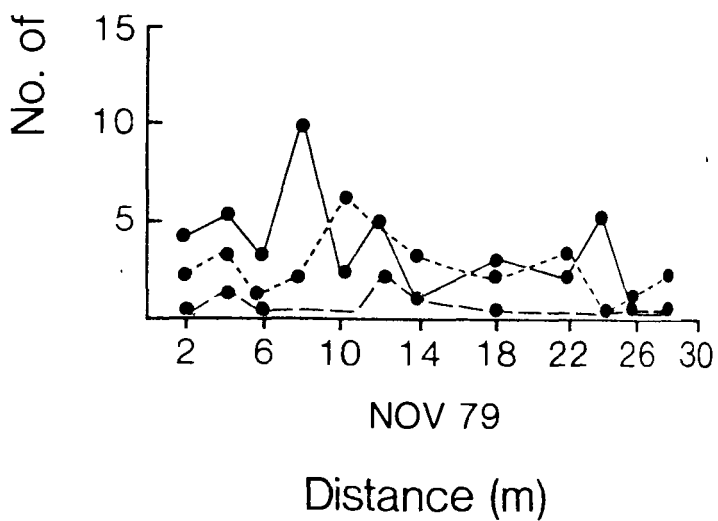
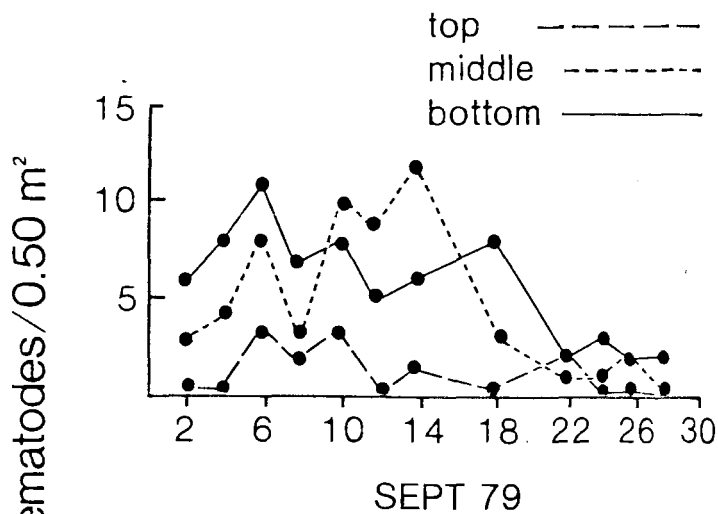


Fig. 16: Density of Monhystera disjuncta (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect from July 1978 to October 1978 (from deep to shallow depth).

Fig. 17: Density of Monhystera disjuncta (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect from November 1978 to March 1979 (from deep to shallow depth).

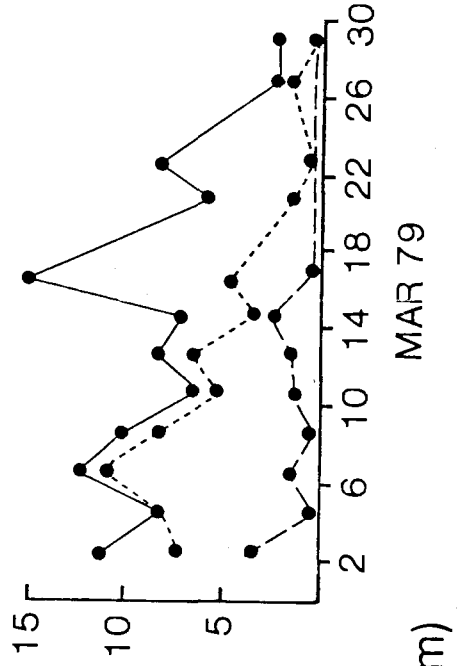
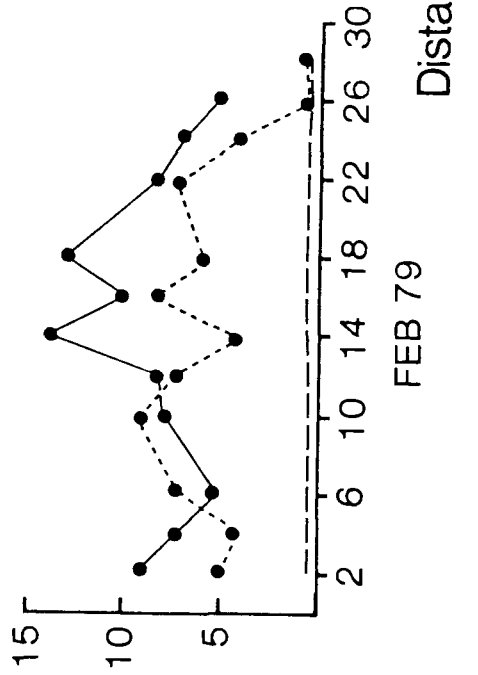
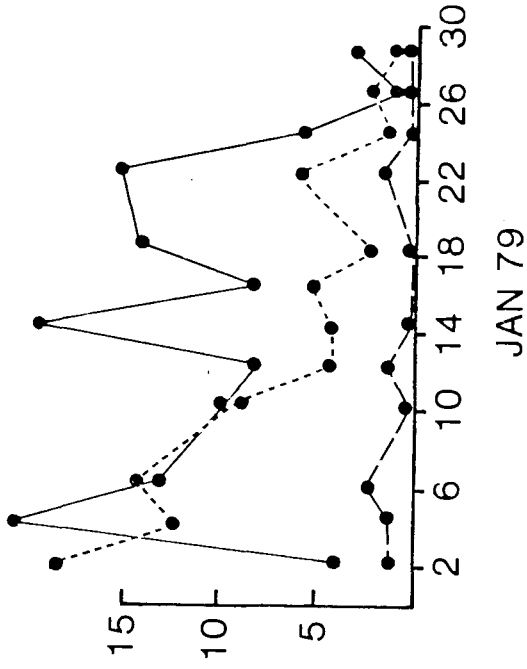
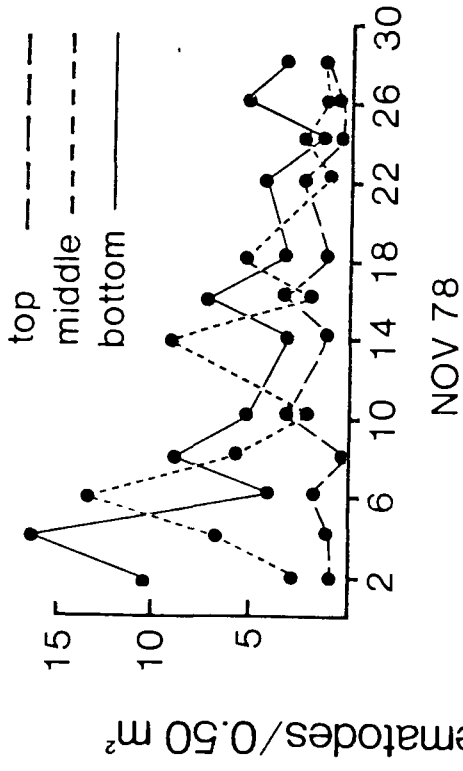
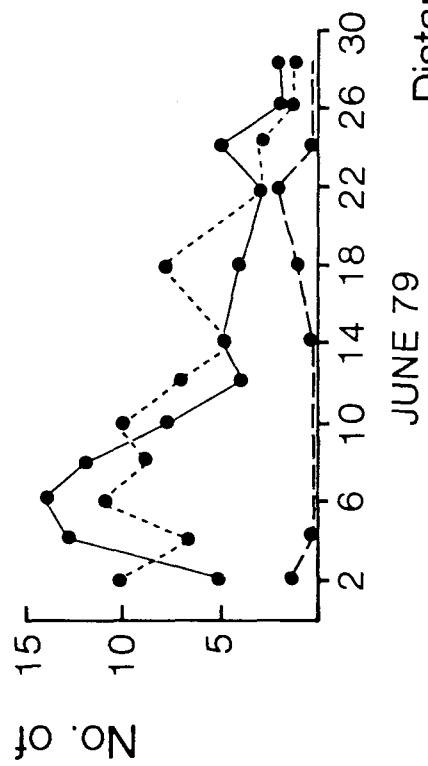
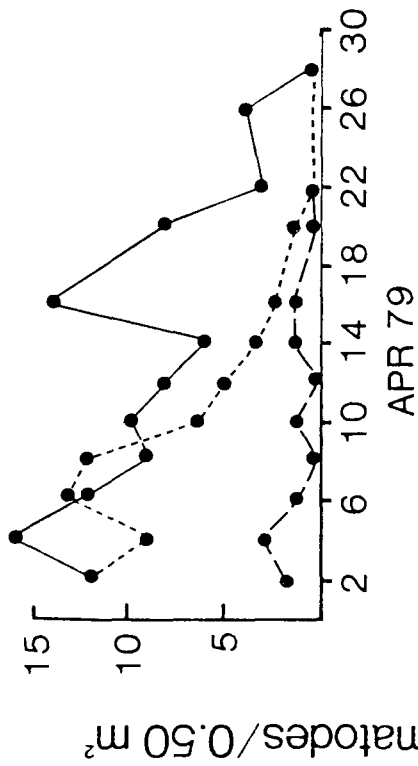
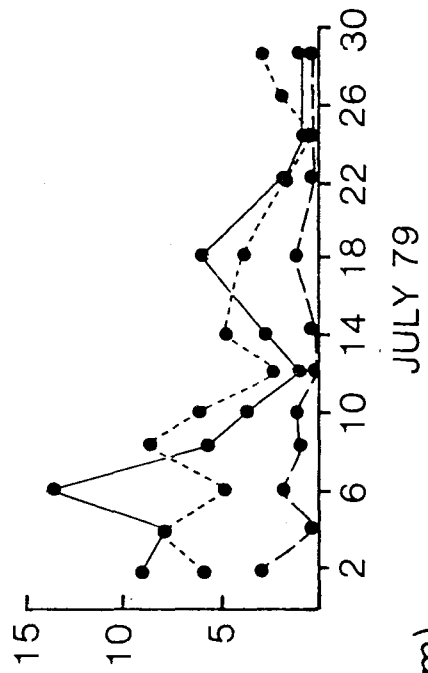
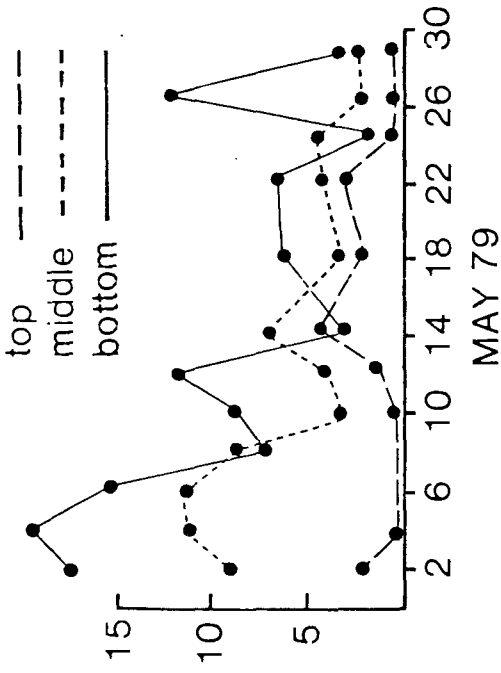


Fig. 18: Density of Monhystera disjuncta (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect from April 1979 to July 1979 (from deep to shallow depth).



Distance (m)

Fig. 19: Density of Monhystera disjuncta (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect for September and November 1979 (from deep to shallow depth).

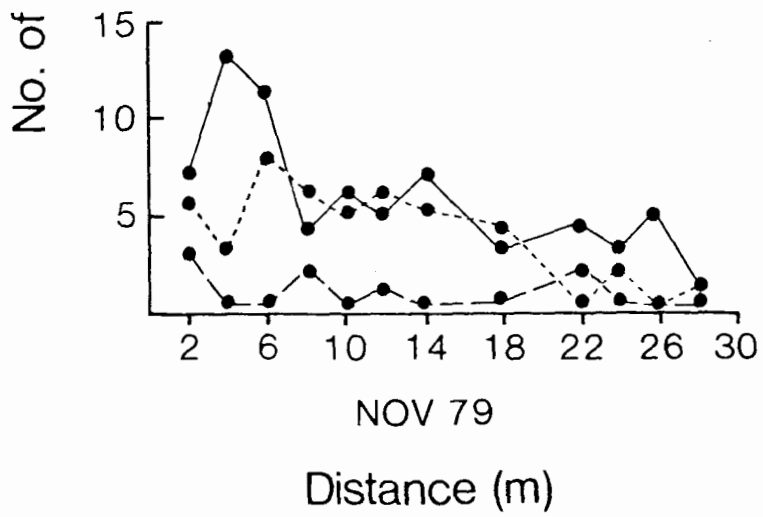
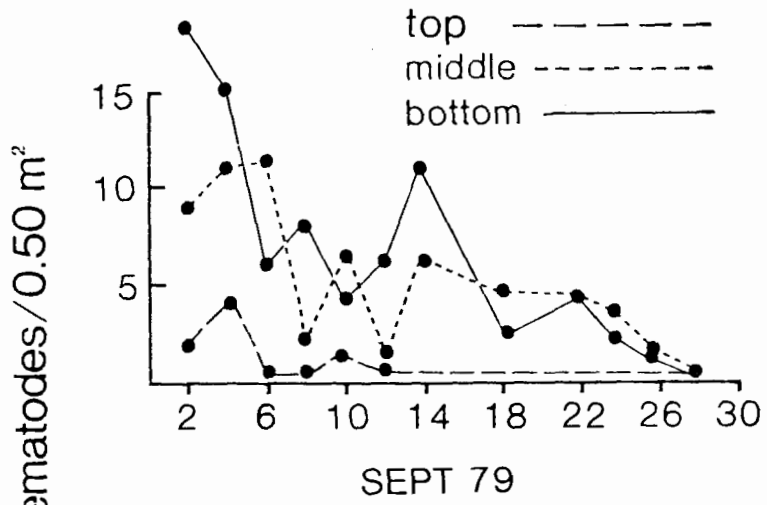


Fig. 20: Density of non-predominant nematode species (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect from July 1978 to October 1978. (from deep to shallow depth).

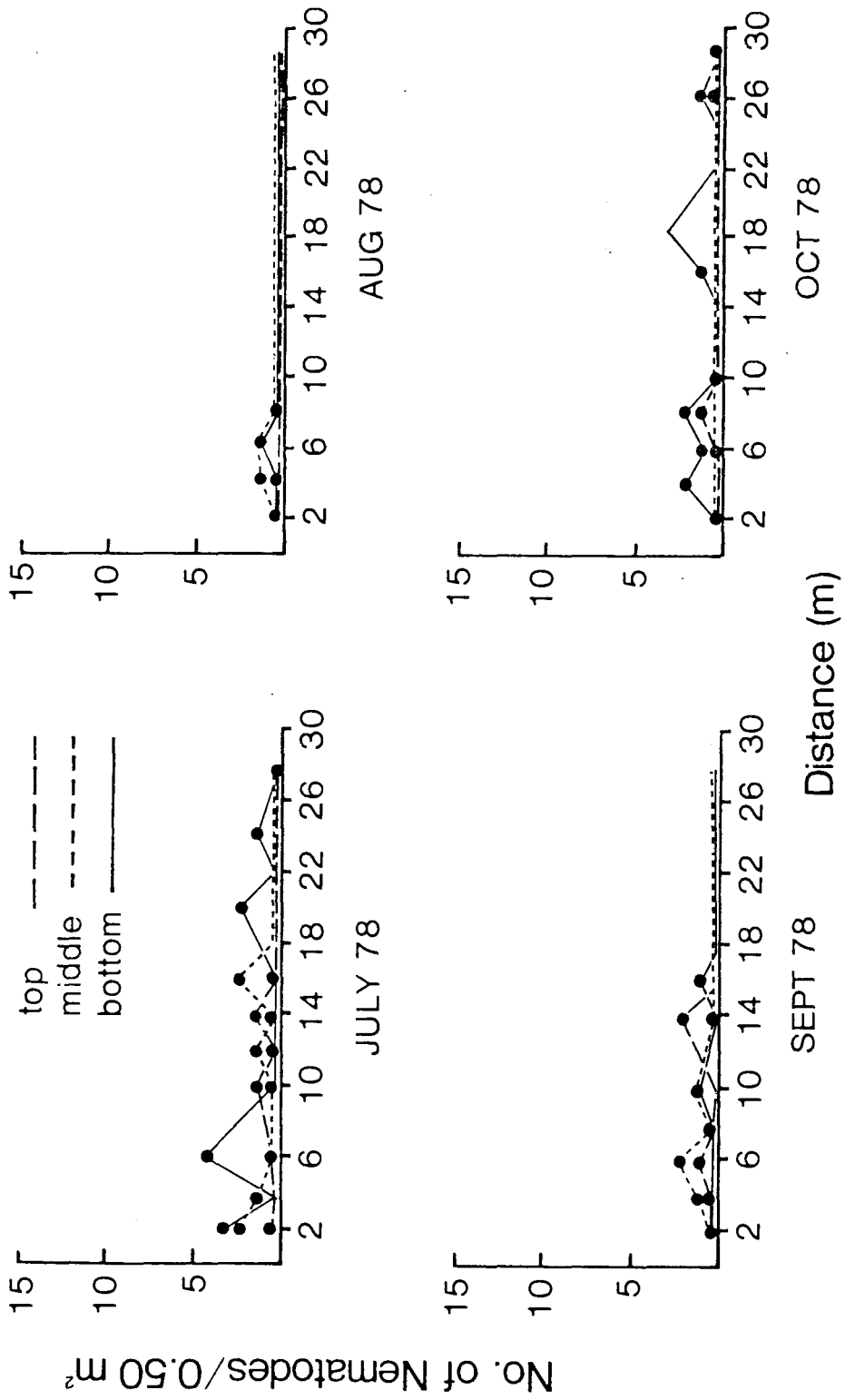


Fig. 21: Density of non-predominant nematode species (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect from November 1978 to March 1979 (from deep to shallow depth).

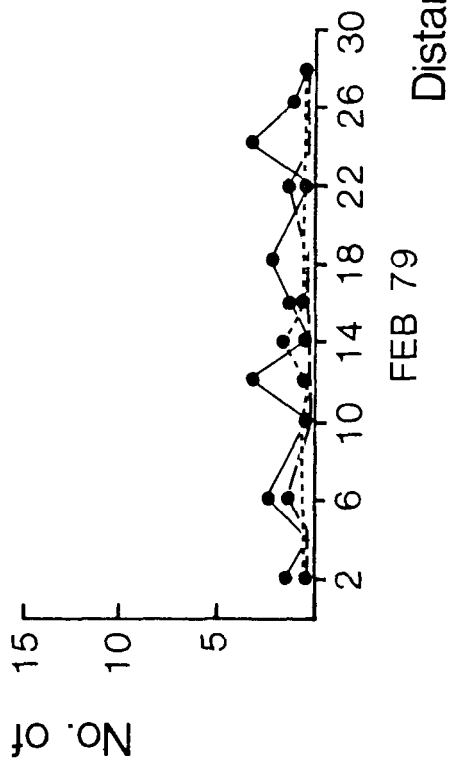
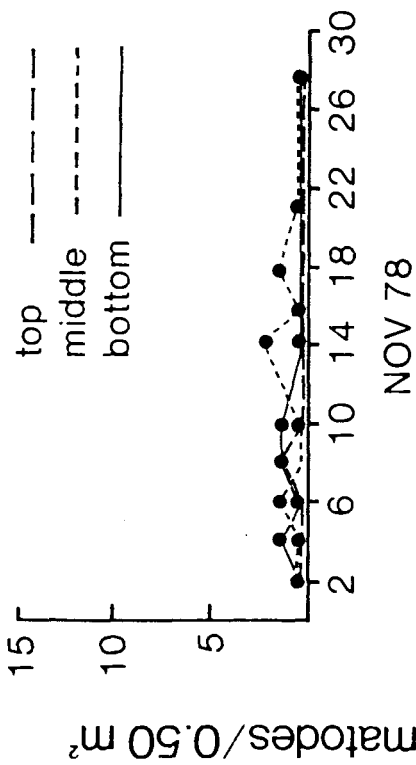
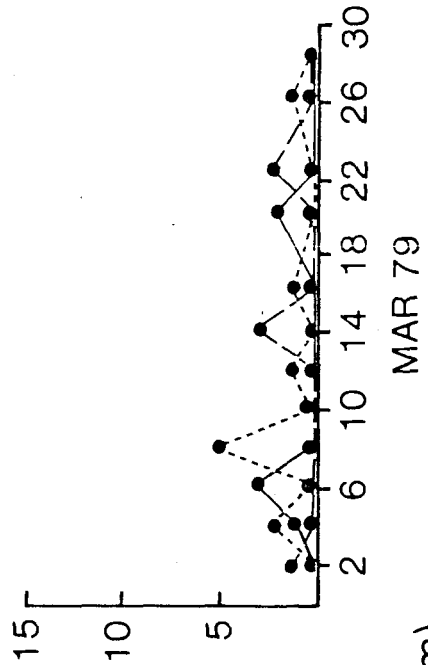
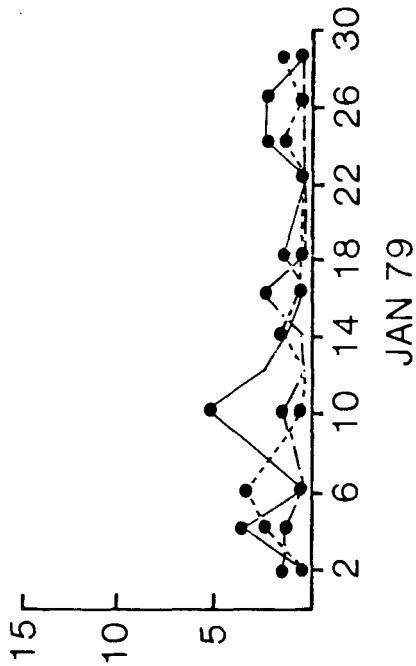


Fig. 22: Density of non-predominant nematode species (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect from April 1979 to July 1979 (from deep to shallow depth).

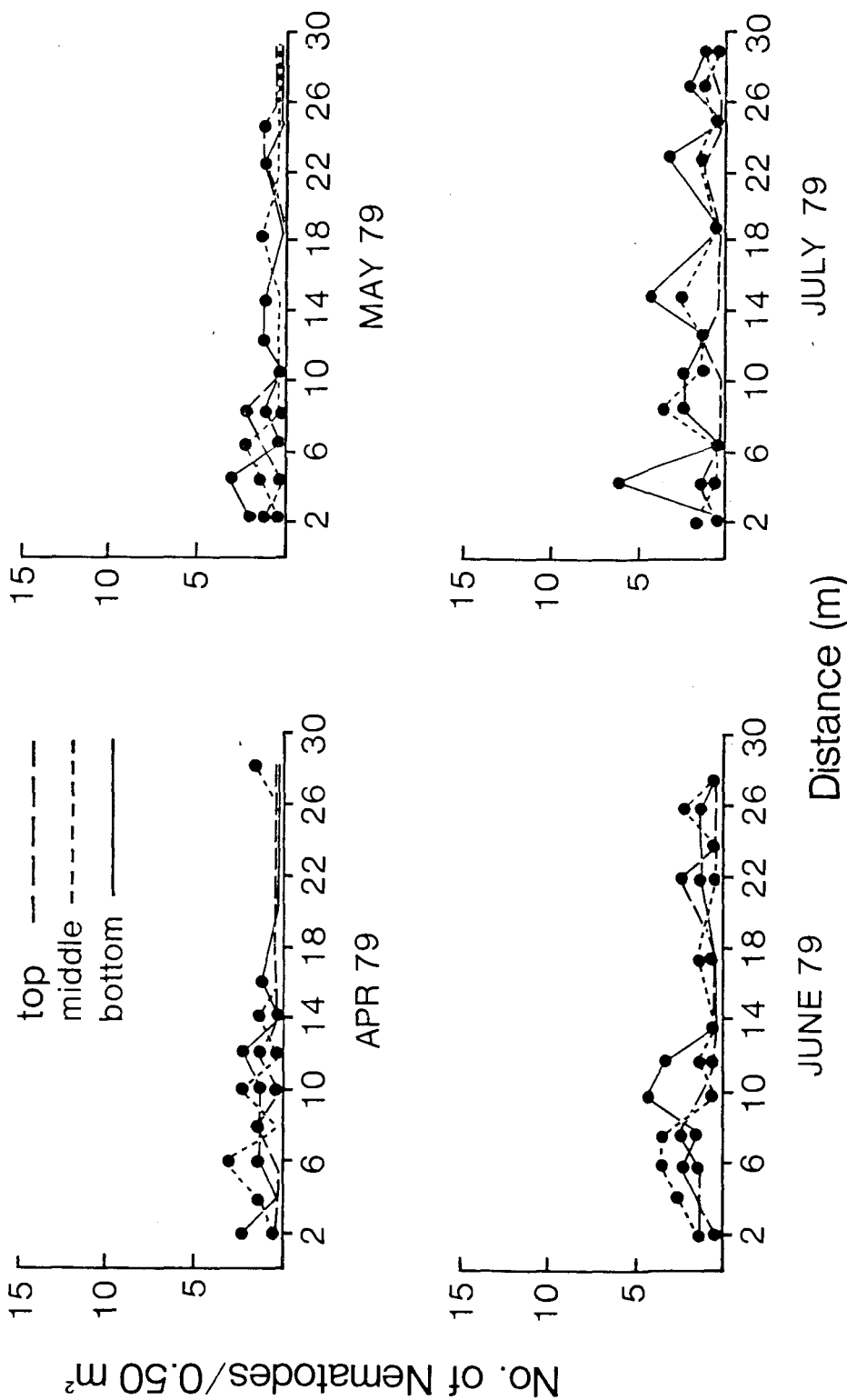


Fig. 23: Density of non-predominant nematode species (nematodes/0.5 m²) on top, middle and bottom blade samples of Macrocystis integrifolia along a depth transect for September and November 1979 (from deep to shallow depth).

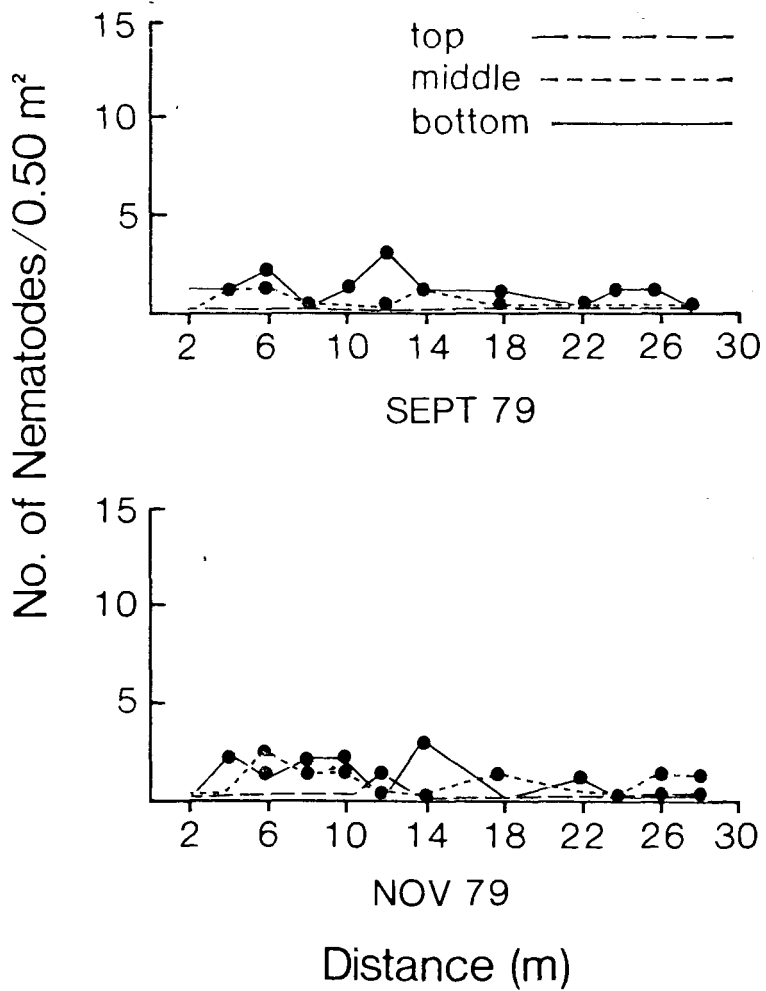


TABLE 6. Distribution of *Prochromadorella neapolitana* with reference to *Macrocystis integrifolia* blade position and depth F values for a grouped two-level ANOVA calculated on the number of nematodes/0.05m² for each sample month.

| Month | Blade Position | | Depth | | Blade Position X Depth | |
|------------|-------------------|--------------|-------------------|--------------|------------------------|--------------|
| | F _{2,24} | Significance | F _{3,24} | Significance | F _{2,24} | Significance |
| July, 1978 | 20.54 | *** | 1.83 | n.s. | 0.30 | n.s. |
| August | 1.90 | n.s. | 0.63 | n.s. | 0.19 | n.s. |
| September | 0.50 | n.s. | 1.11 | n.s. | 1.24 | n.s. |
| October | 1.90 | n.s. | 1.17 | n.s. | 0.77 | n.s. |
| November | 9.02 | *** | 0.63 | n.s. | 0.29 | n.s. |
| December | - | - | - | - | - | - |
| Jan., 1979 | 6.12 | ** | 0.12 | n.s. | 1.02 | n.s. |
| February | 8.05 | ** | 3.37 | n.s. | 2.22 | n.s. |
| March | 3.47 | * | 1.54 | n.s. | 0.57 | n.s. |
| April | 3.31 | n.s. | 0.44 | n.s. | 0.08 | n.s. |
| May | 7.54 | *** | 0.77 | n.s. | 1.06 | n.s. |
| June | 10.33 | *** | 0.91 | n.s. | 0.14 | n.s. |
| July | 13.27 | *** | 0.11 | n.s. | 1.18 | n.s. |
| August | - | - | - | - | - | - |
| September | 1.73 | n.s. | 1.55 | n.s. | 0.41 | n.s. |
| October | - | - | - | - | - | - |
| November | 6.27 | ** | 1.14 | n.s. | 0.89 | n.s. |

*** P<0.005

TABLE 7 Distribution of *Monhystera refringens* with reference to *Macrocystis integrifolia* blade position and depth F values for a grouped two-level ANOVA calculated on the number of nematodes/0.5m² for each sample month.

| Month | Blade Position | | Depth | | Blade Position X Depth | |
|------------|-------------------|--------------|-------------------|--------------|------------------------|--------------|
| | F _{2,24} | Significance | F _{3,24} | Significance | F _{2,24} | Significance |
| July, 1978 | 5.75 | ** | 0.20 | n.s. | 0.30 | n.s. |
| August | 11.46 | *** | 0.64 | n.s. | 0.19 | n.s. |
| September | 15.97 | *** | 2.84 | n.s. | 1.24 | n.s. |
| October | 21.25 | *** | 1.34 | n.s. | 0.77 | n.s. |
| November | 6.37 | ** | 0.61 | n.s. | 0.29 | n.s. |
| December | - | - | - | - | - | - |
| Jan., 1979 | 6.68 | *** | 1.05 | n.s. | 1.02 | n.s. |
| February | 16.22 | *** | 3.33 | * | 2.22 | n.s. |
| March | 7.34 | *** | 1.44 | n.s. | 0.57 | n.s. |
| April | 8.50 | *** | 0.13 | n.s. | 0.08 | n.s. |
| May | 11.73 | *** | 0.70 | n.s. | 1.06 | n.s. |
| June | 9.26 | *** | 0.06 | n.s. | 0.14 | n.s. |
| July | 13.62 | *** | 1.23 | n.s. | 1.18 | n.s. |
| August | - | - | - | - | - | - |
| September | 7.36 | *** | 0.69 | n.s. | 0.41 | n.s. |
| October | - | - | - | - | - | - |
| November | 7.97 | *** | 1.32 | n.s. | 0.89 | n.s. |

*** P<0.005

TABLE 8. Distribution of *Monhystera disjuncta* with reference to *Macrocystis integrifolia* blade position and depth F values for a grouped two-level ANOVA calculated on the number of nematodes/0.05m² for each sample month.

| Month | Blade Position | | Depth | | Blade Position X Depth | |
|------------|-------------------|--------------|-------------------|--------------|------------------------|--------------|
| | F _{2,24} | Significance | F _{3,24} | Significance | F _{2,24} | Significance |
| July, 1978 | 9.25 | *** | 0.16 | n.s. | 0.62 | n.s. |
| August | 23.15 | *** | 0.80 | n.s. | 0.75 | n.s. |
| September | 5.74 | ** | 1.23 | n.s. | 0.84 | n.s. |
| October | 21.62 | *** | 0.72 | n.s. | 0.33 | n.s. |
| November | 4.94 | * | 0.20 | n.s. | 0.38 | n.s. |
| December | - | - | - | - | - | - |
| Jan., 1979 | 7.50 | *** | 1.17 | n.s. | 1.42 | n.s. |
| February | 20.80 | *** | 0.21 | n.s. | 0.30 | n.s. |
| March | 11.87 | *** | 0.01 | n.s. | 0.17 | n.s. |
| April | 8.46 | *** | 0.20 | n.s. | 0.06 | n.s. |
| May | 10.46 | *** | 0.70 | n.s. | 0.27 | n.s. |
| June | 16.29 | *** | 1.16 | n.s. | 1.02 | n.s. |
| July | 5.61 | ** | 0.39 | n.s. | 0.13 | n.s. |
| August | - | - | - | - | - | - |
| September | 5.91 | ** | 1.16 | n.s. | 0.22 | n.s. |
| October | - | - | - | - | - | - |
| November | 11.61 | *** | 0.70 | n.s. | 0.89 | n.s. |

*** P<0.005

TABLE 9. Distribution of the three predominant nematode species with reference to Macrocyctis integrifolia blade position and depth F values for an unreplicated three-level ANOVA calculated on the number of nematodes/0.5m² for each sample month.

| Month | Blade Position | | Depth | Nematode sp. | | Blade Position | | Blade Position | | Nematode sp. | |
|---------------|-------------------|--------------------|-----------|--------------------|-------------------|----------------|---|-------------------|---------|--------------------|---|
| | F _{2,52} | F _{13,52} | | F _{13,52} | F _{2,52} | X Depth | X | F _{4,52} | X Depth | F _{26,52} | X |
| July, 1978 | 53.67 *** | 4.50 *** | 6.75 *** | 1.37 n.s. | 4.73 *** | 1.40 n.s. | | | | | |
| August | 43.17 *** | 3.05 *** | 34.65 *** | 0.89 n.s. | 9.97 *** | 0.81 n.s. | | | | | |
| September | 34.54 *** | 6.44 *** | 47.82 *** | 1.40 n.s. | 8.84 *** | 2.75 *** | | | | | |
| October | 57.18 *** | 1.98 * | 55.12 *** | 1.13 n.s. | 12.11 *** | 1.21 n.s. | | | | | |
| November | 28.88 *** | 5.06 *** | 12.69 *** | 2.02 * | 2.35 | 0.53 n.s. | | | | | |
| December | - | - | - | - | - | - | | | | | |
| January, 1979 | 10.10 *** | 1.63 | 21.82 *** | 0.95 n.s. | 6.59 *** | 1.31 n.s. | | | | | |
| February | 62.34 *** | 3.00 *** | 80.87 *** | 1.68 n.s. | 23.25 *** | 1.84 * | | | | | |
| March | 39.27 *** | 4.63 *** | 56.44 *** | 1.32 n.s. | 15.45 *** | 1.91 * | | | | | |
| April | 42.11 *** | 6.03 *** | 46.10 *** | 1.52 n.s. | 11.22 *** | 2.40 *** | | | | | |
| May | 34.12 *** | 4.00 *** | 17.20 *** | 1.33 n.s. | 3.48 * | 0.84 n.s. | | | | | |
| June | 62.37 *** | 5.83 *** | 0.54 n.s. | 1.82 * | 0.39 n.s. | 0.72 n.s. | | | | | |
| July | 39.28 *** | 5.03 *** | 2.67 n.s. | 1.20 n.s. | 0.60 n.s. | 0.65 n.s. | | | | | |
| August | - | - | - | - | - | - | | | | | |
| September | 31.88 *** | 5.93 *** | 35.88 *** | 1.47 n.s. | 6.69 *** | 2.86 *** | | | | | |
| October | - | - | - | - | - | - | | | | | |
| November | 29.74 *** | 2.91 *** | 7.36 *** | 0.83 n.s. | 1.63 n.s. | 0.71 n.s. | | | | | |

*** P < 0.005

The remainder of the year, mid and bottom blade samples had few individuals and there were very low or no individuals associated with the top blades (Figs. 12-15).

The distribution of P. neapolitana was similar to that of M. disjuncta and M. refringens for the months of July and November 1978 and May to July 1979 (Figs. 8-11). For the remainder of the year distribution was difficult to ascertain because of low numbers.

Distribution of the remaining nematode species is shown in Figures 20-23. However, due to their low numbers and to combined grouping of the species, no distribution trends on the blades could be confidently established.

DISCUSSION I

Although the area of nematode/algal associations is not well documented, published studies show that certain nematode taxa are commonly found in association with a variety of marine algae (Colman, 1940; Hopper, 1970; L'Hardy, 1962). All the fully identified nematode species recorded on the kelp in the present study have been recorded previously as being associated with algae (Wieser, 1953). The three most abundant nematode species have been recorded as being associated with algal species ranging from members of the Laminariales (Chitwood and Murphy, 1964; Moore, 1971, 1973a, 1973b; Wieser, 1952a) to Zostera marina (eelgrass) beds (Hopper and Meyers, 1967a, 1967b). The other species that have been recorded as seaweed associates include Araeolaimus elegans, Paracanthanchus sp., Anticoma acuminata and Oncholaimus dujardinii on Laminaria hyperborea (Moore, 1971) and O. dujardinii on turtle grass (Hopper and Meyers, 1967a) and some intertidal algae (Wieser, 1952b). The other two species, Chromadora nudicapitata and Chromadorina laeta have not been recorded as algal associates in any studies other than the Chilean survey by Wieser (1953). Generally, chromadorids appear to be the dominant group of nematodes associated with algae. In fact, Hopper and Meyers (1967a) found that two chromadorids, Chromadora macrolaimoides and Chromadorina epidemos Filipjev, in two separate Thalassia beds, comprised over 60% of the nematode

fauna. Wieser (1952b), studying the microfauna inhabiting intertidal seaweeds on rocky coasts, and Tietjen and Lee (1973), observing estuarine nematode populations on Enteromorpha intestinalis Link and Z. marina, found chromadorids to be the dominant group. Moore (1971, 1973a, 1973b) found that enoplids were predominant in the holdfasts of Laminaria hyperborea. In the present study on M. integrifolia, the chromadorids represented the largest single number of species, four of nine. However, the monhysterids were the most abundant.

Investigators have sometimes tried to associate nematode feeding strategies, based on Wieser's (1952) classifications, with nematodes found on marine algae. They have found that the epigrowth feeders are generally the dominant group (Hopper and Meyers, 1967a, 1967b; Tietjen and Lee, 1973). In the present study, chromadorids are considered to be equivalent to Wieser's (1952a) epigrowth feeders, the monhysterids equivalent to the nonselective feeders, the oncholaimid as omnivores and the araeolamids and anticomid as selective deposit feeders. As chromadorids are very abundant in algal habitats the dominance of the epigrowth feeding type is supported. Hopper and Meyers (1967b) found the epigrowth feeders to comprise 58% - 85% of the total nematode fauna. However, in the present study as monhysterids are the most abundant species on Macrocystis, there is an overall dominance of the

non-selective feeding type except during the summer months (see Figs. 4-6).

The most striking aspect of nematode distribution on the kelp is the marked seasonal differences in the nematode populations. In particular, the three most abundant species show distinct seasonal variations in their abundance (Figs. 4-6 and Tables 3-5). The most distinct pattern is exhibited by the chromadorid, P. neapolitana, which Wieser (1953) considered to be an epigrowth feeder. For most of the year, P. neapolitana ranges from 2% - 12% of the total nematode population but in the late spring, the population increases greatly to a maximum in mid-summer (see Fig. 4). The increase is characterized by the addition of substantially more juveniles and males. Hopper and Meyers (1967a) found similar results, in that populations of epigrowth feeders were lowest during the early spring and increased to a maximum during the summer.

Observations on two other chromadorids, Chromadora macrolaimoides and Chromadorina germanica Butschli, both associated with Enteromorpha intestinalis (L.) Link, exhibited their highest abundance during the early and mid-summer months respectively (Tietjen and Lee, 1973, 1977). Some researchers have suggested that these large increases in population of epigrowth feeders during the summer months may, in part, be associated to a corresponding increase in the epiflora, especially diatoms, which are a major food source for the species (Hagerman, 1966; Tietjen and Lee, 1973). It has

been suggested that the species diversity of the epiphytes on the alga has a greater effect than does the specificity of the nematodes on the alga itself (Hopper and Meyers, 1967a; Ott, 1967).

In addition to food availability, the influence of temperature was probably an important parameter affecting the reproductive potential of P. neapolitana. Tietjen and Lee (1977a) observed that C. germanica had the shortest generation times, 12-15 days, at 26°/oo and 20-30 C. Chromadorina germanica was most abundant in July and August when the average temperature and salinity was 24 C and 25°/oo respectively. At temperatures below 20 C, the generation time increased to nearly 100 days. It may be that the low winter and spring temperatures in Dodger Channel lengthen the generation time of P. neapolitana and thus, in part, explain the low abundance levels.

The two monhysterids found in the M. integrifolia blade samples exhibit different seasonal variations from P. neapolitana. Both species are over 20% of the relative abundance for most of the year, particularly M. disjuncta which was observed as high as 76% in January 1979. M. disjuncta was the predominant species for most of the year (Fig. 6, Table 5).

M. refringens was in greatest abundance during the late summer and early fall (Fig. 5, Table 4). Both monhysterids have been classified as non-selective deposit feeders (Chitwood and Murphy, 1964). Hopper and Meyers (1967a) observed the increased abundance of non-selective

deposit feeders (Monhystera spp.) primarily in the spring. The seasonal abundance pattern described by Hopper and Meyers (1967a) closely fits my observations on M. disjuncta (see Fig. 6, Table 5). However, M. refringens did not correspond to this pattern and this may reflect different food preferences (see Section V).

M. disjuncta and M. refringens exhibited little variation in the male, female and juvenile population structure between July 1978 and November 1979, except for M. refringens during the early fall. However, as with P. neapolitana, temperature and salinity probably contribute to the variation in seasonal abundance. Observations by Gerlach and Schrage (1971) on the life cycle of M. disjuncta isolated from Laminaria in the North Sea, and reared at various temperatures correspond to my observations (Table 20) that the seasonality may be related to temperature (Zaika and Makarova, 1979). Generally, life cycles and generation times shorten with increasing temperature (Hopper et al., 1973).

Similar results were observed with another monhysterid, Monhystera denticulata Timm, isolated from Z. marina with the average generation time of 18 days at 26° and 15 C while it was 180 days at 5 C (Tietjen and Lee, 1972). M. refringens reflects quite well these trends. However, M. disjuncta exhibits a quite different response in the field. The data on M. disjuncta from the blade samples of Macrocystis indicated large numbers of this species occurring

throughout the year. This likely reflects the fact that little variation in the age' structure has been observed. The temperature and salinity effects on M. disjuncta generation times are not reflected in the field data.

All three major species exhibited the same general distribution pattern on the blades of M. integrifolia. Most nematodes were found in the bottom, fewer in the middle and very few or none in the top blade samples (see Figs. 8-23). Nematode numbers increased with water depth, particularly on the middle and bottom blades. Exceptions to this pattern were observed in the late winter and spring months for M. refringens and P. neapolitana and this was probably due to their reduced levels of abundance (Figs. 8-15). A variety of factors have been suggested that may affect distribution, namely algal shape, surface area, degree of encrustation and degree of silt or detritus deposits (Dahl, 1948; North, 1971; Ott, 1967; Wieser, 1952a, 1959c; Wing and Clendenning, 1971). In particular, increased levels of detritus cause large increases in nematode populations on heavily affected fronds (Wieser, 1952). Mukai (1971), studying epiphytic animals on Sargassum, observed large increases in nematode numbers in early spring which corresponded to the time of maximum degradation of the plant itself. Merrith (1973), studying deposit-feeding nematodes, observed the largest number of them to be at stations with the thickest layer of detritus. Wing and Clendenning (1971) observed that nematode populations were

much higher on blades of M. pyrifera that were totally encrusted and deteriorating than on unencrusted, clean growing blades. They also observed that the nematodes were greatest on the deep rather than on the surface blades. The deep blades are the oldest on the plant, and usually carry the greatest amount of epiphytes (North, 1971). This is also true for M. integrifolia (Lobban, 1978; Roland, 1980). Ott (1967) observed deposit feeders to predominate at the bases of two species of Cystoseria. Also, monhysterids have been found to be good indicators on an increased degree of sedimentation of detritus (Wieser, 1959). Hence, with a much more developed microenvironment associated with the older blades of Macrocystis, which are themselves the lower blade areas of the plant, and the associated concentration of life forms associated with this older tissue, then the distribution and seasonality of the nematodes, particularly Monhystera spp., appear to be closely tied.

RESULTS II

a) Prochromadorella neapolitana

i) Summer conditions

Data for the "cafeteria" experiments are included in Appendices 1-6 and the analyses are presented in Table 10 and 11. Individuals of this species accumulated significantly ($P > 0.005$) more around food wells than around the non-food interwells or, around the wells of the control plate. Observations from the three experiments indicated that P. neapolitana was attracted in greater numbers to diatom than to bacteria sources. In experiment I, nematodes accumulated in the greatest numbers and were equally attracted to diatom species G. marina and R. curvata. Diatom species N. jamalinensis and S. tabulata showed equivalent accumulations of P. neapolitana. Values obtained from wells containing N. jamalinensis were very close to nematode numbers observed from the control wells.

In experiment II, almost 50% of the nematodes were found to accumulate in food wells containing C. scutellum. Fewer nematodes accumulated around diatom species A. longissipes and A. acutiuscula but did so equally ($\bar{X} = 19.0$ and 16.75 respectively). Nematode accumulations in food wells containing G. olivaceum were not significantly different from those around the control wells. P. neapolitana did not accumulate significantly more around any

TABLE 10. Total, male and female mean recruitment of *Prochromadorella neapolitana* to diatom and bacteria species (see Table 1) under simulated summer conditions

| Expt N=4 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|-------|-----------|------|-----------|------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{x} | S.D. |
| I | D1 | 37.00 | 8.60 | 19.50 | 6.61 | 17.50 | 4.51 |
| | D2 | 32.25 | 6.85 | 14.50 | 2.89 | 17.75 | 8.22 |
| | D3 | 5.50 | 3.70 | 3.75 | 2.75 | 1.75 | 0.96 |
| | D4 | 12.00 | 5.10 | 6.25 | 3.59 | 5.75 | 3.10 |
| II | D5 | 3.25 | 2.22 | 2.50 | 2.38 | 0.75 | 1.50 |
| | D6 | 48.00 | 5.72 | 23.00 | 6.98 | 25.00 | 4.90 |
| | D7 | 19.00 | 4.90 | 9.75 | 0.96 | 9.25 | 4.99 |
| | D8 | 16.75 | 4.99 | 7.00 | 3.56 | 9.75 | 2.75 |
| III | B1 | 4.00 | 2.94 | 1.50 | 1.73 | 2.50 | 3.70 |
| | B2 | 2.75 | 2.06 | 1.50 | 1.30 | 1.25 | 1.89 |
| | B3 | 15.75 | 3.95 | 9.75 | 2.63 | 6.00 | 1.83 |
| | B4 | 3.75 | 2.87 | 1.50 | 1.73 | 2.25 | 2.87 |
| I | Outside | 3.00 | 2.31 | 1.50 | 1.73 | 1.50 | 0.58 |
| II | Outside | 4.25 | 0.50 | 2.25 | 1.71 | 2.00 | 1.41 |
| III | Outside | 64.00 | 12.03 | 29.50 | 4.65 | 34.50 | 9.18 |
| I | Interwell | 2.56 | 2.50 | 1.13 | 1.15 | 1.44 | 1.75 |
| II | Interwell | 2.19 | 1.28 | 1.38 | 1.20 | 0.81 | 0.91 |
| III | Interwell | 2.44 | 2.22 | 1.56 | 1.63 | 0.88 | 1.02 |
| I | Control | 4.75 | 2.92 | 3.00 | 2.39 | 1.75 | 1.67 |
| II | Control | 3.13 | 3.04 | 2.25 | 2.60 | 0.88 | 1.13 |
| III | Control | 3.63 | 3.66 | 2.38 | 2.26 | 1.25 | 1.58 |

TABLE 11. Analysis of attractiveness and preference of *Prochromadorella neapolitana* to diatom and bacteria species (see Table 1) using Chi-square, two-level ANOVA and Student-Newman-Keuls testing under simulated summer and winter conditions.

| | Diatom sp. D1-D4 | | Diatom sp. D5-D8 | | Bacteria sp. B1-B4 | |
|---------------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|
| | Summer | Winter | Summer | Winter | Summer | Winter |
| $\chi^2_{1,2}$ | 3960.78 *** | 2681.96 *** | 3969.73 *** | 2527.95 *** | 238.58 *** | 115.15 *** |
| 2 Level ANOVA | | | | | | |
| $F_{3,9}$ (Plate Effect) | 0.12 n.s. | 0.08 n.s. | 0.12 n.s. | 1.29 n.s. | 0.38 n.s. | 1.37 n.s. |
| $F_{3,9}$ (Food Effect) | 18.17 *** | 6.50 * | 49.39 *** | 44.13 *** | 22.67 *** | 17.01 *** |
| Student-Newman-Keuls Test | <u>D3 D4 D1 D2</u> | <u>D3 D4 D2 D1</u> | <u>D5 D8 D7 D6</u> | <u>D5 D7 D8 D6</u> | <u>B2 B4 B1 B3</u> | <u>B4 B2 B1 B3</u> |

*** P < 0.005

one of three species of bacteria (namely B1, B2 and B4) than around interwells and control wells. However, significantly more accumulated around species B3 than around control wells and this species attracted nematodes significantly more ($P > 0.005$) than the other three species. In general, males and females accumulated with equal facility at any of the food sources.

ii) Winter conditions

Data for the "cafeteria" experiments are included in Appendices 1-6 and the analyses are presented in Table 11 and 12. P. neapolitana exhibited the same overall pattern of accumulations around particular food wells as found under simulated summer conditions, with a few exceptions. This nematode accumulated to a greater degree in food wells with diatoms but the mean number of nematodes found in a particular food well was proportionally lower than under summer conditions. Also, the number of nematodes not associated with any particular well, food or control, was higher than those observed under summer conditions.

In experiments II and III, the only difference from the observations under summer conditions were changes in the mean accumulation of nematodes in wells containing diatom species, A. longispes and A. acutiuscula and bacteria species B4 and B2. The male/female ratio of those nematodes attracted to food wells was approximately 1:1.

TABLE 12. Total, male and female mean recruitment of Prochromadorella neapolitana to diatom and bacteria species (see Table 1) under simulated winter conditions.

| Expt N=4 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|------|-----------|------|-----------|------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{X} | S.D. |
| I | D1 | 27.50 | 6.56 | 12.00 | 4.97 | 15.50 | 5.80 |
| | D2 | 23.50 | 6.61 | 11.75 | 5.12 | 11.75 | 5.44 |
| | D3 | 7.50 | 4.65 | 3.75 | 1.94 | 3.75 | 5.56 |
| | D4 | 14.25 | 6.80 | 8.75 | 5.06 | 5.50 | 3.87 |
| II | D5 | 4.50 | 2.08 | 2.00 | 1.41 | 2.50 | 2.65 |
| | D6 | 36.75 | 3.59 | 19.00 | 7.53 | 17.75 | 8.10 |
| | D7 | 14.25 | 6.70 | 7.75 | 5.91 | 6.50 | 2.65 |
| | D8 | 15.00 | 3.16 | 7.25 | 2.99 | 7.75 | 2.63 |
| III | B1 | 3.25 | 2.63 | 1.50 | 0.58 | 1.75 | 2.22 |
| | B2 | 2.50 | 1.00 | 1.75 | 0.96 | 0.75 | 0.96 |
| | B3 | 12.75 | 4.11 | 4.75 | 2.87 | 8.00 | 3.56 |
| | B4 | 2.00 | 1.41 | 1.00 | 0.00 | 1.00 | 1.41 |
| I | Outside | 18.75 | 4.57 | 8.50 | 6.24 | 10.25 | 1.89 |
| II | Outside | 18.25 | 7.68 | 7.75 | 3.86 | 10.50 | 9.00 |
| III | Outside | 71.25 | 2.87 | 35.25 | 1.71 | 36.00 | 4.08 |
| I | Interwell | 2.13 | 1.26 | 1.44 | 1.41 | 0.69 | 0.79 |
| II | Interwell | 2.81 | 2.10 | 1.56 | 1.31 | 1.25 | 1.48 |
| III | Interwell | 2.06 | 1.77 | 1.38 | 1.31 | 0.68 | 0.95 |
| I | Control | 4.13 | 2.90 | 2.13 | 2.30 | 2.00 | 1.60 |
| II | Control | 2.88 | 2.59 | 1.63 | 2.07 | 1.25 | 1.67 |
| III | Control | 3.25 | 1.58 | 1.75 | 1.67 | 1.50 | 0.93 |

b) Monhystera refringens

i) Summer conditions

Data for the "cafeteria" experiments are included in Appendices 7-12 and the analyses are presented in Table 13 and 14. This nematode species exhibited an equal accumulation to both diatom and bacteria food wells. In all three experiments, M. refringens was observed to exhibit a significant ($P > 0.005$) accumulation of individuals in food wells when compared with control wells. In experiment I, higher nematode numbers were found in association with only diatom species S. tabulata. In comparison, the remaining three diatom species had low mean accumulations of nematodes (G. marina = 10.5, R. curvata = 11.5 and N. jamalinensis = 14.0). Nematode attraction to wells containing S. tabulata exhibited the highest mean accumulation ($\bar{X} = 40.0$) in comparison to all three experiments.

In experiment II, M. refringens exhibited higher mean accumulations in wells with A. acutiuscula and A. longissipes (22.5 and 21.75 respectively) as compared to the accumulations in wells containing diatom species C. scutellum and G. olivaceum (15.0 and 9.25 respectively). In comparison, the nematode was found to show no significant accumulations to any single diatom species. When presented bacterial food sources, M. refringens was found to be equally attracted to bacteria species B2, B3 and B4 with a significantly lower mean accumulation in B1 food wells. In general, none of the

TABLE 13. Total, male and female mean recruitment of Monhystera refringens to diatom and bacteria species (see Table 1) under simulated summer conditions.

| Expt N=4 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|------|-----------|------|-----------|-------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{X} | S.D. |
| I | D1 | 10.50 | 5.20 | 6.75 | 4.65 | 3.75 | 2.63 |
| | D2 | 11.50 | 6.24 | 3.00 | 0.82 | 8.50 | 6.76 |
| | D3 | 14.00 | 4.55 | 5.50 | 3.70 | 8.50 | 6.19 |
| | D4 | 40.00 | 8.29 | 21.75 | 6.18 | 18.25 | 12.53 |
| II | D5 | 9.25 | 3.10 | 6.25 | 4.79 | 3.00 | 2.16 |
| | D6 | 15.00 | 6.98 | 7.50 | 2.74 | 7.50 | 6.56 |
| | D7 | 21.75 | 9.36 | 7.25 | 4.11 | 14.50 | 11.00 |
| | D8 | 22.25 | 6.85 | 12.75 | 7.14 | 9.50 | 6.24 |
| III | B1 | 9.25 | 3.50 | 5.00 | 2.71 | 4.25 | 2.50 |
| | B2 | 21.50 | 4.93 | 10.25 | 3.40 | 11.25 | 6.08 |
| | B3 | 24.00 | 5.29 | 8.00 | 2.94 | 16.00 | 5.94 |
| | B4 | 17.25 | 2.63 | 10.75 | 3.69 | 6.50 | 6.03 |
| I | Outside | 10.00 | 3.37 | 7.00 | 2.45 | 3.00 | 1.41 |
| II | Outside | 12.50 | 2.65 | 6.50 | 3.87 | 6.00 | 4.24 |
| III | Outside | 13.00 | 3.92 | 8.00 | 3.74 | 5.00 | 3.46 |
| I | Interwell | 3.50 | 2.80 | 1.50 | 1.26 | 2.00 | 2.19 |
| II | Interwell | 4.81 | 2.40 | 2.44 | 1.79 | 2.38 | 2.06 |
| III | Interwell | 3.75 | 3.07 | 2.00 | 2.28 | 1.75 | 1.61 |
| I | Control | 3.25 | 3.11 | 2.00 | 2.51 | 1.25 | 1.49 |
| II | Control | 3.50 | 2.14 | 1.50 | 1.69 | 2.00 | 2.00 |
| III | Control | 4.50 | 3.21 | 2.38 | 2.07 | 2.13 | 2.30 |

TABLE 14. Analysis of attractiveness and preference of *Monhystera refringens* to diatom and bacteria species (see Table 1) using Chi-square, two-level ANOVA and Student-Newman-Keuls testing under simulated summer and winter conditions.

| | Diatom sp. D1-D4 | | Diatom sp. D5-D8 | | Bacteria sp. B1-B4 | |
|---------------------------------|------------------|-------------|------------------|-------------|--------------------|-------------|
| | Summer | Winter | Summer | Winter | Summer | Winter |
| $\chi^2_{1,2}$ | 3017.22 *** | 2317.41 *** | 2480.77 *** | 2317.41 *** | 2691.98 *** | 1854.0 *** |
| 2-Level ANOVA | | | | | | |
| F _{3,9} (Plate Effect) | 0.10 n.s. | 0.19 n.s. | 0.06 n.s. | 0.37 n.s. | 0.32 n.s. | 0.50 n.s. |
| F _{3,9} (Food Effect) | 15.83 *** | 13.39 *** | 2.42 n.s. | 3.92 n.s. | 7.98 ** | 2.88 n.s. |
| Student-Newman-Keuls Test | D1 D2 D3 D4 | D3 D1 D2 D4 | D5 D6 D7 D8 | D5 D7 D6 D8 | B1 B2 B3 B4 | B1 B2 B3 B4 |

*** P<0.005

TABLE 15. Total, male and female mean recruitment of Monhystera refringens to diatom and bacteria species (see Table 1) under simulated winter conditions.

| Expt N=4 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|------|-----------|-------|-----------|------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{X} | S.D. |
| I | D1 | 13.25 | 5.12 | 6.50 | 4.65 | 6.75 | 2.99 |
| | D2 | 15.25 | 4.92 | 7.50 | 3.70 | 7.75 | 2.87 |
| | D3 | 11.75 | 3.59 | 5.00 | 3.56 | 6.75 | 1.89 |
| | D4 | 33.00 | 5.48 | 17.00 | 9.35 | 16.00 | 4.08 |
| II | D5 | 11.25 | 4.79 | 6.50 | 7.33 | 4.75 | 2.63 |
| | D6 | 19.25 | 3.95 | 6.50 | 3.32 | 12.75 | 3.50 |
| | D7 | 16.75 | 6.65 | 11.75 | 7.41 | 5.00 | 2.16 |
| | D8 | 20.50 | 5.00 | 8.50 | 4.36 | 12.00 | 6.48 |
| III | B1 | 9.50 | 4.93 | 5.00 | 3.56 | 4.50 | 2.65 |
| | B2 | 20.00 | 8.04 | 11.50 | 7.60 | 8.50 | 5.57 |
| | B3 | 19.75 | 6.85 | 8.75 | 2.50 | 11.00 | 7.87 |
| | B4 | 11.50 | 3.11 | 5.25 | 2.50 | 6.25 | 4.43 |
| I | Outside | 19.75 | 3.30 | 9.75 | 6.70 | 10.00 | 4.97 |
| II | Outside | 22.50 | 5.00 | 10.75 | 11.56 | 11.75 | 7.93 |
| III | Outside | 25.00 | 6.06 | 12.25 | 6.60 | 12.75 | 7.14 |
| I | Interwell | 1.75 | 1.06 | 1.06 | 0.85 | 0.69 | 0.95 |
| II | Interwell | 2.38 | 1.41 | 1.50 | 1.37 | 0.88 | 1.20 |
| III | Interwell | 3.56 | 1.79 | 1.81 | 1.60 | 1.75 | 1.73 |
| I | Control | 4.13 | 2.70 | 2.38 | 2.20 | 1.75 | 1.49 |
| II | Control | 2.25 | 1.67 | 1.125 | 0.99 | 1.125 | 1.25 |
| III | Control | 3.50 | 2.20 | 1.50 | 1.60 | 2.00 | 2.20 |

nematode accumulations in food wells was as low as those found in the control wells. The male/female ratio of the attracted nematodes was approximately 1:1.

ii) Winter conditions

Data for the "cafeteria" experiments are included in Appendices 7-12 and the analyses are presented in Table 14 and 15. In experiments I and II, this nematode exhibited a similar display of accumulation compared with summer except that relatively fewer nematodes occurred in the food wells and more of them occurred outside any particular well. In experiment II, the mean number of nematodes in wells with C. scutellum increased. Nematode accumulations in A. longissipes were observed to decrease. In experiment III, the mean accumulation of nematodes in all four bacteria wells were not significantly different from each other. A 1:1 ratio with respect to male/female accumulations in wells was observed in all experiments.

c) Monhystera disjuncta

i) Summer conditions

Data for the "cafeteria" experiments are included in Appendices 13-18 and the analyses are presented in Table 16 and 17. In all three experiments, nematodes accumulated in significant numbers ($P < 0.005$) in the food wells. M. disjuncta showed a marked attraction to bacteria as compared to diatoms. In experiment I, M. disjuncta exhibited a low response to all diatom species except S. tabulata in which

TABLE 16. Total, male and female mean recruitment of Monhystera disjuncta to diatom and bacteria species (see Table 1) under simulated summer conditions.

| Expt N=4 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|-------|-----------|-------|-----------|-------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{X} | S.D. |
| I | D1 | 9.75 | 4.43 | 5.50 | 4.36 | 4.25 | 2.50 |
| | D2 | 5.00 | 2.94 | 1.00 | 0.82 | 4.00 | 3.27 |
| | D3 | 7.75 | 4.99 | 5.25 | 2.87 | 2.50 | 3.00 |
| | D4 | 21.75 | 6.08 | 12.25 | 6.70 | 9.50 | 5.57 |
| II | D5 | 8.75 | 3.30 | 5.25 | 3.31 | 3.50 | 3.00 |
| | D6 | 9.50 | 5.45 | 7.25 | 5.00 | 2.25 | 1.26 |
| | D7 | 0.50 | 0.58 | 0.25 | 0.50 | 0.25 | 0.50 |
| | D8 | 5.50 | 2.52 | 3.00 | 4.08 | 2.50 | 2.38 |
| III | B1 | 18.00 | 5.90 | 7.50 | 6.60 | 10.50 | 1.30 |
| | B2 | 19.25 | 3.86 | 8.50 | 4.93 | 10.75 | 5.62 |
| | B3 | 27.75 | 7.27 | 19.25 | 5.44 | 8.50 | 4.51 |
| | B4 | 14.75 | 2.63 | 6.25 | 2.06 | 8.50 | 1.91 |
| I | Outside | 45.50 | 19.23 | 20.00 | 15.01 | 25.50 | 10.41 |
| II | Outside | 63.00 | 6.27 | 25.25 | 3.59 | 27.75 | 5.50 |
| III | Outside | 9.75 | 4.27 | 3.75 | 2.50 | 6.00 | 3.37 |
| I | Interwell | 2.56 | 2.16 | 1.50 | 1.46 | 1.06 | 1.53 |
| II | Interwell | 3.19 | 2.56 | 2.25 | 2.18 | 0.94 | 1.12 |
| III | Interwell | 2.63 | 1.78 | 1.19 | 0.66 | 1.44 | 1.55 |
| I | Control | 2.38 | 2.33 | 2.00 | 2.14 | 0.38 | 0.52 |
| II | Control | 3.38 | 2.83 | 2.00 | 2.07 | 1.38 | 1.41 |
| III | Control | 4.00 | 2.00 | 2.50 | 1.31 | 1.50 | 1.93 |

Table 17. Analysis of attractiveness and preference of *Monhyстера disjuncta* to diatom and bacteria species (see Table 1) using Chi-square, 2 level ANOVA and Student-Newman-Keuls testing under simulated summer and winter conditions.

| | Diatom sp. D1-D4 | | Diatom Sp. D5-D8 | | Bacteria Sp. B1-B4 | |
|---------------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|
| | Summer | Winter | Summer | Winter | Summer | Winter |
| $\chi^2_{1,2}$ | 875.22 *** | 698.23 *** | 213.53 *** | 311.31*** | 3301.74 *** | 2474.85 *** |
| 2 Level ANOVA | | | | | | |
| $F_{3,9}$ (Plate Effect) | 4.94 * | 0.20 n.s. | 0.41 n.s. | 0.81 n.s. | 0.63 n.s. | 1.04 n.s. |
| $F_{3,9}$ (Food Effect) | 19.24 *** | 2.92 n.s. | 4.85 * | 1.70 n.s. | 4.08 * | 2.95 n.s. |
| Student-Newman-Keuls Test | <u>D2 D3 D1 D4</u> | <u>D2 D3 D1 D4</u> | <u>D7 D8 D5 D6</u> | <u>D7 D5 D6 D8</u> | <u>B4 B1 B2 B3</u> | <u>B4 B1 B2 B3</u> |

*** $P < 0.005$

the relatively high accumulations were later attributed to bacterial contamination.

The accumulations of nematodes in the food wells containing diatom species of experiment II (see Table 1) were even lower than the values observed in experiment I. The mean number of nematodes found in the wells containing A. longissipes and A. acutiuscula had values comparable to the control wells. The mean number of nematodes outside the wells was relatively high ($\bar{X} = 63.0$) compared to the other two nematode species. In experiment III, the majority of the nematodes were found to accumulate in the wells with rod-shaped (B1 and B3) rather than with coccoid bacteria (B2 and B4).

Data and analyses for the two additional "cafeteria" experiments are presented in Appendices 19-20 and Table 18 respectively. The results indicated, as observed above, a higher accumulation of nematodes in wells containing rod as opposed to coccoid bacteria. The male/female ratio in all experiments was approximately 1:1.

ii) Winter conditions

Few nematodes were attracted to diatom food sources as measured by the accumulations at the food wells (Tables 17 and 19). The number of individuals not in food wells was high. In experiments I and II, none of the diatom sources showed any significant accumulations. In experiment III, M. disjuncta showed a reduced response to bacterial sources, such that it accumulated with relatively equal

TABLE 18. Total, male and female mean recruitment of *Monhystera disjuncta* to rod and coccoid bacteria species (see Table 1) under simulated summer conditions.

| Expt N=8 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|-------|-----------|------|-----------|-------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{X} | S.D. |
| IV | B1 | 16.25 | 5.52 | 7.62 | 3.11 | 8.63 | 6.57 |
| | B3 | 25.13 | 10.02 | 13.63 | 5.32 | 11.50 | 8.97 |
| V | B2 | 14.75 | 5.68 | 9.38 | 5.90 | 5.37 | 2.96 |
| | B4 | 10.63 | 6.28 | 5.25 | 3.37 | 5.38 | 4.34 |
| IV | Outside | 8.75 | 5.12 | 3.50 | 3.51 | 5.25 | 4.43 |
| V | Outside | 39.50 | 12.45 | 15.00 | 7.75 | 24.50 | 11.27 |
| IV | Interwell | 1.81 | 1.52 | 0.56 | 0.73 | 1.25 | 1.18 |
| V | Interwell | 2.44 | 2.00 | 1.44 | 1.03 | 1.00 | 1.86 |
| IV | Control | 3.38 | 4.69 | 2.38 | 3.07 | 1.00 | 1.77 |
| V | Control | 2.25 | 1.98 | 1.25 | 1.49 | 1.00 | 1.60 |

TABLE 19. Total, male and female mean recruitment of *Monhystera disjuncta* to diatom and bacteria species (see Table 1) under simulated winter conditions.

| Expt N=4 | Species | Total | | Male | | Female | |
|-------------|-----------|-----------|-------|-----------|------|-----------|------|
| | | \bar{X} | S.D. | \bar{X} | S.D. | \bar{X} | S.D. |
| I | D1 | 10.50 | 7.19 | 4.50 | 2.87 | 6.00 | 4.83 |
| | D2 | 5.75 | 3.50 | 3.75 | 3.59 | 2.00 | 2.00 |
| | D3 | 6.25 | 4.57 | 3.50 | 3.51 | 2.75 | 1.89 |
| | D4 | 17.25 | 6.24 | 6.75 | 6.29 | 10.50 | 7.00 |
| II | D5 | 7.50 | 4.80 | 5.50 | 4.36 | 2.00 | 2.16 |
| | D6 | 8.25 | 4.65 | 4.25 | 3.20 | 4.00 | 4.24 |
| | D7 | 3.00 | 0.82 | 2.00 | 2.00 | 1.00 | 0.82 |
| | D8 | 9.25 | 4.79 | 3.25 | 2.99 | 6.00 | 2.71 |
| III | B1 | 16.25 | 4.79 | 7.75 | 5.06 | 8.50 | 2.89 |
| | B2 | 18.00 | 3.83 | 8.50 | 3.70 | 9.50 | 3.32 |
| | B3 | 22.75 | 6.80 | 10.50 | 2.52 | 12.25 | 5.85 |
| | B4 | 12.75 | 3.30 | 5.25 | 3.77 | 7.50 | 5.20 |
| I | Outside | 47.75 | 6.50 | 24.75 | 2.75 | 23.00 | 8.37 |
| II | Outside | 58.50 | 14.20 | 28.25 | 5.74 | 30.25 | 8.77 |
| III | Outside | 18.50 | 8.74 | 10.50 | 6.34 | 8.00 | 4.97 |
| I | Interwell | 3.13 | 2.00 | 1.69 | 1.45 | 1.44 | 1.09 |
| II | Interwell | 3.38 | 3.03 | 1.69 | 2.09 | 1.69 | 1.82 |
| III | Interwell | 2.94 | 1.34 | 1.88 | 1.09 | 1.06 | 1.06 |
| I | Control | 2.50 | 2.33 | 0.75 | 0.71 | 1.75 | 1.91 |
| II | Control | 4.50 | 3.16 | 1.75 | 1.04 | 2.75 | 3.37 |
| III | Control | 3.38 | 2.26 | 1.63 | 1.19 | 1.75 | 2.19 |

attraction to all four bacteria species. Also, the mean number of nematodes outside any particular well was significantly greater than summer condition values. As seen in all previous experiments, a 1:1 ratio of male/female accumulations was observed in the food wells.

d) Life Cycle Observations on Monhystera disjuncta

M. disjuncta was the only one of the three species to reproduce continuously in culture. P. neapolitana was never observed to reproduce and so cultures could be maintained only by the continual supplement with new individuals. M. refringens was observed only once to reproduce and so the observations are scant and inconclusive.

Observations on the life cycle of five female M. disjuncta (A-E) and their progeny (N) are summarized in Table 20. Under environmental conditions of $10\text{ C} \pm 1\text{ C}$, 30.8% and a 16/8 h day/night cycle, the mean number of days from egg deposition of the original female to egg deposition of the progeny was 23.25 days. A breakdown of this turnover time indicates that on average 4.3 days were required from egg deposition to hatching, 14.1 days from hatching to the appearance of recognizable sexual characteristics and a further 4.3 days for the appearance of the first egg in the uterus. Development of the egg to its deposition was observed to take an average of 2.3 days. Females deposited, on the average, 14.4 eggs.

TABLE 20. Duration of development and generation time (in days) of Monhystera disjuncta in cultures maintained under simulated summer conditions.

| | N=5 A | N=5 B | N=4 C | N=5 D | N=3 E | Mean |
|---|-----------------|-----------------|-----------------|-----------------|-----------------|----------------|
| I Development of eggs (egg deposition- hatching) | 3-4 (3.5) | 5-6 (5.5) | 1-2 (1.5) | 3-4 (3.5) | 7-8 (7.5) | 4.3 |
| II Postembryonic development (hatching - recognizable sexual characteristics) | 13-14 (13.5) | 12-13 (12.5) | 14-15 (14.5) | 14-15 (14.5) | 15-16 (15.5) | 14.1 |
| III Maturation (development of sexual characteristics - 1st egg in uterus) | 2-3 (2.5) | 3-4 (3.5) | 3-4 (3.5) | 4-5 (4.5) | 7-8 (7.5) | 4.3 |
| IV Development of eggs in uterus (1st egg in uterus - egg deposition) | 1-2 (1.5) | 2-3 (2.5) | 2-3 (2.5) | 2-3 (2.5) | - | 2.25 (N=4) |
| I + II + III | 19-22 (20.5) | 20-23 (21.5) | 18-21 (19.5) | 21-24 (22.5) | 29-32 (30.5) | 22.9 |
| I + II + III + IV | 20-24 (22) | 22-26 (24) | 20-24 (22) | 23-27 (25) | - | 23.25 (N=4) |
| Egg number | 15 | 19 | 18 | 12 | 8 | 14.4 |

DISCUSSION II

Very little detail is known of the microbiological assemblages found on macrophytic algal species. Some researchers have endeavoured to characterize these relationships under laboratory conditions and extrapolate the results so as to understand subsequent field observations.

A variety of techniques have been developed to culture marine nematodes and their food sources (Lee and Muller, 1975; Tietjen et al., 1970). Some studies have tried to characterize feeding preferences by way of gut contents (Jennings and Colman, 1970) but this has proved to be unsuccessful as researchers have found nematodes to exhibit both selective ingestion and digestion (Meyers and Hopper, 1966; Tietjen and Lee, 1977b; Viglierchio et al., 1969).

The three dominant species in this study were cultured and subjected to food preference testing using a technique modified from Lee et al. (1977). The nematodes were exposed to eight diatom and four bacteria species isolated from M. integrifolia blades. The eight diatom species were all identified as benthic types (Lee et al., 1975; Rao and Lewin, 1976) and five of the species had previously been identified as associates of M. integrifolia (see Table 1) (Roland, 1980).

Each nematode species exhibited some distinct food preferences. P. neapolitana, classified as an epigrowth

feeder, selected primarily for diatom species, in particular, Cocconeis scutellum, Gramatophora marina, and Rhoicosphenia curvata (see Tables 10-13). The preferential response of P. neapolitana for bacteria was little different from the controls except for the rod-shaped bacteria species, B3. Chromadorina germanica responds similarly in that it is attracted to varying degrees to a variety of diatom species but only to one bacterium to any extent (Tietjen and Lee, 1977; Lee et al., 1977). Chromadora macrolaimoides was observed to effectively utilize eleven of the twenty species of microalgae presented but only five species were able to sustain growth indefinitely (Tietjen and Lee, 1973). Calorific content of the algae does not appear to be a factor governing food preference selection in nematodes but selective digestion and ingestion are evident such that a selective diet in conjunction with a particular set of digestive enzymes may, in fact, dictate selection (Tietjen and Lee, 1977b).

The monhysterids, M. disjuncta and M. refringens, though both classified as non-selective deposit feeders, exhibited different feeding preferences from each other. M. disjuncta preferred particular species of bacteria rather than diatoms (see Table 18-22). Except for one species of diatom, Synedra tabulata, which was later found to be contaminated with bacteria, this nematode species showed no detectable pattern in relation to diatom food sources. Studies on Rhabditis marina, a bacterivorous species,

indicated that it avoided some species of diatoms but otherwise was randomly distributed on both control and experimental plates (Lee et al., 1977). Food size was a significant factor for bacterivorous nematodes (Tietjen and Lee, 1977). Hence, all the diatom species may well have been too large for M. disjuncta to successfully ingest.

M. refringens was observed to be attracted to both particular species of diatoms and bacteria (see Tables 14 - 17). However, M. refringens appears to be much more of a generalist than does M. disjuncta in its feeding preferences. Both species ingested rod and coccoid bacteria but the rod-shaped species were preferred over the coccoid forms.

Rhabditis marina fed heavily on Pseudomonas sp. and Flavobacterium sp., both rod bacteria, over the coccoid species, Micrococcus sp. (Lee et al., 1970). Wilt et al. (1973) found that bacteriophagous nematodes preferentially migrated and fed on Vibrio sp. (rod) under various conditions. Comparison of the feeding strategies of two bacteria feeding nematodes showed them to be highly selective for particular bacterial species, primarily rod forms (Tietjen, 1967, 1969).

Thus, if all marine nematodes are as selective as the three predominant species from this study, then spatial and

seasonal variations in the abundance of microorganisms may account for variations in abundance and distribution (Lee et al., 1977; Tietjen and Lee, 1973, 1977b). However, caution must be used because data from such laboratory studies are not directly applicable to feeding conditions in the field because of different nutrient values of food sources, growth and reproduction rates, feeding rates (Duncan et al., 1974) and alternate feeding sources (Chia and Warwick, 1970; Tietjen and Lee, 1975).

However, food is still likely to be a main contributing factor governing the population dynamics of nematode species under field conditions (Schiemer et al., 1980).

GENERAL DISCUSSION

This study has contributed to our understanding of the spatial and temporal distribution of nematodes living on M. integrifolia and has initiated our appreciation of microbial food chains involved in the nutrition of these animals.

From the previous sections, the three dominant nematode species isolated from the blades of M. integrifolia exhibited distinctive patterns of abundance and distribution and specific feeding preferences on the epiflora associated with the blades of this kelp. From these observations and previous studies, a possible explanation of the variations observed for these three nematode species may be derived.

From Roland's (1980) study on the per cent occurrence of several diatom taxa on M. integrifolia blades and the feeding preference data from this study, it appears that the observed seasonal and spatial distributions of the three predominant nematode species may be related to the variation in availability of particular food sources. Kita and Harada (1962) observed that the abundance of animals on Zostera blades was related to the abundance of epiphytes and microalgae. P. neapolitana appears to prefer the diatom species C. scutellum (Tables 10-12). This diatom species occurs in high levels of abundance in April to July which is the time of increasing abundance of P. neapolitana (Fig. 4). The large decrease in the relative

occurrence of this diatom species in August corresponds with the decreased abundance of P. neapolitana. The increased per cent occurrence of the diatom Rhoicosphenia curvata in October may be an explanation for the sudden increase in P. neapolitana numbers found in the November samples. P. neapolitana was observed to concentrate when in high abundance in the bottom blades. Roland (1980) found higher percentages of occurrence of diatoms on top blades with smaller densities in subsequently lower blades of M. integrifolia. It may be that only a particular level of food availability can effectively maintain a large population of this species. Above this level, the nematode abundance does not alter substantially. Also, Roland's (1980) study did not sample to the depth of senescent blades, which are primarily the bottom blades and, hence, the per cent occurrence of the food sources are unknown. P. neapolitana showed little or no preference for bacteria and thus did not utilize this food source though Roland (1980) found that both the rod and coccoid bacteria froms on the blades of Macrocystis exhibited no seasonality.

In contrast, the high relative abundance of M. disjuncta may, in part, be related to this unlimited unvaried seasonality of the bacteria. Studies have indicated that nematode numbers are directly related to decreases and increases in bacteria population levels (Boucher and Chamroux,

1976; Chamroux et al., 1977). Thus, with M. disjuncta's preference for bacteria, primarily the rod forms, and little association with diatom species, the observed high abundance of this species, in conjunction with its short generation time, may explain its seasonal distribution. Many investigators have observed large bacterial populations associated with seaweed species and most have found the bacteria in highest abundance on the older and deteriorating parts of the plant and to be primarily of the rod type (Kong and Kwong-yu Chan, 1979; Laycock, 1974; Mazure and Field, 1980). Also, most investigators on nematode feeding preference have observed the selective and non-selective deposit feeders to survive and reproduce more readily on rod-shaped bacterial sources (Hopper and Meyers, 1967a; Tietjen, 1967; Tietjen and Lee, 1973, 1977b). Therefore, the high abundance of M. disjuncta throughout the year and on the older blades of Macrocystis may, in a good part, be related to the bacterial populations. Studies have shown that epiphyte population encrustations tend to increase with tissue age (Ballantine, 1979; North, 1978) and that most nematodes are found associated with deteriorating heavily encrusted blades (Wing and Clendenning, 1971). Thus, the relation between these two factors may contribute substantially to nematode abundance and distribution.

M. refringens shows a more complex interaction between food availability and abundance. In the food preference experiments, M. refringens is preferentially associated with

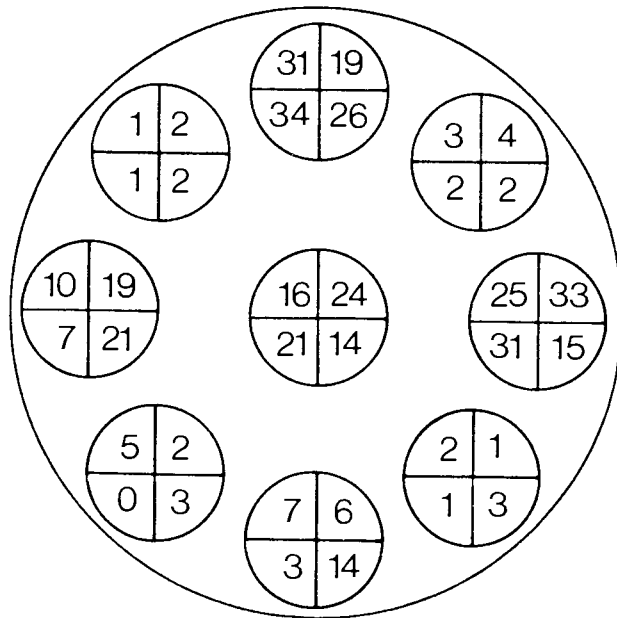
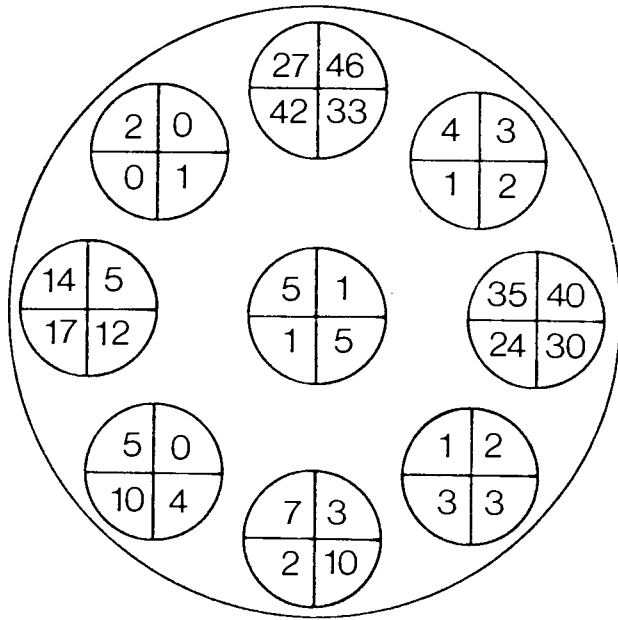
both particular diatom and bacteria species (Tables 13-15). Its most preferred diatom species, Synedra tabulata, is at its greatest abundance during late summer and fall (Roland, 1980). This corresponds to the observed increases in abundance of M. refringens that reached their highest levels in the fall. The remainder of the diatoms presented to M. refringens reflected fairly similar responses of attraction. These diatoms predominate primarily in the mid- and late summer but in relatively low per cent occurrence. The only exception is Cocconeis scutellum, which was most abundant during mid-summer and was the preferred food of P. neapolitana. In contrast, M. refringens was attracted to C. scutellum only moderately and showed little associated response to it in the field. With respect to bacteria, M. refringens exhibited almost the same response as M. disjuncta in terms of preference but not to the same degree. As a result, the variations in abundance of M. refringens may be in part related to the diatom food sources but its higher relative abundance compared to P. neapolitana through the year may be related to its partial affinity for bacterial food sources.

Thus, if marine nematodes are as selective in their feeding habits as this and other studies have indicated, then spatial and temporal variations in the abundance of epiphytic microorganisms could account for the spatial and seasonal variations in abundance of marine nematodes. Hence,

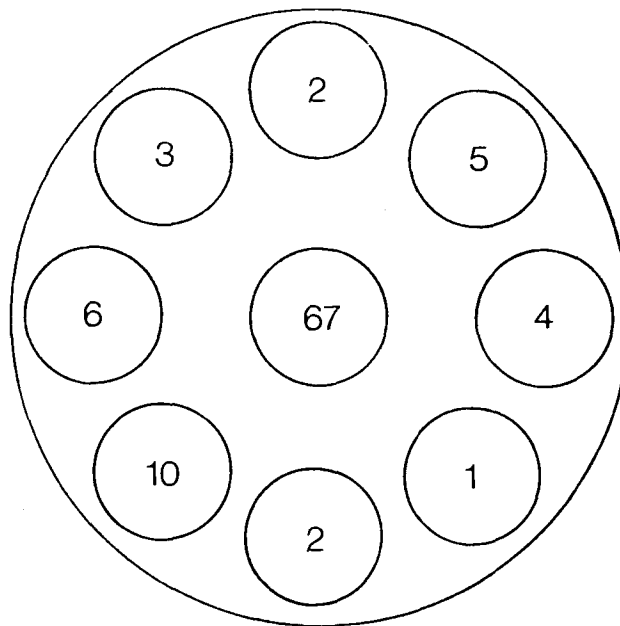
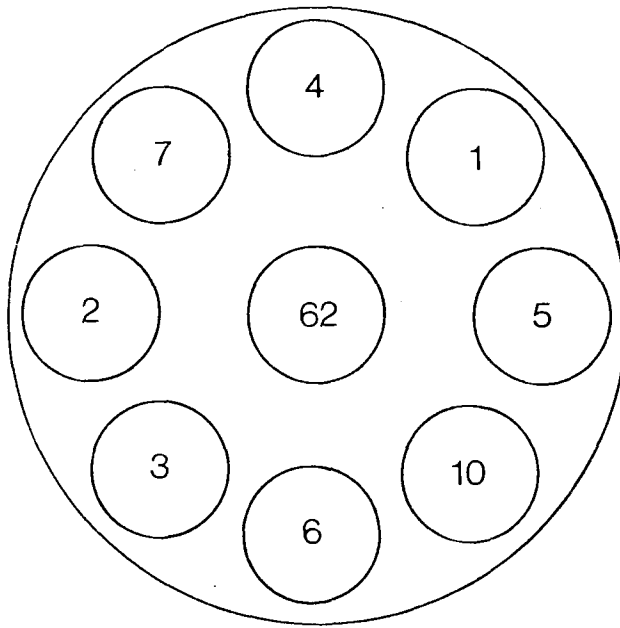
selective feeding by different nematodes on different food sources may reduce or perhaps even eliminate competition for a few chosen food sources. This would enable cohabitation of a small area, like algal blades, by a variety of nematode species which, on the basis of buccal morphology, would appear to feed on the same foods. This may be the case for M. disjuncta and M. refringens and the dominance of species types with the epigrowth feeding classification. Thus quantification or even precise definition of competitive interactions between marine nematodes in relation to preferred food sources in association with seaweeds should be pursued.

APPENDICES

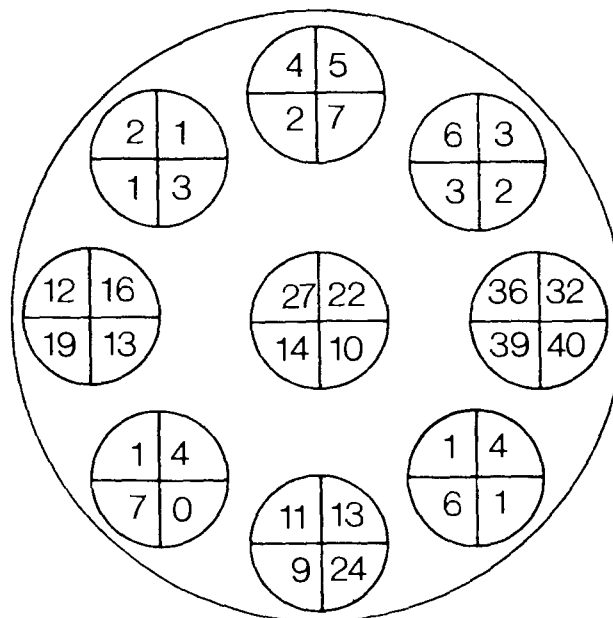
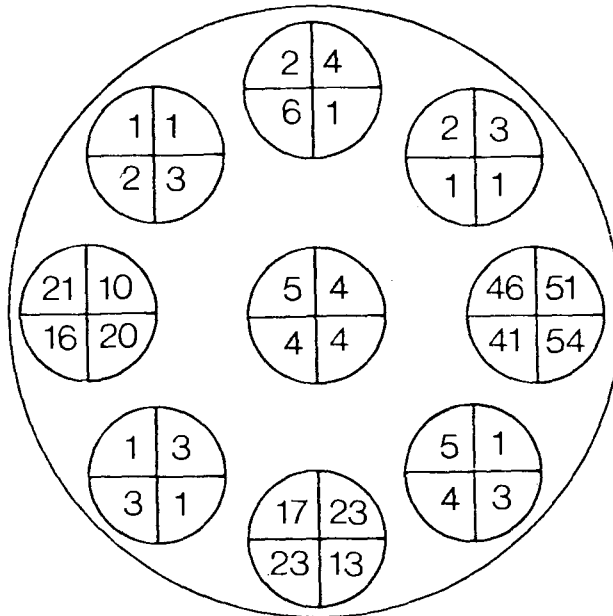
Appendix 1: Selective accumulation of Prochromadorella
neapolitana to alternating wells of diatom
species DI-D4 (see Table 1). Diagram records
four replicates for each condition.



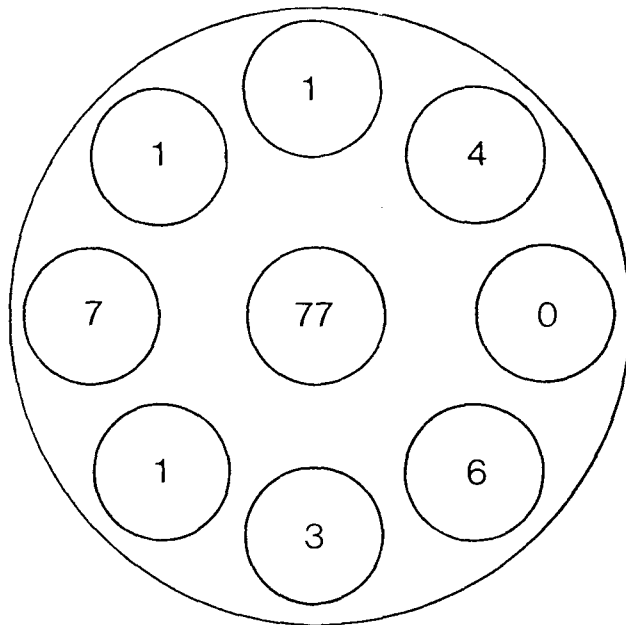
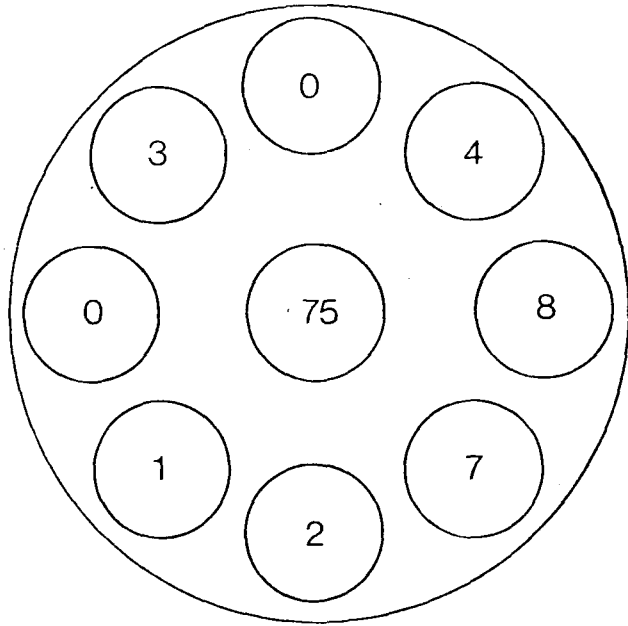
Appendix 2: Accumulation of Prochromadorella neapolitana
to control wells.



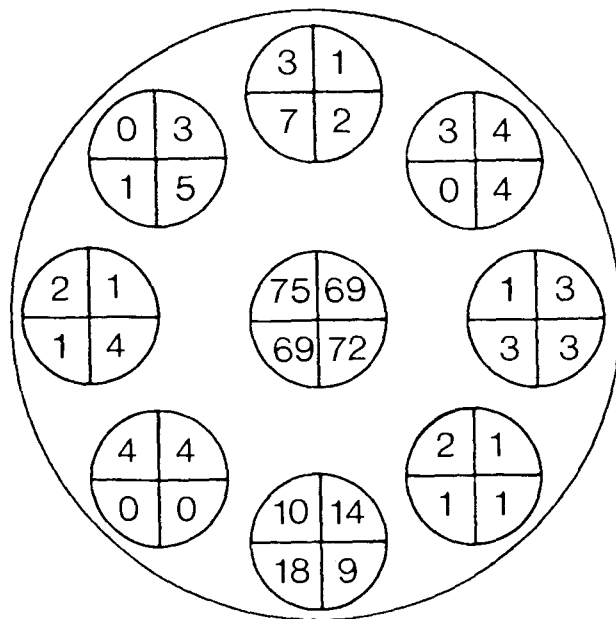
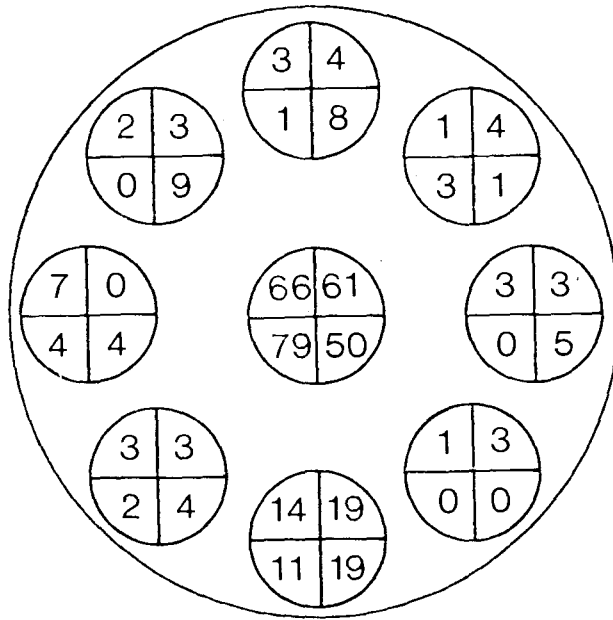
Appendix 3: Selective accumulation of Prochromadorella
neapolitana to alternating wells of diatom
species D5-D8 (see Table 1).



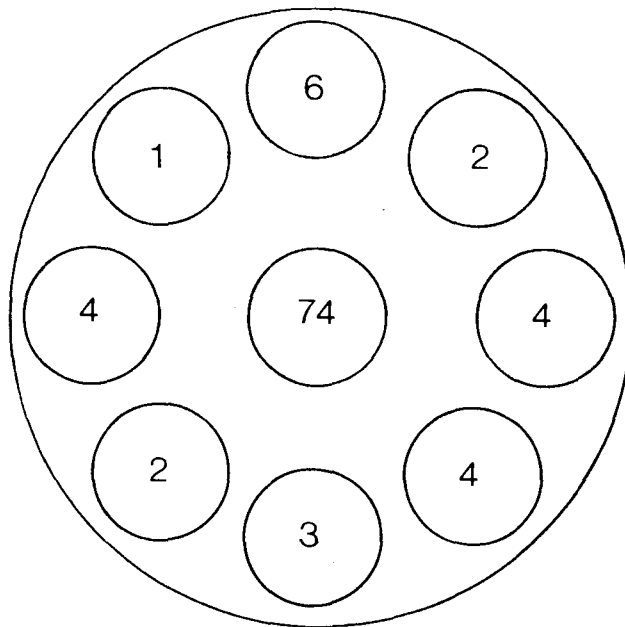
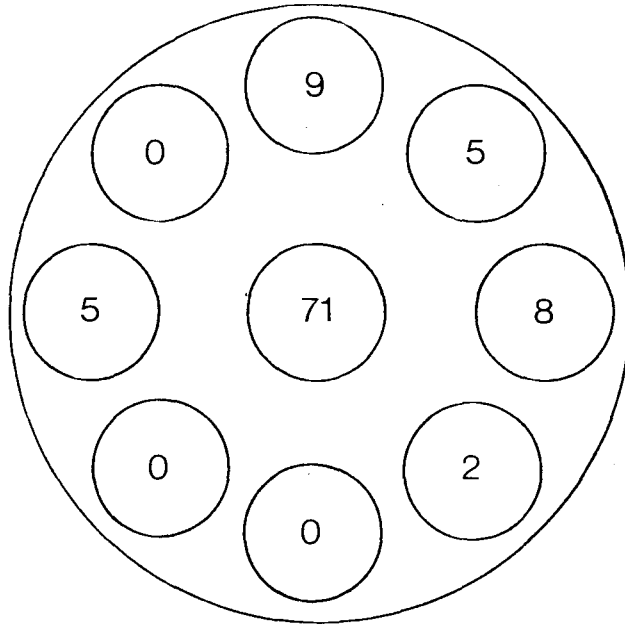
Appendix 4: Accumulation of Prochromadorella neapolitana to control wells.



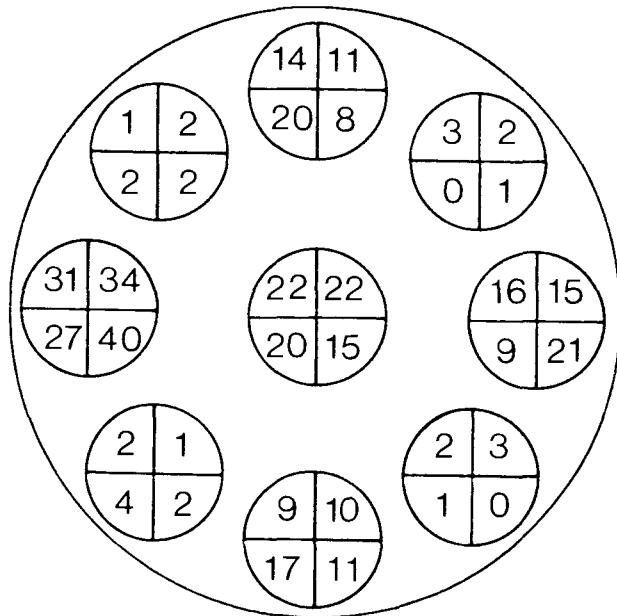
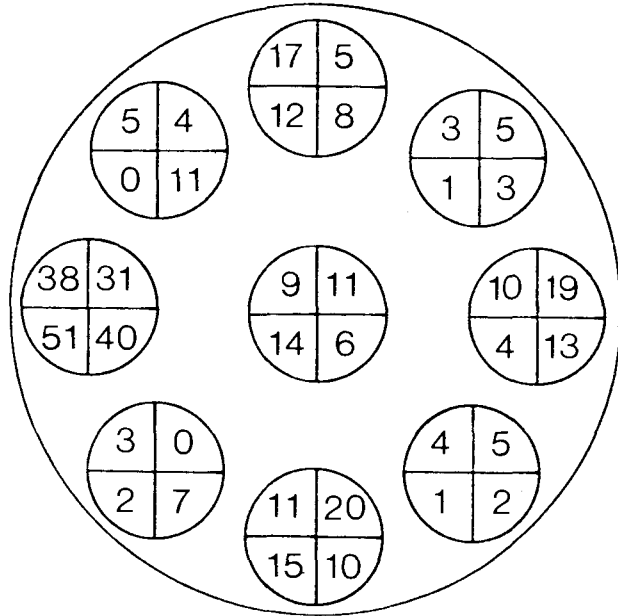
Appendix 5: Selective accumulation of Prochromadorella neapolitana to alternating wells of bacteria species B1-B4 (see Table 1). Diagram records four replicates for each condition.



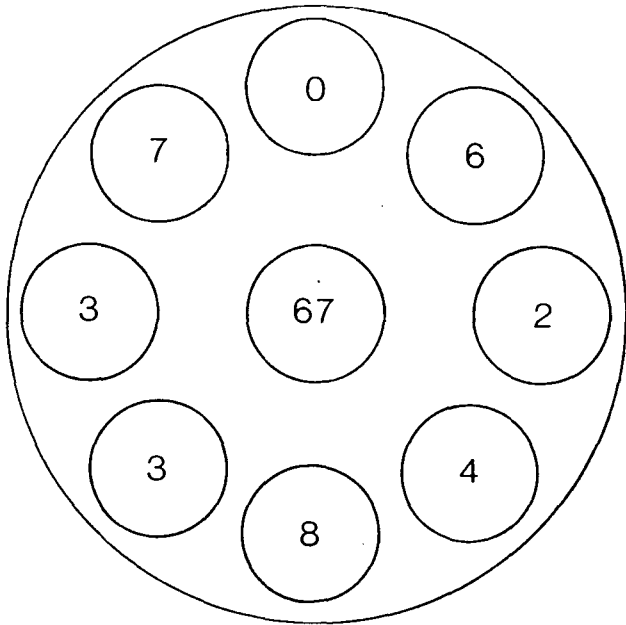
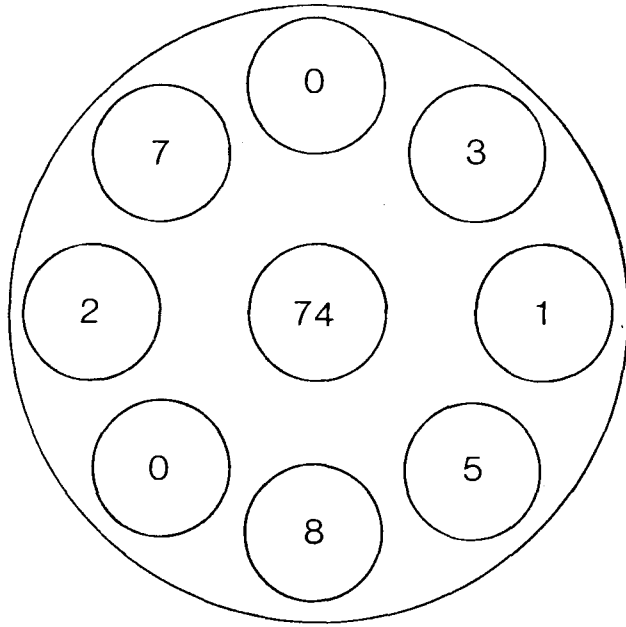
Appendix 6: Accumulation of Prochromadorella neapolitana
to control wells.



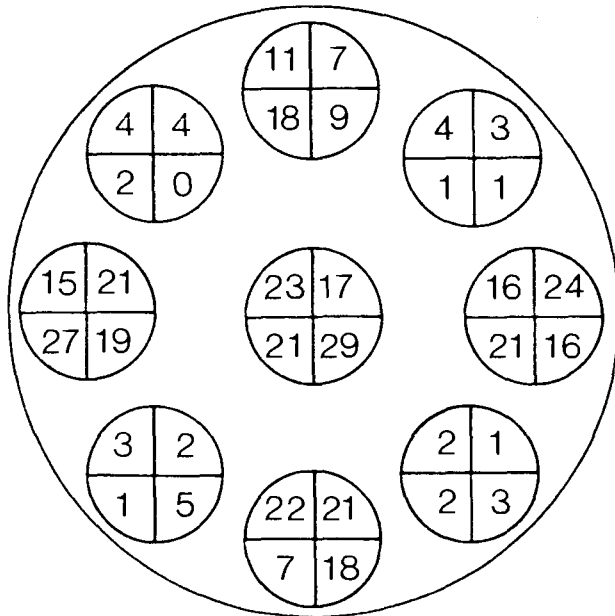
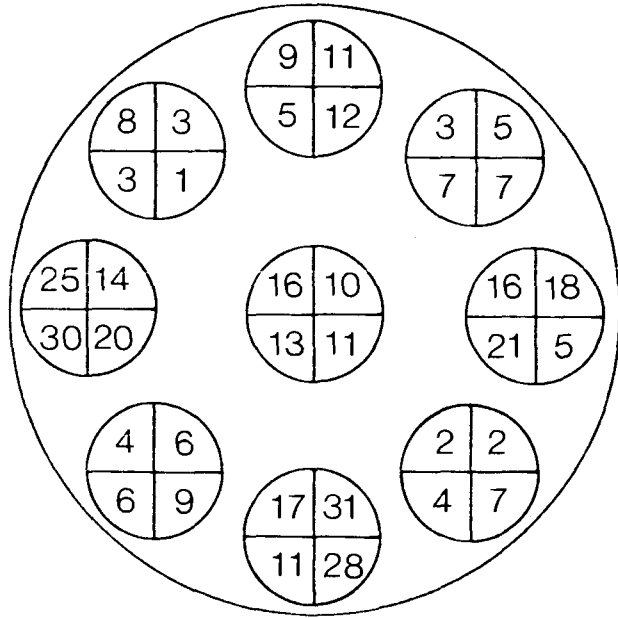
Appendix 7: Selective accumulation of Monhystera refringens to alternating wells of diatom species D1-D4 (see Table 1). Diagram records four replicates for each condition.



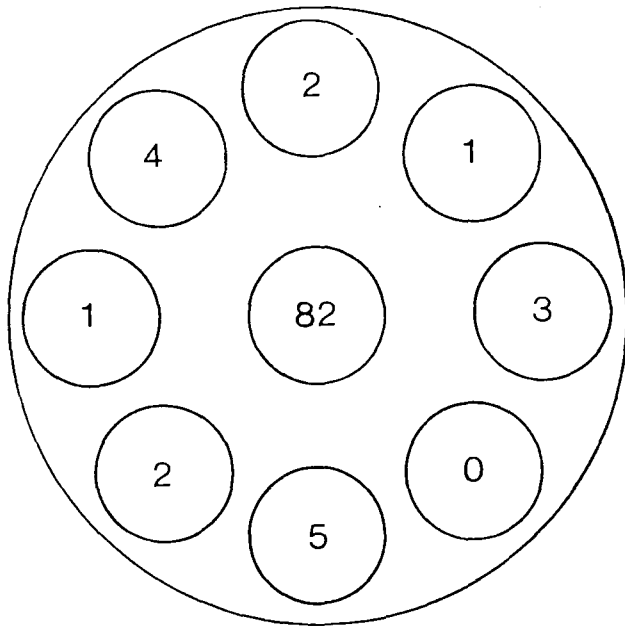
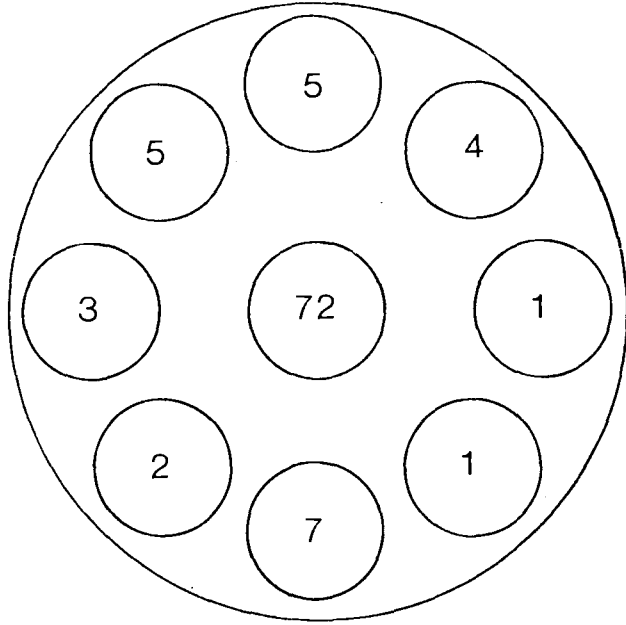
Appendix 8: Accumulation of Monhystera refringens to
control wells.



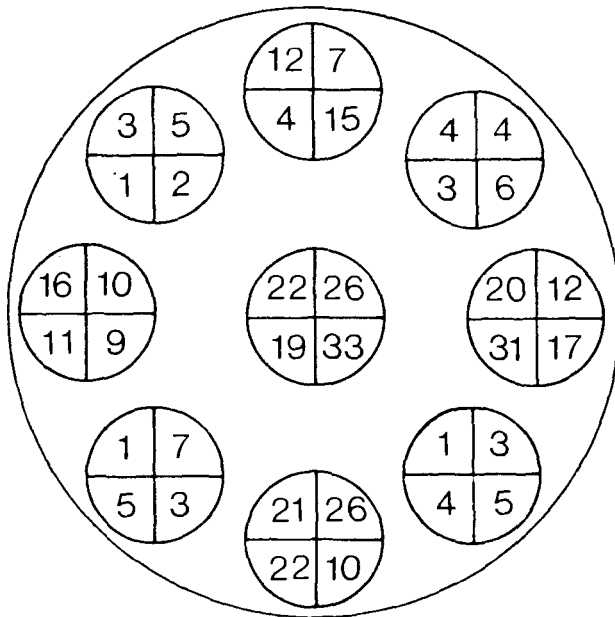
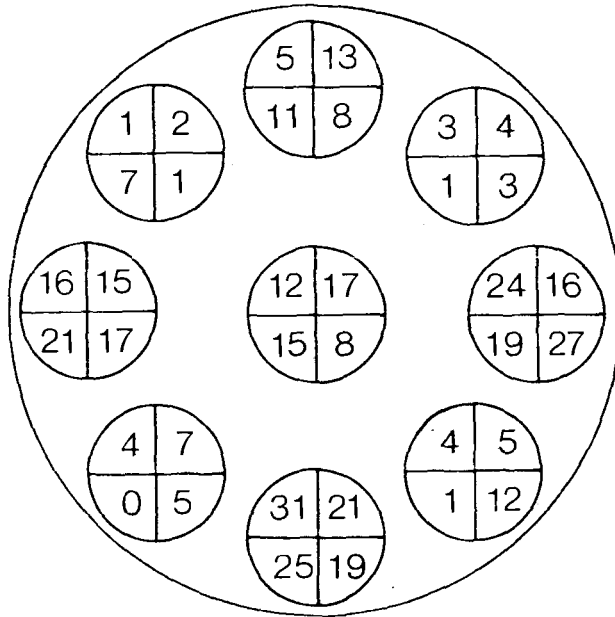
Appendix 9: Selective accumulation of Monhystera refringens to alternating wells of diatom species D5-D8 (see Table 1). Diagram records four replicates for each condition.



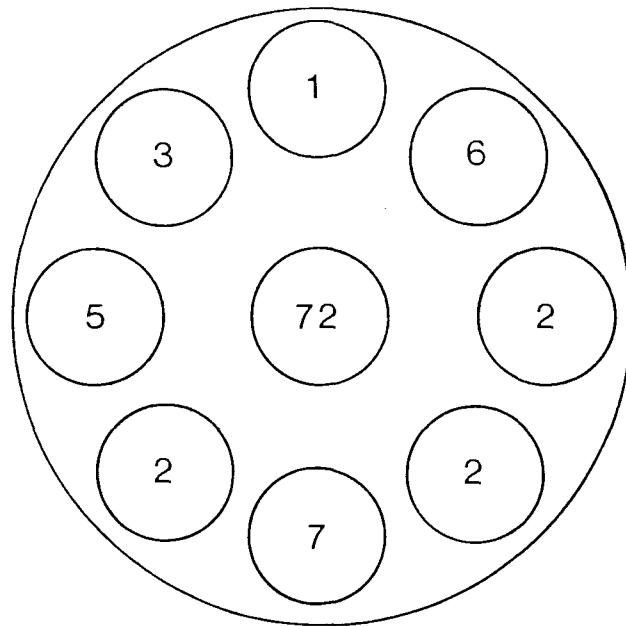
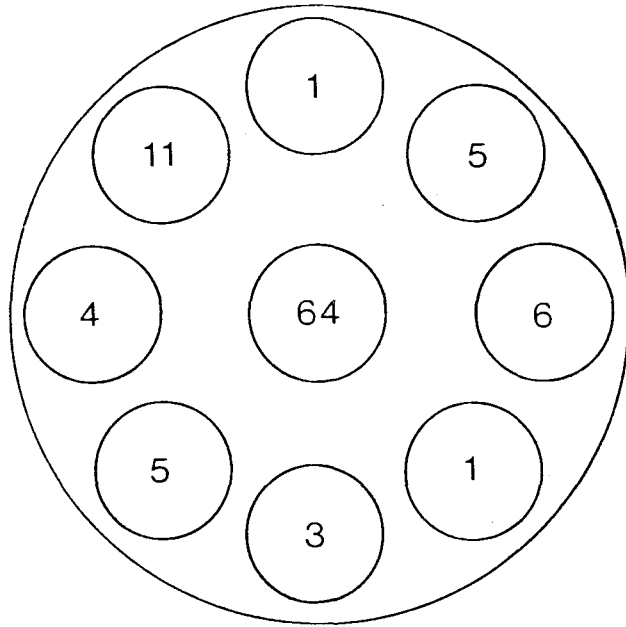
Appendix 10: Accumulation of Monhystra refringens to
control wells.



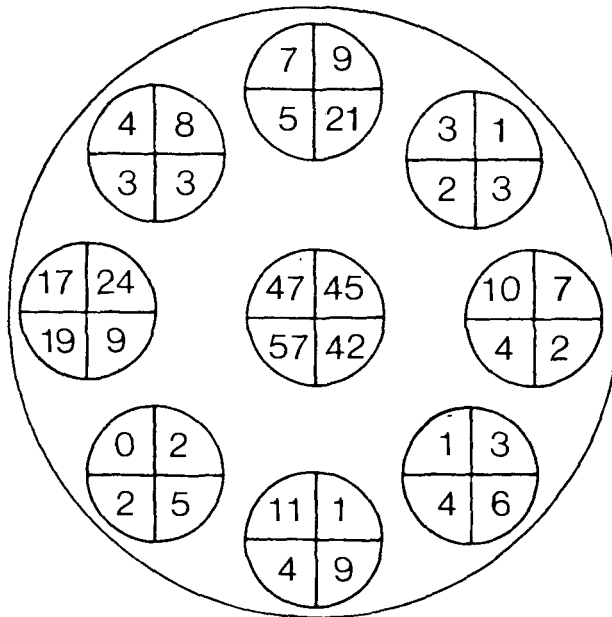
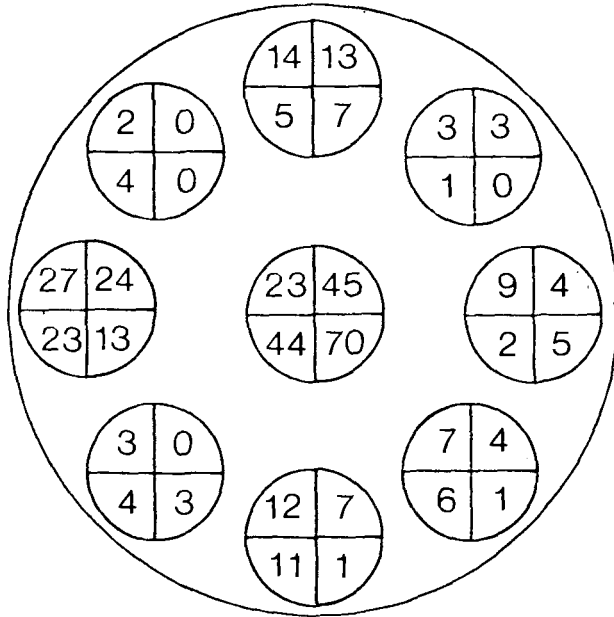
Appendix 11: Selective accumulation of Monhystera refringens to alternating wells of bacteria species B1-B4 (see Table 1). Diagram records four replicates for each condition.



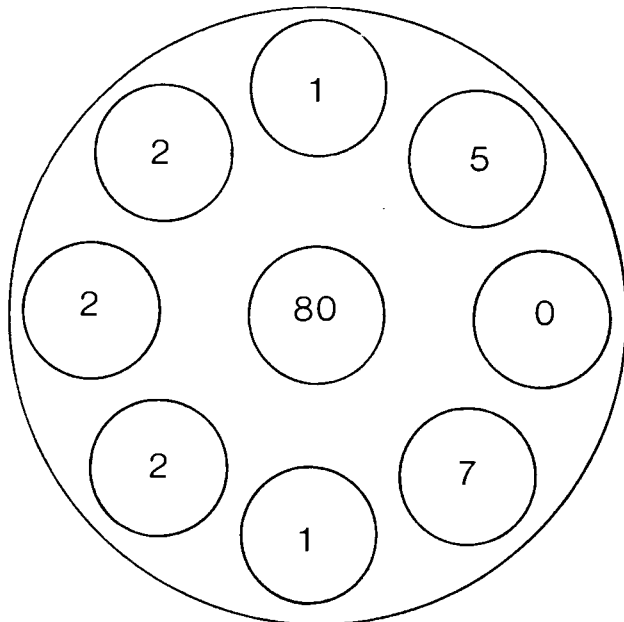
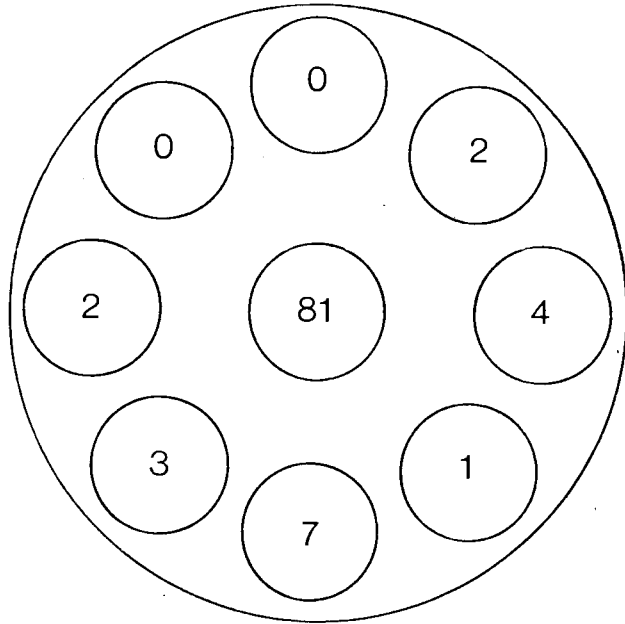
Appendix 12: Accumulation of Monhystera refringens to control wells.



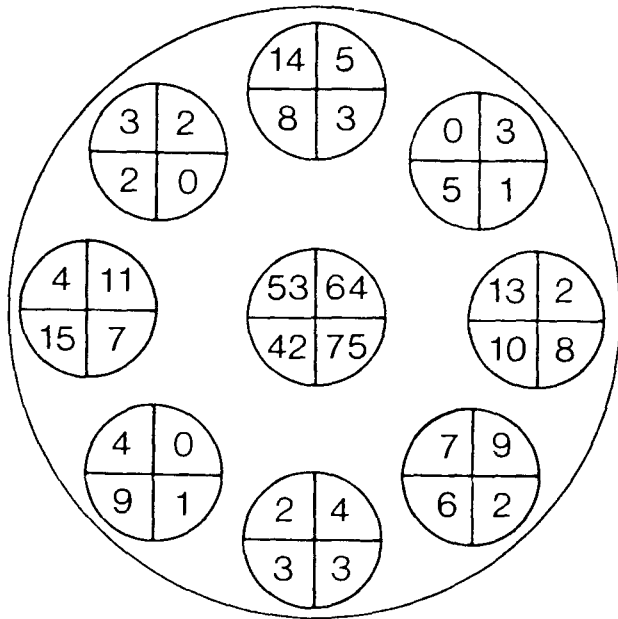
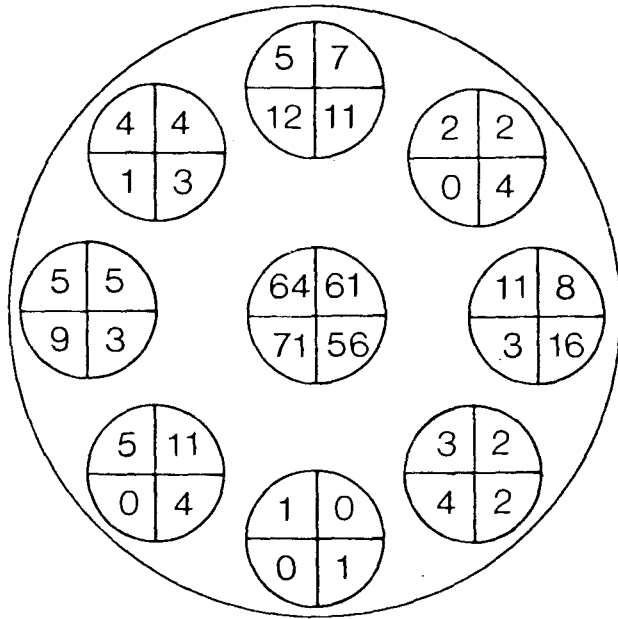
Appendix 13: Selective accumulation of Monhystera disjuncta
to alternating wells of diatom species D1-D4
(see Table 1). Diagram records four replicates
for each condition.



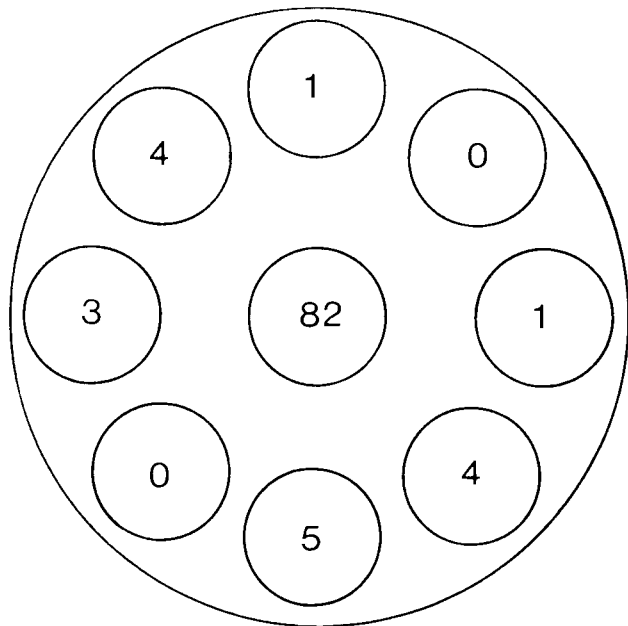
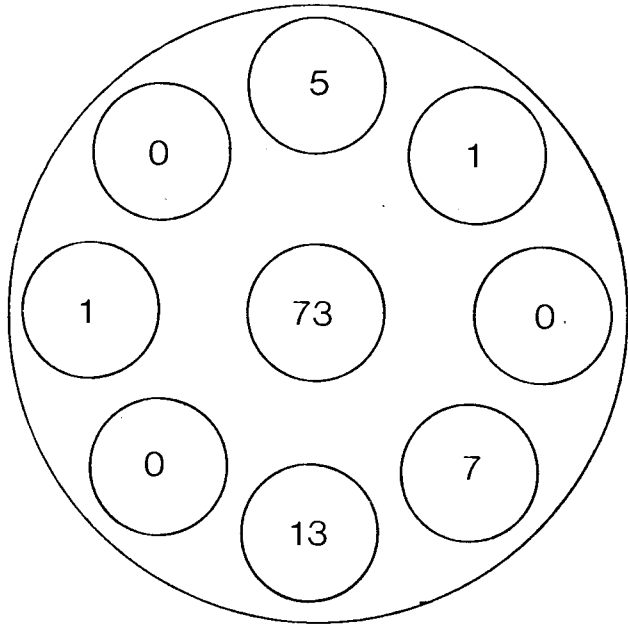
Appendix 14: Accumulation of Monhystera disjuncta to control wells.



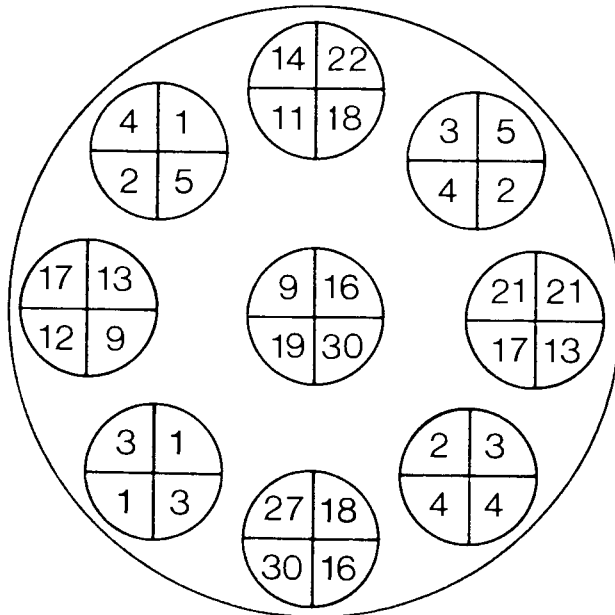
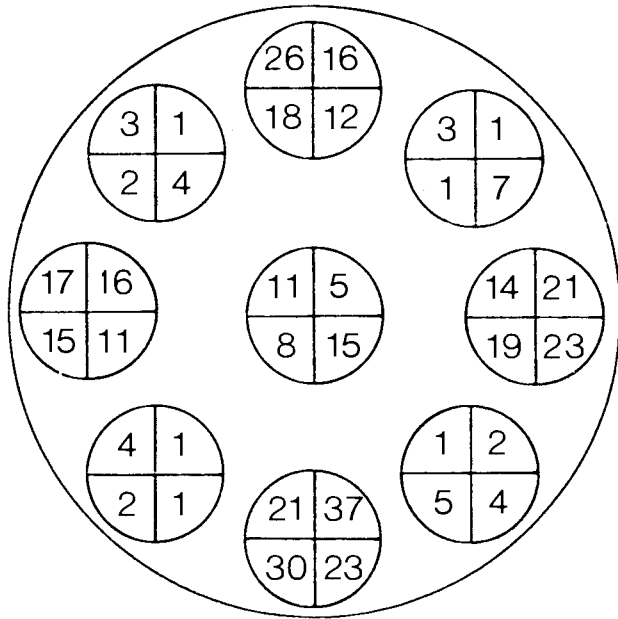
Appendix 15: Selective accumulation of Monhystera disjuncta
to alternating wells of diatom species D5-D8
(see Table 1). Diagram records four replicates
for each condition.



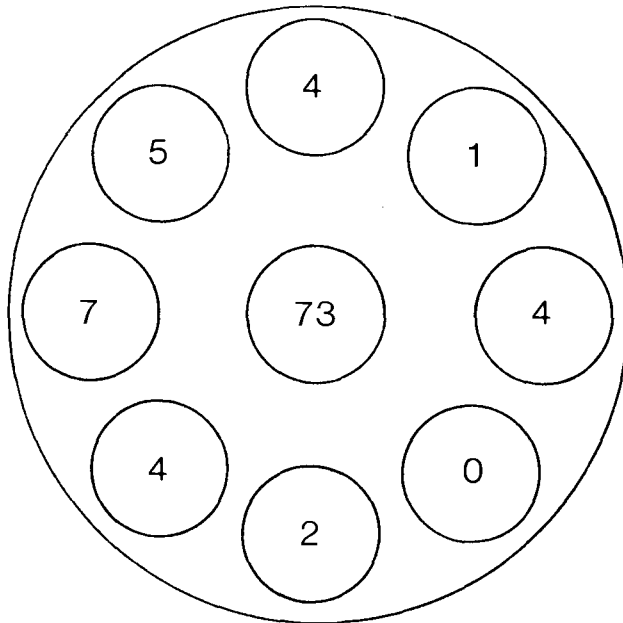
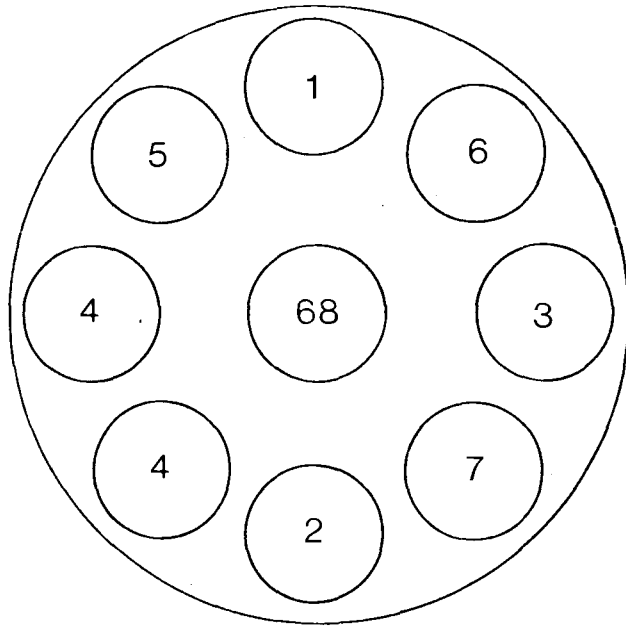
Appendix 16: Accumulation of Monhystera disjuncta to control wells.



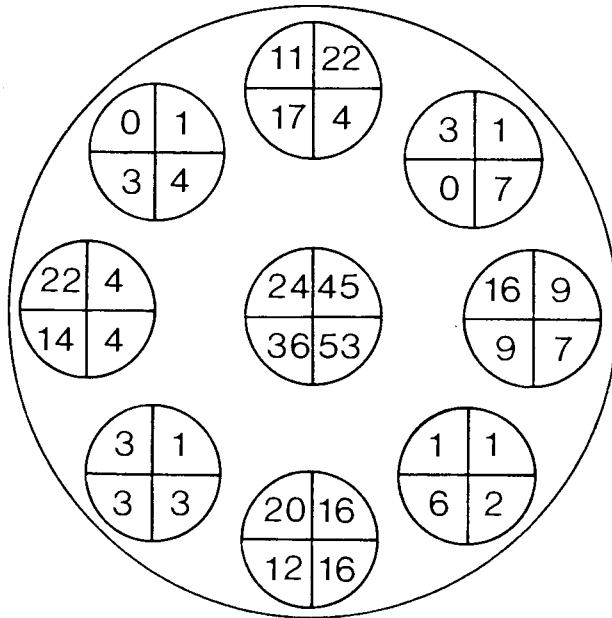
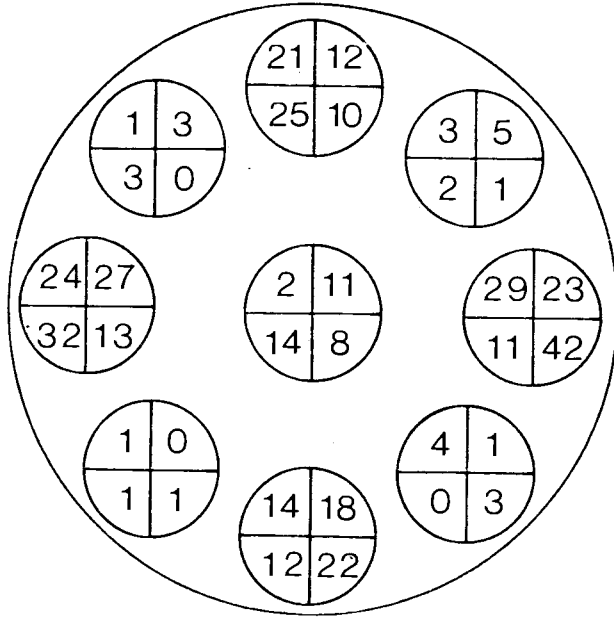
Appendix 17: Selective accumulation of Monhystera disjuncta to alternating wells of bacteria species BI-B4 (see Table 1). Diagram records four replicates for each condition.



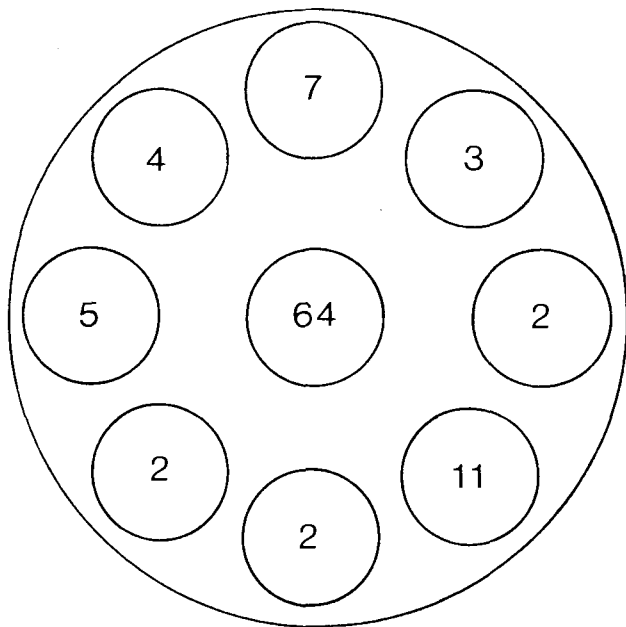
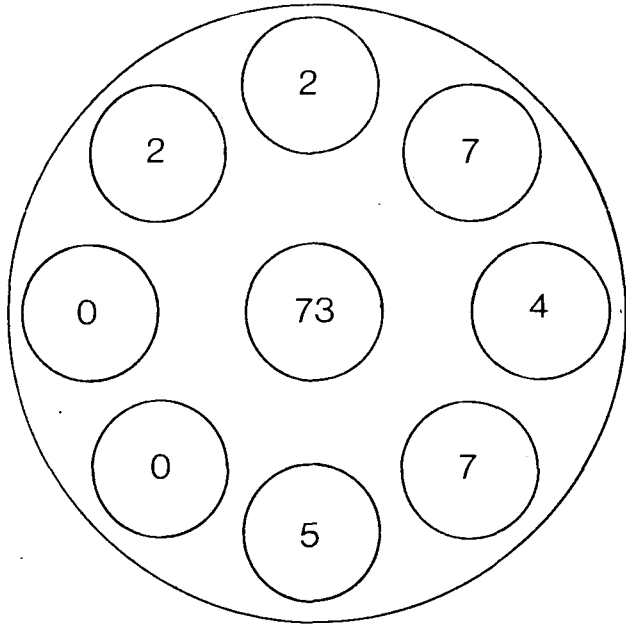
Appendix 18: Accumulation of Monhystera disjuncta to control wells.



Appendix 19: Selective accumulation of Monhystera disjuncta to alternating wells of (i) rod or (ii) coccoid bacteria species (see Table 1). Diagram records four replicates for each condition.



Appendix 20: Accumulation of Monhystera disjuncta to control wells.



Appendix 21: Two level ANOVA calculated on age structure (males, females, juveniles/ 0.5 m²) densities of Prochromadorella neapolitana.

| Source of Variance | Degrees of Freedom | Sum of Squares | Mean Square | F | Significance |
|--------------------|--------------------|----------------|-------------|-------|--------------|
| Age Structure | 2 | 1708.8 | 854.3 | 14.56 | *** |
| Time | 13 | 12615.3 | 970.4 | 16.54 | *** |
| Residual | 26 | 1525.2 | 58.7 | | |
| Total | 41 | 15849.3 | | | |

Appendix 22: Two level ANOVA calculated on age structure (males, females, juveniles/0.5 m²) densities of Monhystera refringens.

| Source of Variation | Degrees of Freedom | Sum of Squares | Mean Square | F | Significance |
|---------------------|--------------------|----------------|-------------|-------|--------------|
| Age Structure | 2 | 2032.0 | 1016.0 | 17.92 | *** |
| Time | 13 | 9777.4 | 752.1 | 13.27 | *** |
| Residual | 26 | 1473.9 | 56.7 | | |
| Total | 41 | 13283.6 | | | |

— ||| —

Appendix 23: Two level ANOVA calculated on age structure (males, females, and juveniles/0.5 m²) densities of Monhystera disjuncta.

| Source of Variation | Degrees of Freedom | Sum of Squares | Mean Square | F | Significance |
|---------------------|--------------------|----------------|-------------|-------|--------------|
| Age Structure | 2 | 12922.4 | 6461.2 | 30.05 | *** |
| Time | 13 | 5931.1 | 456.2 | 2.12 | N.S. |
| Residual | 26 | 5589.6 | 215.0 | | |
| Total | 41 | 24443.1 | | | |

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