EPIPHYTISM AND ENDOPHYTISM OF MACROCYSTIS INTEGRIFOLIA AND NEREOCYSTIS LUETKEANA: SEASONALITY, SUCCESSION AND TACTICS ON

TEMPORARY, LIVING SUBSTRATE

by

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in the Department

of

Biological Sciences

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Epiphytism and Endophytism of <u>Macrocys</u>tis integrifolia and Nereocystis luetkeana: seasonality, succession and tactics

on temporary living substrate

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ABSTRACT

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Epiphytism and endophytism of the stipes and blades of the large kelps Macrocystis integrifolia Bory and Nereocystis luetkeana (Mertens) Postels and Ruprecht were studied in a semi-exposed kelp bed in Bamfield Inlet, Vancouver Island, British Columbia. Major factors influencing epiphytism were seasonal immigration, specific features of the host, and the life spans of the blades and stipes. A spring increase in settlement of ectoprocts, hydroids, suctorians, and macroalgae was followed by summer or fall declines in species numbers and abundance. In contrast, diversity of diatoms on blades was lowest in spring and increased in fall and winter. Variations in densities of bacteria and endophytic algae were not seasonal. Growing tissue appeared to inhibit settlement and/or growth of organisms and host morphology effected positioning of organisms. The life span of the host tissue was important to the degree of development of organisms and assemblages of organisms. Abundant organisms on blades (life span of approximately 2 months) were restricted to bacteria, microalgae, and small animals. Only on stipes (life spans 6 - 14 months), did macroalgae become abundant.

Algae which were abundant had simple thalli with high surface to volume ratios, reproduced vegetatively on the host and had more rapid rates of development than less abundant seaweeds such as Fucales germlings and Laminariales which were solitary, complex, and failed to attain reproductive maturity within the host life span. Most abundant animals were small at maturity, reproduced by budding and produced disseminules within the life span of the host. Mortality mainly resulted from death of the host but also occurred from grazing (Druehl <u>et al</u>, 1977; Roland 1978). Density dependent mortality from competition for space occurred

seasonally and more rarely from overgrowth of certain organisms by others.

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TABLE OF CONTENTS

TABLE OF CONTENTS	Page
APPROVAL	II
ABSTRACT	111
ACKNOWLEDGEMENTS	v
LIST OF TABLES	viii
LIST OF FIGURES	i×
INTRODUCTION	· . 1
DESCRIPTION OF STUDY SITE AND STUDIED HOST SPECIES	2
METHODS AND MATERIALS	6
Sampling Procedure	6
Treatment of Samples	б
Statistical Treatment	7
Artificial Substrate	8
Settlement of Membranipora serrilamella	9
Taxonomic Notes	9
RESULTS	9
Phylum Ectoprocta	10
Phylum Cnidaria	12
Phylum Protozoa	12
Division Phaeophyta	13
Order Ectocarpales	, 13
Order Laminariales	14
Division Chlorophyta	14
Family Ulvaceae	14
Family Chaetophoraceae	15

·	Page
Division Rhodophyta	16
Diatoms on blades of Macrocystis and Nereocystis	32
Bacteria on blades of Macrocystis and Nereocystis	41
Epiphytism of Artificial Substrates	43
Glass Slides	43
Plastic "Plants"	43
DISCUSSION	44
Colonization	44
Host Effects	46
Host Life Span, Life Histories, and Succession	49
REFERENCES CITED	52

vIII

LIST OF TABLES

Number

L

Epiphytes and endophytes, except diatoms and bacteria, of the stipes and blades of Macrocystis and Nereocystis

- 2 Seasonal occurrence of diatom taxa on blades of Macrocystis
- 3 Seasonal occurrence of diatom taxa on blades of <u>Néréocystis</u>

35

Page

17

LIST OF FIGURES

Number		Page
: 1.	Locations of study sites	4
2.	Semi-diagrammatic sketches of <u>Macrocystis</u> integrifolia and <u>Nereocystis luetkeana</u>	. 5
3.	Colonization curves of stipes and blades of <u>Macrocystis</u> and <u>Nereocystis</u>	18
4.	Percent cover of the encrusting bryozoan, <u>Membranipora serrilamella</u> Osburn on blades of <u>Macrocystis</u> from June 1976 through December 1977	19
5.	Size class histograms of <u>Membranipora serrilamella</u> Osburn colonies on blades of <u>Macrocystis</u>	20
6.	Percent cover of the encrusting bryozoan, Membranipora serrilamella Osburn from February 1977 through December 1977	21
7.	Density of the hydroid, <u>Obelia</u> sp., on <u>Macrocystis</u> blades from June 1976 through December 1977	22
8.	Density of suctorians on blades of <u>Macrocystis</u> from June 1976 through December 1977	23
9.	Percent chance of occurrence per mm ² for Ectocarples on stipes of <u>Nereocystis</u> from February 1977 through December 1977	24
10.	Density of the endophyte <u>Streblonema</u> <u>aecidioides</u> f. <u>pacificum</u> Setchell and Gardner from July 1976 through December 1977 in blades of <u>Macrocystis</u>	25
11.	Percent chance of occurrence per mm ² of the endophyte <u>Bulbocoleon piliferum</u> Pringsheim	26
12.	Photographs of <u>Membranipora serrilamella</u> Osborn and <u>Obelia</u> sp	27
13.	Photographs of suctorians	28
14.	Photographs of epiphytised stipes of <u>Macrocystis</u> and <u>Nereocystis</u>	29

Number

Page

15.	Photographs of <u>Streblonema</u> <u>aecidioides</u> f. <u>pacificum</u> Setchell and Gardner in blades of <u>Macrocystis</u>	30
16.	Photographs of <u>Bulbocoleon piliferum</u> Pringsheim in blades of <u>Macrocystis</u>	31
17.	Seasonal variation in diatom densities on blades of Macrocystis	36
18. 19.	Seasonal variation in diveristy (H') and equidability (<u>H'</u>) of diatoms on blades of <u>Macrocystis</u> and Log ₂ S <u>Nereocystis</u> Dendrograms derived from cluster analysis of sampling dates for diatoms on blades of <u>Macrocystis</u> and Nereocystis	37 38
20, 21	Scanning electron photomicrographs of diatoms and bacteria on <u>Macrocystis</u>	39, 40
22.	Density of bacteria on blades of Macrocystis	42

×

INTRODUCTION

The life history traits of colonizers of unstable environments (Southwood, <u>et al</u>, 1974; Nichols, 1976), and the significance of seasonality and substrate stability to community structure in temperate marine waters have been theoretically and experimentally examined (e.g. Emerson and Zelder, 1978; Osman, 1977). Epiphytes (organisms attached to plants) of benthic marine plants in temperate latitudes live in a seasonally changing environment and, since host tissue is typically short lived, inhabit unstable substrates. However, an epiphytic ecosystem is peculiar in that the substrate may have evolved responses to colonizers (e.g. Sieburth, <u>et al</u>, 1974), and the often continual formation of new substrate through plant growth creates a habitat for colonization which is predictable and not dependent on perturbations for creation of new space.

Epiphytism of seaweeds and seagrasses has been studied descriptively and experimentally (e.g. Sloan <u>et al</u>, 1957; Harlin, 1973a,b, 1975; Boaden <u>et al</u>, 1976); however, I am not aware of any comprehensive study of an epiphytic ecosystem. Markham (1969) described the vertical distribution of seaweed epiphytes on the stipes of <u>Nereocystis luetkeana</u> (Mertens) Postels and Reprecht near San Juan Island, Washington, but epiphytism has not been described for plants on the west coast of British Columbia. Except for brief mention (Scagel, 1947; Tokida, 1960; Lobban, 1976) epiphytism of <u>Macrocystis integrifolia</u> Bory has not been studied. I have described the seasonal and within plant distributions of epiphytes and endophytes of the stipes and blades of the large kelps <u>N. luetkeana</u> and <u>M. integrifolia</u>. Succession of epiphytic and endophytic organisms was followed. Life history and natural history characters of of the epiphytes and endophytes were noted. Artificial substrates were used to compare the seasonality and succession of biotic with aboitic substrates.

Description of Study Site and Studied Host Species

The main study site was a kelp bed in Bamfield Inlet on Vancouver Island, British Comumbia (Fig. 1). <u>Macrocystis</u>, the most abundant of the two species, formed a fringing bed close to shore. <u>Nereocystis</u> was scattered throughout the bed of <u>Macrocystis</u> and along the outer perimeter. The depth range of the two species was from 0 to -4m below 0 tide. Monthly variations in temperature, salinity, and solar radiation throughout the period of this study have been reported by Lindsay and Saunders (1977, 1978). Surface water temperatures were 5 - 10°C in winter and over 15°C in summer. Salinities were between 25 and 28°/oo in winter and around 30°/oo in summer. Solar radiation in winter was less than 100 gcal/cm²/day and over 400 gcal/cm²/day in summer. In summer the kelp bed was fairly sheltered from waves but was semi-exposed in winter due to changing prevailing winds. Mean higher high water and mean lower low water were 3.3 and 0.9 m respectively above 0 tide.

These kelps were unique for a study of epiphytism due to their large size (length of mature stipes 3-4 m) and pattern of growth (Fig. 2). The main intercalary meristematic region is near the apex of the frond (stipe plus blades) of <u>Macrocystis</u> and at the junction of the stipe and blades of <u>Nereocystis</u>. These meristems continually produce new tissue, although the rate varies seasonally (Lobban, 1978; Foreman, R.E., Dept. of Botany, University of British Columbia). The holdfast of Macrocystis is perrennial and new fronds are produced throughout the year. The stipe, which bears the blades, is present approximately 6 months and any blade lives about 2 months (Lobban, 1978). The Nereocystis sporophyte is an annual which begins growth in March and April, ceases growth by late summer and dies during spring and early summer of the following year. The stipe is present 12-14 months and any piece of blade tissue probably remains about 2 months. The blades of Macrocystis are covered with furrows and ridges and blades of Nereocystis are smooth. Because new tissue is added at the meristems with progressively older tissue found distal to the meristems, certain successive events can be followed without the passing of time. Each species has tissues of widely varying ages throughout all seasons, allowing the importance of the state of tissue maturity to be examined. In this study I recognized three qualitative ages or states of tissue maturity: growing tissue, mature tissue (not growing but healthy) and senescent tissue (decaying).

Figure I. Locations of study sites.

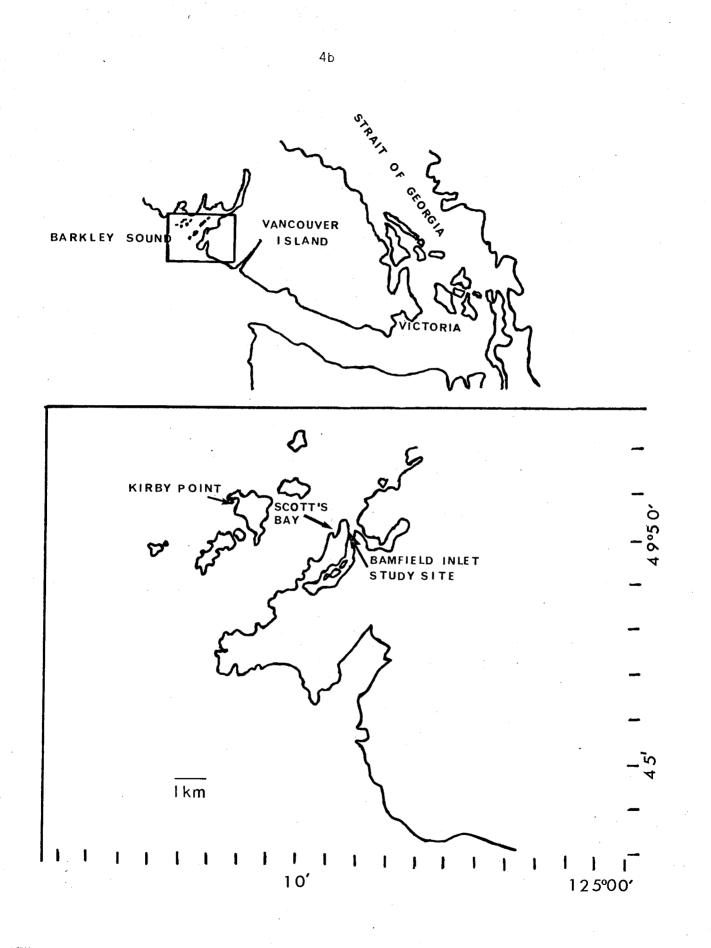
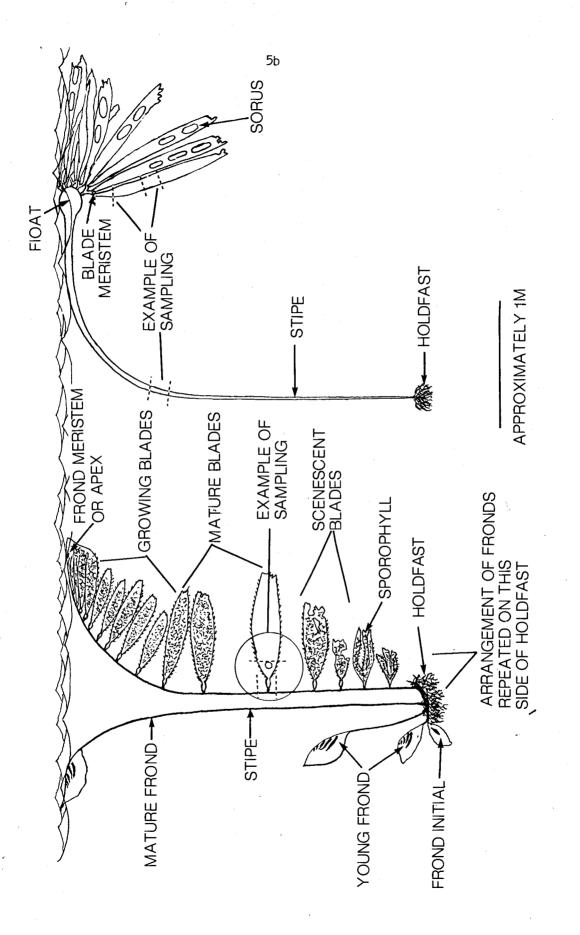


Figure 2. Semi-diagrammatic sketches of <u>Macrocystis</u> integrifolia (left) and <u>Nereocystis</u> luetkeana (right).



METHODS AND MATERIALS

6

Sampling Procedure

Plants of <u>Macrocystis</u> and <u>Nereocystis</u> were randomly sampled each 4-7 weeks from July 1976 to December 1977. Fronds of <u>Macrocystis</u> (n=10-18) were subsampled at 0.1 m below the frond meristem, then at 0.5 m intervals to the frond base. At each interval, 10 cm of the stipe and the first 10 cm of the blade from the float were sampled (Fig. 2). For low magnification work and diatom study by acid clearing, 3.5 cm diameter subsamples were taken 10 cm from the float. For scanning electron microscopy (SEM), five 1 cm diameter subsamples, cut 10 cm from the float, were taken at 0.1, 1.0, and 2.0 m from the frond meristem.

Blades of <u>Nereocystis</u> (n=5) were subsampled 5-15 and 45 - 55 cm from the float (Fig. 2). Stipes (n=2-6) were sampled at each 0.5 m interval from the float from February to December 1977. Each sample was 10 cm long.

Taxa missed due to sampling error were calculated with a species area curve, where area was the number of stipes or blades. Virtually all sample sizes taken were well into the asymptote of the curves.

Epiphytes and endophytes were occasionally observed on <u>Macrocystis</u> and <u>Nereocystis</u> in other areas, over a range of wave exposures.

Treatment of Samples

Plant tissue was preserved in 10% formalin in seawater. The preserved tissue was examined for epiphytes and endophytes at three levels of magnification. I. Epiphytes clearly discerned with the unaided eye (e.g. bryozoans'and macro-algae) were quantified on the entire piece of tissue sampled. 2. Epiphytes requiring low (X45) magnification (e.g. suctorians and hydroids) were quantified with a dissecting microscope by counting organisms occurring in 100 randomly chosen 1 mm² quadrats on the tissue sampled. 3. Epiphytes requiring high (500-10,000 X) magnification (e.g. diatoms and bacteria) were examined with light or scanning electron (SEM) microscopy.

Tissue for SEM observations was preserved in 5% glutaraldehyde in 1:1 sea water and fresh water. Tissue was refrigerated up to two months. then dehydrated in a series of acetone washings, critical point dried with CO2, mounted onto stubs with silver paint, gold coated by vacuum diffusion (NRC 3115) and viewed with the SEM (ETEC Corp.). Diatoms were counted and identified in 10 random quadrats at 900X (0.81 mm²). Epiphytic diatoms were also examined by dissolving samples of plant tissue, after removal of large epiphytes, with 1:1 sulfuric and nitric acids. The resultant suspension was allowed to settle for 24 hr, the supernatant poured off, and the settled diatoms diluted with distilled water. This was repeated several times to dilute the acid to a low concentration. Measured amounts of the final volume of suspended diatoms were spread evenly onto coverslips, dried and mounted in Hyrax. Random transects on 3 slides were counted to quantify the number of diatoms per unit area of host tissue. Typically 500 diatoms were counted in each sample, as this number was always into the asymptote of species versus sample size curves (Saunders, 1968) indicating that a large change in sample size added very few species.

Statistical Treatment

Diversity of diatoms sampled was estimated by the information measure (H!) (Shannon and Weaver, 1949) where n_i is the number of

individuals in the ith species, N is the total number of individuals in the sample, and s is the number of species in the sample.

$$H' = -\sum_{i=1}^{n} \frac{n_i}{(N-1)} \log_2 (\frac{n_i}{N})$$

Evenness of distribution of individuals among the species was calculated as $\underline{H'}_{Log_2 \ s}$ (Pielou, 1975). This measure aids in interpreting the calculated diversity index since a community with a few evenly distributed species can have the same H' as one with many unevenly distributed species (Pielou, 1975).

The niche breadth of diatom taxa were calculated by the expression

$$B_{i} = \exp \left[\sum_{r=1}^{n} \left(\frac{n_{ir}}{N_{i}}\right) \log_{e} \left(\frac{n_{ir}}{N_{i}}\right)\right]$$

where n_{ir} is the number of individuals of the ith taxon in the rth sample in all Q samples (Levins, 1968; McIntire and Overton, 1971; Main and McIntire, 1974). This measures the evenness of distribution of a taxon throughout the samples considered (Main and McIntire, 1974).

Cluster analysis of cases (Biomedical Program series at Simon Fraser University computing center) was used to cluster sampling dates of diatoms. The chi-square test of equality defined the distance between data of any two cases and was used to form clusters.

Artificial Substrate

"Plants" composed of plastic 5 cm x 50 cm strips tied at 0.5 m intervals to tygon tubing, and of open boxes tied to a rope at 0.5 m intervals, containing glass slides held in place with elastic bands, were suspended in the kelp bed. Glass slides were replaced at 1-2 month intervals from May 1976 to April 1977, and the plastic was sampled without replacement each 1-2 months from May 1976 to June 1977.

Settlement of Membranipora serrilamella

9

Larvae and colonies in small size classes were counted on proximal (youngest) and distal (oldest) portions of growing and senescent blades (5x15 cm samples of blades, n = 3 - 5). Assuming that growth of colonies on young and old tissue was similar, areas of preferred settlement could be determined.

Taxonomic Notes

Invertebrates were identified using Light (1975) and Kozloff (1975). Suctorians were identified using Kudo (1971) and Ricketts and Calvin (1968). Seaweeds were identified in Setchell and Gardner (1920, 1925), and Widdowson (1974). Dr. R.F. Scagel confirmed the identification of <u>Streblonema aecidoides f. pacificum</u>. Dr. C. Tanner identified species of <u>Ulva</u>. Diatoms were identified by using van Heurk (1962) and Rao and Lewin (1976), and certain identifications were confirmed by Drs. J. Lewin and M. Taylor. Dr. M. Hawkes identified the <u>Porphyras</u>.

RESULTS

All epiphytes and endophytes, except diatoms and bacteria, found through sampling or by casual observation on the stipes and blades of <u>Macrocystis</u> and <u>Nereocystis</u> are listed in Table1; Tables 2 and 3 list diatoms found on blades of <u>Macrocystis</u> and <u>Nereocystis</u> respectively. Colonization curves of epiphytes and endophytes recorded through the sampling procedure (not casual occurrences) of the stipes and blades of the two host kelps are in Fig. 3, and diversity indices (H') of diatoms on the blades of both hosts are in Fig. 18. Colonization curves show most immigration during June, July and August for blades of both host species with more taxa on blades of <u>Macrocystis</u>, but diversity of diatoms was lowest in early spring and summer on blades of both plants. Mortality of organisms was mainly due to death of the host, and post summer immigration of organisms, except for diatoms, was low. The result is seen as a decrease in the number of epiphytic taxa after summer in Fig. 3.

Stipes of <u>Nereocystis</u> along the perimeter of the bed were epiphytised only by diatoms and bacteria until August when host growth ceased. From that point on several additional epiphytes were present through December; in the spring additional species settled. None of these epiphytes became extinct until death of the plants. <u>Nereocystis</u> inside the bed of <u>Macrocystis</u> became senescent by June and were heavily epiphytised until their death in August (Table 1). Stipes of <u>Macrocystis</u> had one epiphyte other than diatoms and bacteria during the summer months but occasionally a senescent stipe was discovered which bore several large epiphytes (Table 1).

Phylum Ectoprocta

<u>Membranipora serrilamella</u> settled on the stipes and blades of both hosts during summer but persisted through winter only on stipes of <u>Nereocystis</u>. <u>Hippothoa hyalina</u> occurred rarely on blades of <u>Macrocystis</u> and stipes of <u>Nereocystis</u>, and <u>Tricellaria occidentalis</u> occurred rarely on blades of <u>Macrocystis</u>.

<u>Membranipora serrilamella</u> was present on the blades of <u>Macrocystis</u> from May to August 1976 and in June and August 1977 (Fig. 4); stipes were more rarely encrusted at these times. Larvae settled on the newest tissue but blades less than 1.5 m from the frond meristem rarely had more than 5% cover. Heaviest coverages occurred during May and June 1976 when most blades 2.5 and 3.0 m from the apex were 25 to 100% encrusted

(Fig. 12a). By August 1976 the older zooids of colonies were gone, leaving only an outer perimeter of young zooids (Fig. 12b). Two encrustations occurred in 1977, a small one in June and one much larger in August; however, neither was the magnitude which occurred in 1976. In 1976, abundance was significantly correlated with distance of the blade from the apex, but in 1977 the correlation coefficients were not significant. Median densities closely paralleled the means but variance to mean ratios were high indicating that coverage was clumped for blades of similar distance from the apex within the bed.

The larvae of <u>Membranipora</u> settled only in the furrows of the blades of <u>Macrocystis</u>, but would settle and grow on the relatively smooth stipe. Larvae appeared to settle indiscriminately on young and older blades. The frequency of colonies in the same small size classes were distributed fairly equally on the proximal (growing) and distal (not growing) ends of growing blades (about 0.6 m from the meristem), and fairly equally apportioned between growing and nearly senescent blades about 1.6 m from the frond meristem (Fig. 5). During spring settlement 1 have observed dense numbers of larvae on the youngest and oldest blades of <u>Macrocystis</u>. Colonies grew by radiating from the larvae, encrusting the blade surface. Eventually they would grow over the edge of the blade, covering the spinules along the edge of the blades and continue grow n on the other side of the blade. Colonies were never observed to overgrow one another.

From June to August 1976 <u>Membranipora serrilamella</u> heavily encrusted the stipe of <u>Nereocystis</u> and persisted until February 1977 when quantitative measurements were initiated (Fig. 6). Senescent plants in February and April 1977 had encrustations of less than 25% cover in the

upper 0.5 m of stipe. <u>Membranipora</u> was absent from the stipes of plants which began growing in spring 1977 until October 1977. Colonies occurred from the holdfast to the blades but highest percent covers were between the float and 0.5 m of stipe. Blades were only rarely epiphytised by this bryozoan and only during summer.

Phylum Cnidaria

<u>Obelia</u> sp. (Hydrozoa) was common on blades of <u>Macrocystis</u> but occurred rarely on stipes of <u>Macrocystis</u>. One specimen of the anemone Metridium sp. occurred on a Macrocystis stipe.

<u>Obelia</u> colonies were present on blades of <u>Macrocystis</u> from June 1976 to October 1976 and from June 1977 to October 1977 (Fig. 7). Maximum densities exceeding 0.30 colonies/mm², occurred during the summer of both years, on blades Im or more from the apex (Fig. 12c,d). Mean densities on the plants ranged from around 0.025 to 0.10 colonies/mm². Colonies were never observed at 0.1 m and only rarely at 0.5 m from the apex. Abundance was positively correlated with distance from the apex at 5% confidence level for 7 of 8 months. Median densities were nearly always less than the mean, and variance to mean ratios were much greater than one indicating a clumped distribution within the bed on similarly positioned blades. Colonies of <u>Obelia</u> 3 to 4 cm in length could possess gonozooids. Densities could be reduced from overgrowth by Membranipora.

Phylum Protozoa

Suctorians (<u>Acineta</u> sp. and <u>Ephelota</u> sp.) were found on blades of both hosts. <u>Folliculina</u> sp. <u>Vorticella</u> sp. and <u>Zoothamnion</u> sp. rarely occurred on blades of <u>Macrocystis</u>.

Suctorians were present on blades of <u>Macrocystis</u> from June 1976 to December 1977 (Fig. 8, 15). Maximum mean densities of $2 - 4/mm^2$ occurred on blades I or 2 m from the frond meristem during June, 1976 to April, 1977. At these times they formed a visible layer on the blades. During the other months of both years the densities were visibly lower and typically less than $1/mm^2$. Abundance was not significantly correlated with distance from the apex but maximum densities were never observed on growing tissue. Variance to mean ratios were high, indicating a clumped distribution within the bed on similarly positioned blades. Distribution on the blades was clumped as they were present only in the furrows.

Suctorians were present on blades of <u>Nereocystis</u> during June and July of both years but densities never exceded 0.25/mm² on blade tissue 0.5 m from the float. They were never observed on the stipes. Densities of suctorians on blades of <u>Macrocystis</u> and <u>Nereocystis</u> could be reduced from overgrowth by <u>Membranipora</u> (Fig. 13a).

Division Phaeophyta

Members of the Order Ectocarpales were present on the stipes and blades of both hosts. Members of the Order Laminariales were found in spring on stipes of both species, and germlings of the Order Fucales occurred on blades of Macrocystis during summer.

Order Ectocarpales

Ectocarpales were present throughout the year on senescent blade tissue of <u>Nereocystis</u>, but did not appear on the stipes of plants which began growth early in the spring until August. They totally covered the stipe by December (Fig. 9, 14c). Distribution was not significantly correlated with distance from the float. <u>Nereocystis</u> growing inside the bed of <u>Macrocystis</u> developed heavy mats by July. Epiphytic Ectocarpales

occurred only on stipes of Macrocystis which were aging.

The brown algal endophyte, <u>Streblonema aecidioides</u> f. pacificum, was present in the blades of <u>Macrocystis</u> from July 1976 through December 1977 with the exception of August 1977, when none were found among the 11 sampled fronds (Fig. 10). Maximum density of eruptions of the thallus through the epidermis (Fig. 16a, b, c, d) appeared to occur during the summer with mean densities reaching I - 2/mm². Mean abundance of eruptions tended to be positively correlated with distance from the frond meristem, 5 to 10 linear correlation coefficients being significant at the 5% confidence level. No eruptions were seen on blades 0.1 and 0.5 m from the apex. Young unerupted thalli were occasionally seen by sectioning blades close to the apex (Fig. 11c). Median densities were typically less than the mean and variance to mean ratios were greater than one, indicating a clumped distribution within the bed on similarly positioned blades.

Order Laminariales

<u>Alaria</u> sp. occurred on stipes of <u>Nereocystis</u> in April and persisted until death of the host in densities of approximately 0.1/cm². During summer 1976 a senescent stipe of <u>Macrocystis</u> was found with <u>Laminaria</u> <u>groenlandica</u> (Fig. 14a). None of these epiphytic kelps were reproductive. Division_Chlorophyta

Family Ulvaceae

<u>Nereocystis</u> outside the bed of <u>Macrocystis</u> which began growing in the spring of 1977 had <u>Ulva</u> on the float by August and through October and December the distribution remained restricted to the float. In February, 1977 the previous generation of plants had <u>Ulva</u> present on the upper 0.5 m

of stipe (approx. 20/cm²) most of which were less than 0.5 cm in length. By April 1977 <u>Ulva</u> occurred as low as 1.5 m from the float on these host plants and were present in lesser densities of approximately 5/cm²; many of the plants were 5-10 cm in length. All <u>Ulva</u> large enough to be identified were U. stenophylla or U. fenestrata.

<u>Enteromorpha linza</u> occurred on the upper 0.5 m of the stipe of <u>Nereocystis</u> in February 1977 and on the upper 1 m in April 1977 and respective densities were approximately 10/cm² and 3/cm².

Family Chaetophoroaceæ

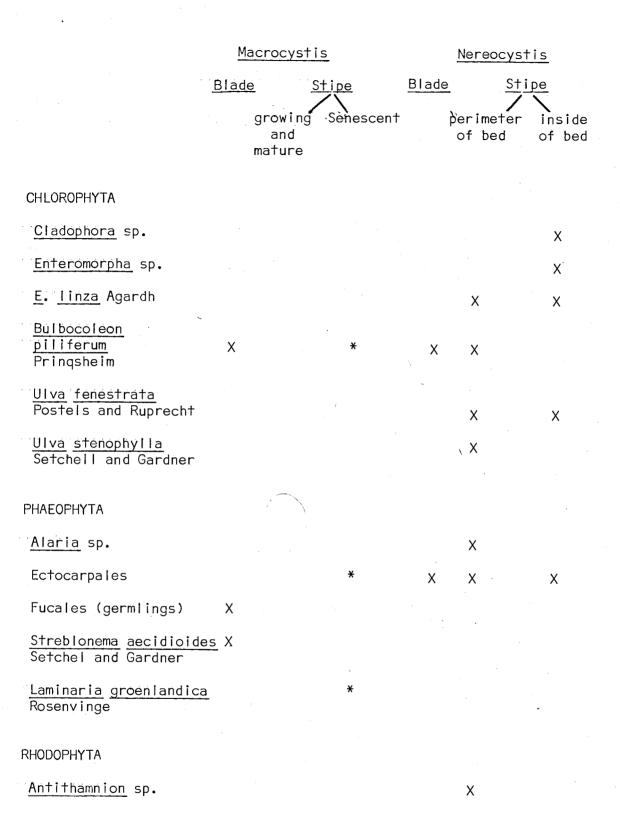
The endophyte <u>Bulbocoleon piliferum</u> was present between the epidermal blade cells of <u>Macrocystis</u> from July 1976 through December 1977 (Fig. 11, 16). Presence or absence/mm² was recorded as individual plants were difficult to discern. Seasonal trends were not evident. From December 1976 through December 1977 mean occurrence/mm² on blades I m or more from the frond meristem ranged from approximately 25 to 100%, and median frequencies were nearly always greater than the mean indicating the commonness of the alga. Variance to mean ratios were nearly always greater than one, indicating a clumped distribution of the endophyte on similarly positioned blades on stipes within the bed, but occasionally (e.g. February 1977) variance to mean ratios were less than one, indicating a more uniform distribution. Frequency of occurrence was positively and significantly associated with increasing distance from the apex meristem. The alga occurred only rarely in stipes of <u>Macrocystis</u> and blades and in stipes of Macrocystis.

Cell's with spores occurred in blades of <u>Macrocystis</u> 2 m or more from the frond meristem. Occasionally <u>Bulbocoleon</u> became so dense that

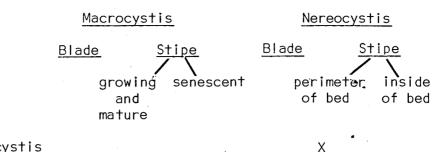
the host epidermal cells eroded (Fig. 16b).

Division Rhodophyta

<u>Porphyra</u> (less than 1 cm long) occurred in February 1977 on <u>Nereocystis</u> from 0.1 to 2 m below the float in respective maximum densities of $5/cm^2$ and $35/cm^2$ (n=4). By April, <u>Porphyra</u> occurred in maximum densities of $0.01/cm^2$ and $5/cm^2$, 0.1 m and 1.0 m respectively from the float (n=2). All large enough to identify were <u>P. nereocystis</u>; and blades in April could be fertile. Nonreproductive, less than 1 cm long <u>Antithamnion</u>-type filaments were present in February and April with maximum densities ($25/cm^2$ and $55/cm^2$ respectively) I to 2 m from the float of <u>Nereocystis</u>. Table I. Epiphytes and endophytes, except bacteria and diatoms, found through sampling (X) and casual observation(*) on the stipes and blades of <u>Macrocystis</u> and <u>Nereocystis</u>. Senescent stipes of <u>Macrocystis</u> which were rarely observed are distinguished from growing and mature stipes found through the regular sampling procedure. Stipes of <u>Nereocystis</u> from inside the bed of <u>Macrocystis</u> are distinguished from those along the perimeter of the bed due to differences in sampling procedure.



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Porphyra nereocystis Anderson

PROTOZOA

Follicullina sp.	*
Suctorians	Х
Vorticella sp.	×
Zoothamnion sp.	×

CNIDARIA

Coryne sp. * Obelia sp. X

Metridium sp.

ECTOPROCTA

<u>Membranipora</u> X <u>serrilamilia</u> Osburn

<u>Hippothoa</u> <u>hyalina</u> Linnaeus

<u>Tricellaria</u> <u>occendentalis</u> Trusk Х

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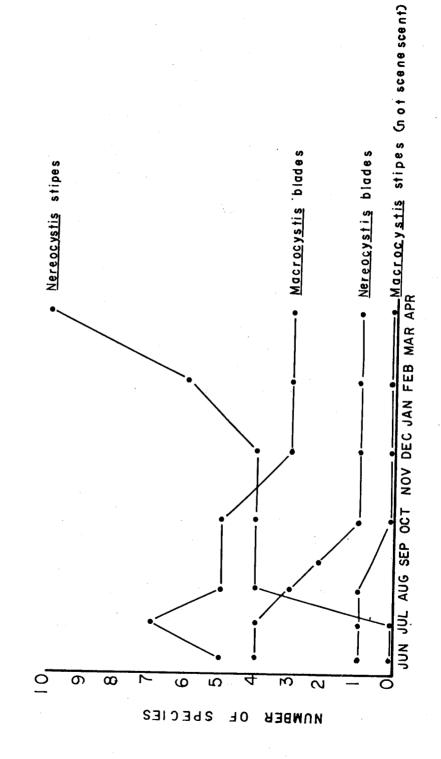
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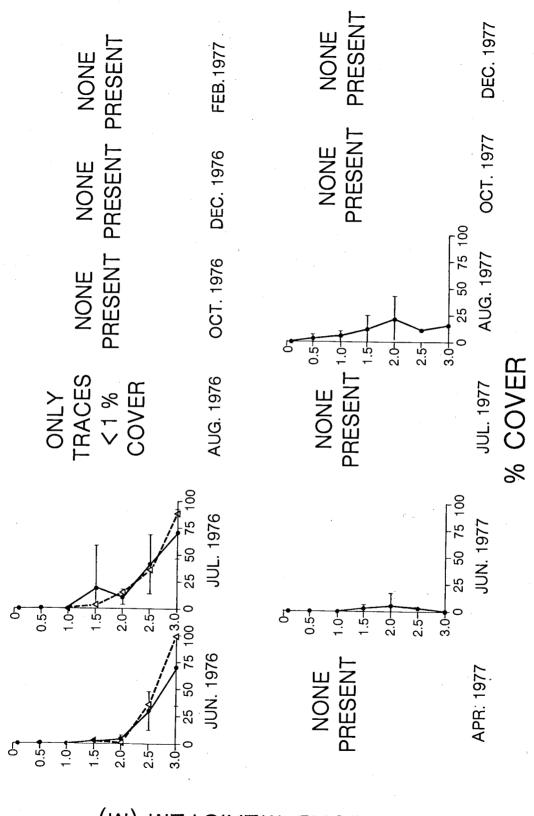
Figure 3. Colonization curves for blades and stipes of

Macrocystis and Nereocystis.



18b

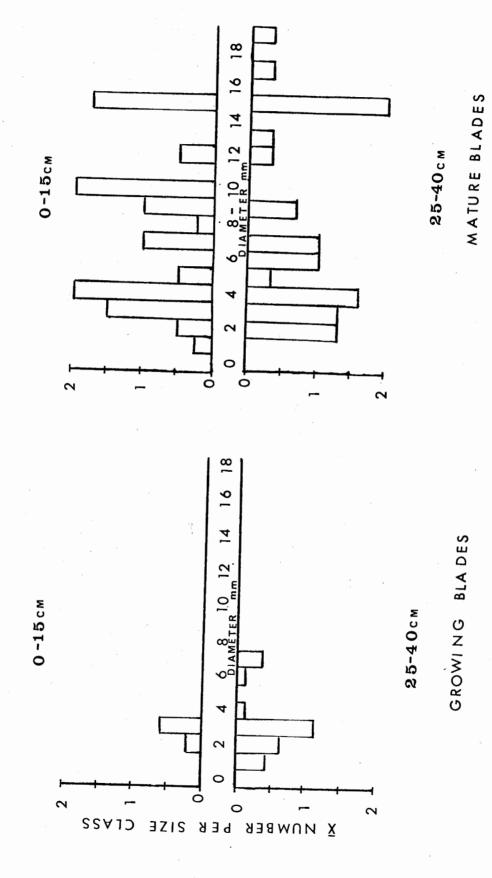
Figure 4. Percent cover of the encrusting bryozoan, <u>Membranipora</u> <u>serrilamella</u> Osburn, on blades of <u>Macrocystis</u>, from June 1976 through December 1977. Solid lines indicate mean values with standard deviations and dashed lines indicate median values.



FROND MERISTEM (M)

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Figure 5. Size class histograms of <u>Membranipora serrilamella</u> Osburn colonies on blades of <u>Macrocystis</u>. 15 x 5 cm samples of blade were taken between 0 and 15 cm from the blade float (n=5), and 25 and 40 cm from the float (n=5) on growing blades between 0.6 and 0.7 m from the apex meristem. Mature to scenescent blades were similarly sampled from 1.7 - 1.8 m from the apex meristem at 0 - 15 cm (n=4) and 25 - 40 cm (4=3) from the blade float.

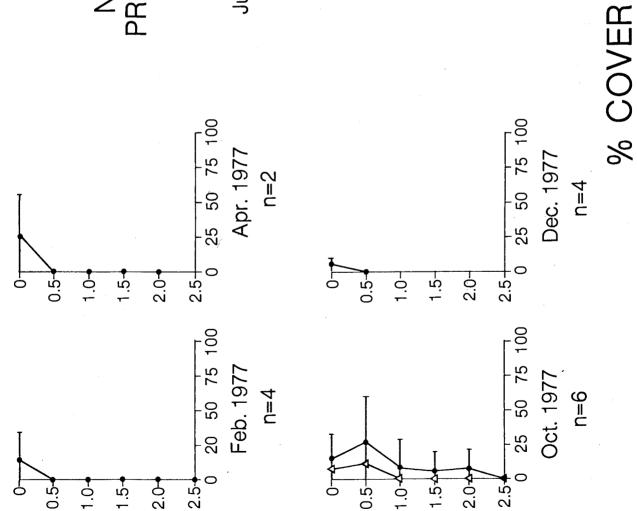


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Figure 6.

Percent cover of the encrusting ectoproct, <u>Membranipora</u> <u>serrilamella</u> Osburn, on stipes of <u>Nereocystis</u> February 1977 through December 1977. Solid points indicate mean values with standard deviations and triangular points indicate median values. The number of stipes sampled is indicated by n.



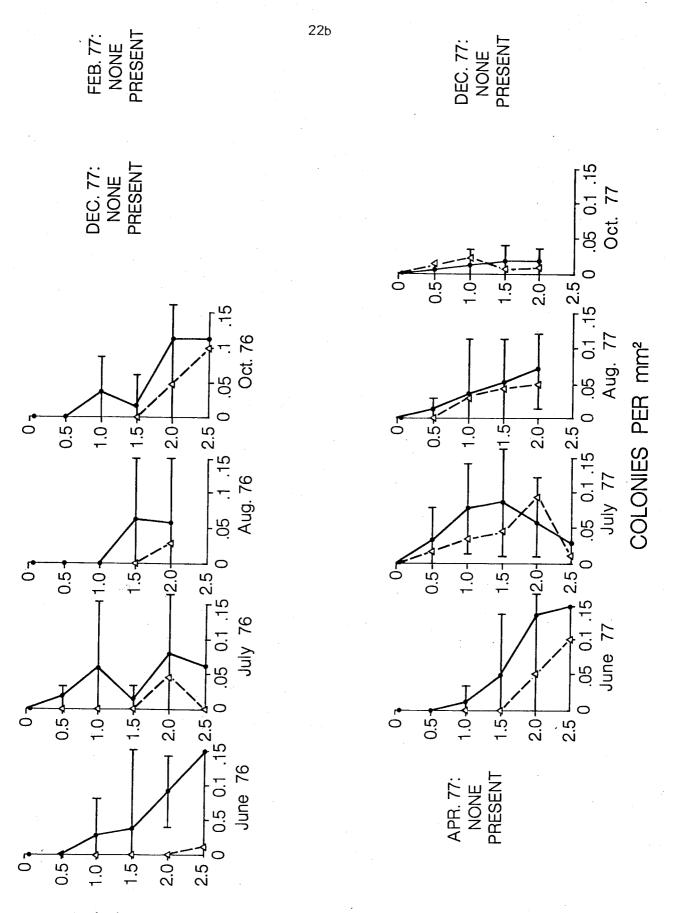


NONE PRESENT

NONE

Jun. 1977 n=6

Aug. 1977 n=6 Figure 7. Density of the hydroid, <u>Obelia</u> sp. (colonies/mm²), on blades of <u>Macrocystis</u> from June 1976 through December 1977. Solid lines indicate mean values with standard deviations and dashed lines indicate median values.



DISTRUCE FROM FROND MERISTEM (m)

Figure 8. Density (nos./mm²) of suctorians on blades of <u>Macrocystis</u> from June 1976 through December 1977. Solid lines indicate mean values with standard deviations and dashed lines indicate median values.

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(M) MATZIRAM DISTANCE FROM FROND

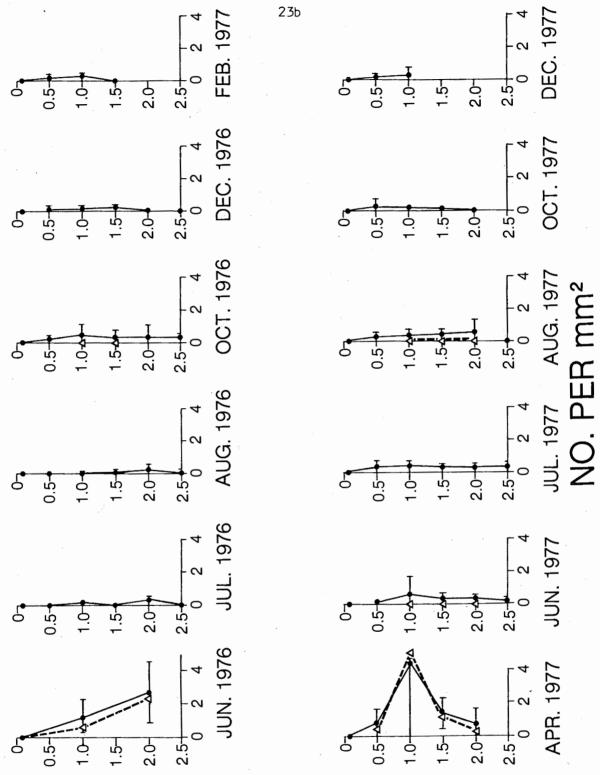
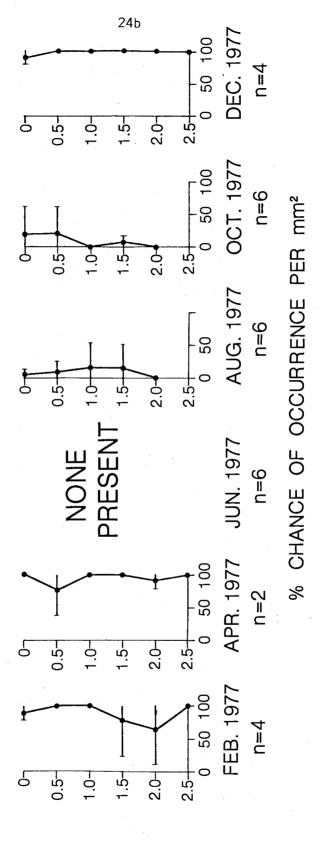


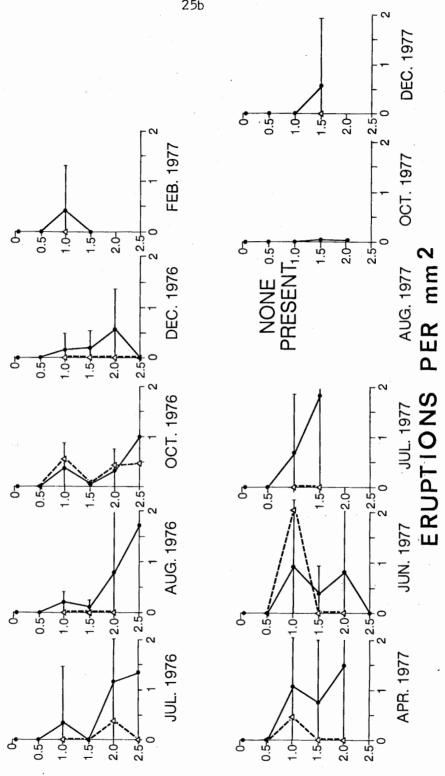
Figure 9.

Percent chance of occurrence per mm² for Ectocarpales on stipes of <u>Nereocystis</u> from February 1977 through December 1977.



PISTANCE FROM (M)

Figure 10. Density (eruptions/mm²) of the endophyte <u>Streblonema</u> aecidioides f. pacificum Setchell and Gardner from July 1976 through December 1977 in blades of Macrocystis. Solid lines indicate mean values with standard deviations and dashed lines are median values.

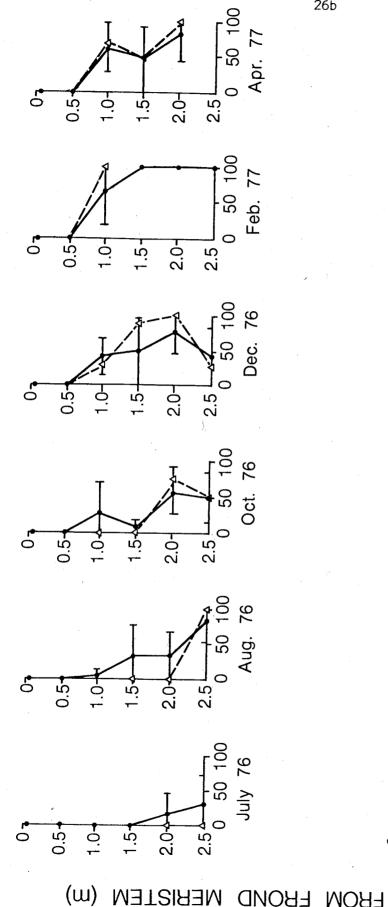


(M) **EROND MERIS** M 11 DISTANCE FROM

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Figure II. Percent chance of occurrence per mm^2 of the endophyte Bulbocoleon piliferum Pringsheim. Solid lines indicate mean values with standard deviations and dashed lines are median values.

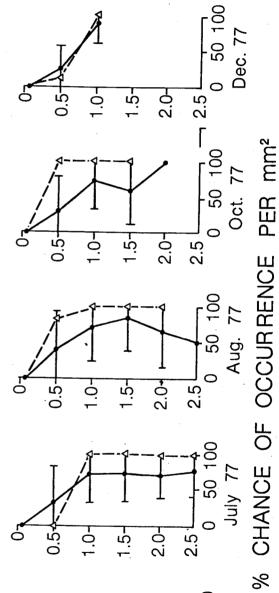
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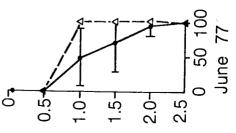


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Figure 12. A. Several colonies of <u>Membranipora</u> <u>serrilamella</u> Osburn encrusting a blade of <u>Macrocystis</u>.

- B. Colonies of <u>Membranipora serrilamella</u> on <u>Macrocystis</u> with central zooids eroded, possibly from being eaten by the nudibranch <u>Doridella</u>.
- C. <u>In situ</u> photograph of <u>Obelia</u> colonies on blades of <u>Macrocystis</u>.
- D. Dense cover of Obelia on blades of Macrocystis.

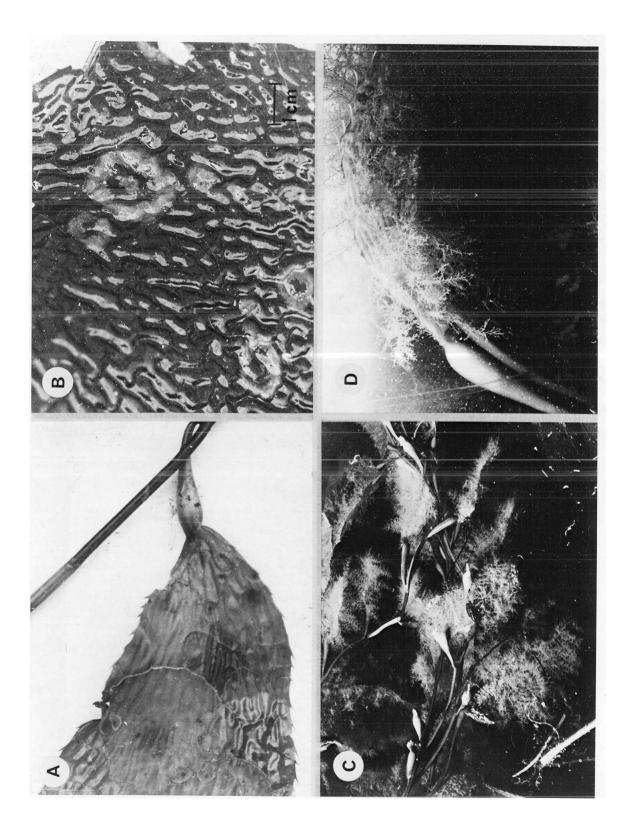
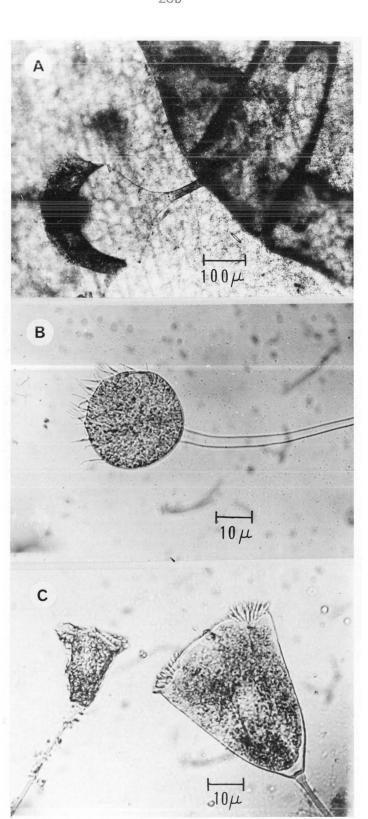


Figure 13.

A. Suctorian, probably <u>Ephelota</u> sp. being overgrown by <u>Membranipora</u>.

B. Suctorian, probably Ephelota sp.

C. Suctorians, probably a species of Acineta.



- A. Senescent stipe of <u>Macrocystis</u> collected in July, 1976 epiphytised with <u>Laminaria groenlandica</u> (arrow) and several other seaweeds.
 - B. Senescent stipe of <u>Nereocystis</u> growing inside the bed of <u>Macrocystis</u>, epiphytised with <u>Membranipora</u> (arrow) and several seaweeds. Collected in July, 1976.
 - C. Mature float and upper portion of a stipe of <u>Nereocystis</u>, epiphytised with Ulvales (arrow) on top of float, and Ectocarpales (appearing as a rough layer) below. This pattern of epiphytism was seen during winter.
 - d. Senescent stipe of <u>Nereocystis</u> collected in April, 1977, epiphytised with Ulvales (solid arrow) and Porphyra nereocystis Anderson (open arrow).

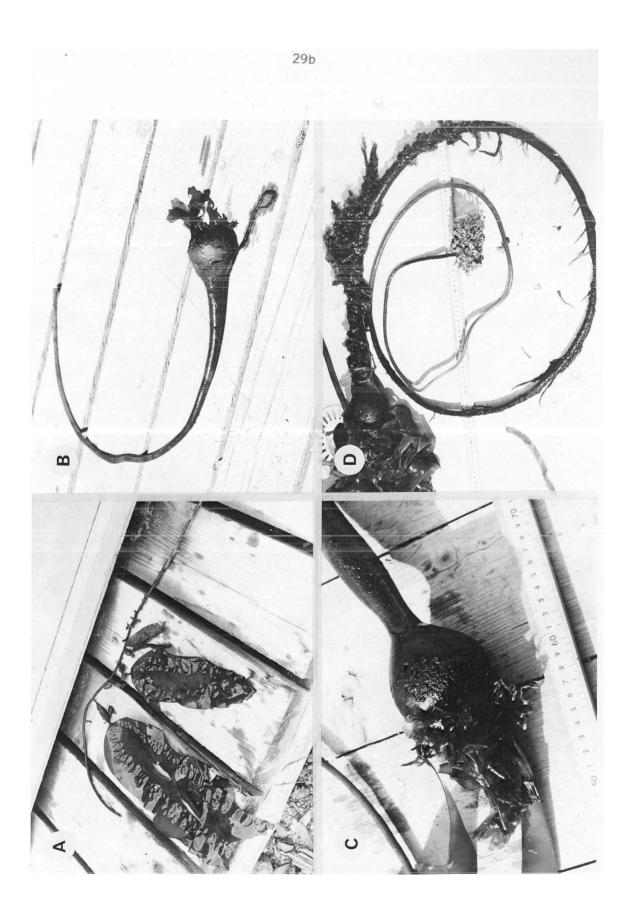


Figure 15. <u>Streblonema aecidioides</u> f. <u>pacificum</u> Setchell and Gardner, an endophytic alga, in blades of <u>Macrocystis</u>. A. Cross section of a plant about to erupt through the epidermis showing penetriation of rhyzoidal filaments into cortex. B. SEM micrograph of mature plant. C. Cross section of immature plant before eruption. D. Cross section of a newly erupted plant.

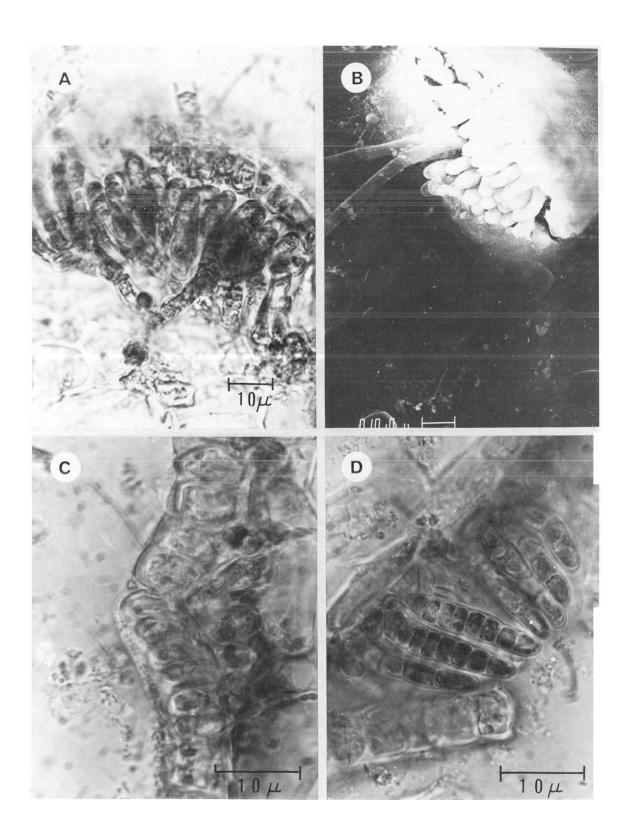
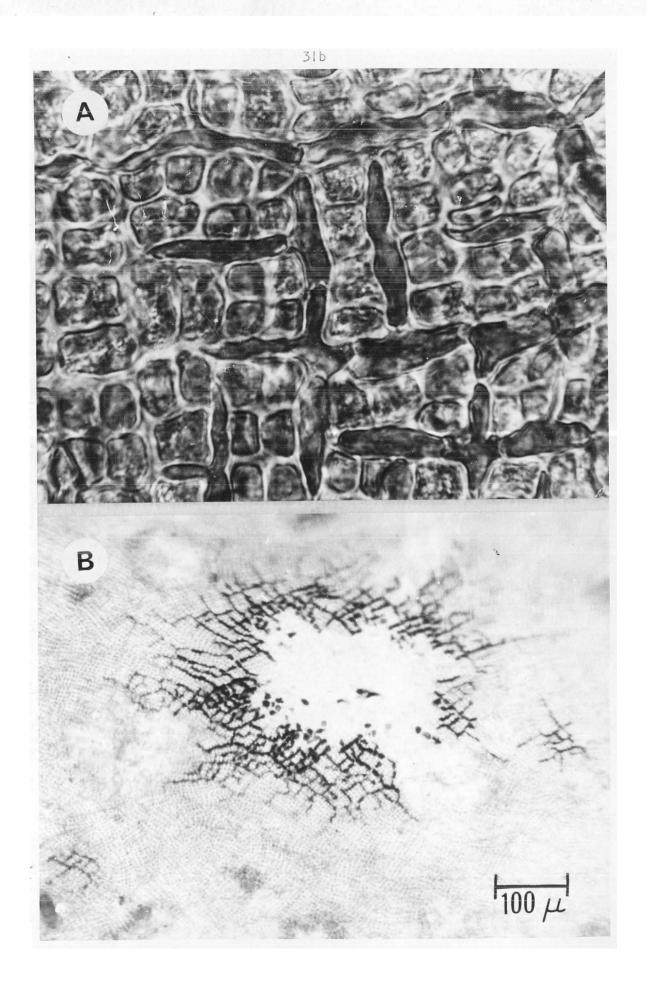


Figure 16.

A. Cells of <u>Bulbocoleon piliferum</u> Pringsheim (arrows) between the epidermal cells of a blade of <u>Macro-</u> <u>cystis</u>. 1 cm = 5µ.

B. <u>Bulbocoleon</u> cells (arrow) which have become so dense the epidermal blade cells of <u>Macrocystis</u> were destroyed.



Diatoms on Macrocystis and Nereocystis Blades

Diatoms separated into 27 taxa on blades of <u>Macrocystis</u> and 18 on blades of <u>Nereocystis</u> (Tables 2 and 3). <u>Cocconets scuteilum</u> had the highest average occurrence (50.8% on <u>Macrocystis</u> and 47.0% on <u>Nereocystis</u>). Niche breadth values (Tables 2 and 3) indicated that <u>Cocconets scutellum</u> and <u>Synedra tabulata</u> were most evenly distributed on <u>Macrocystis</u> and <u>C. scutellum</u>, <u>S. tabulata</u> and <u>Gomphonema</u> no. I were distributed most evenly on <u>Nereocystis</u>.

Densities of diatoms were measured only on blades of <u>Macrocystis</u> (Fig. 17). Total densities on blades 0.1 m from the frond meristem were relatively low (less than 70/mm²), during all seasons, but increased during June of 1976 and 1977 when <u>C. scutellum</u> formed a dense unialgal mat. On blades I and 2 m from the apex low densities (approx. 100 - $200/mm^2$) occurred during August - October 1976 and July - August 1977. These were periods of heightened diversity following a dominance by <u>C. scutellum</u>. Densities were higher at all other times (more than $400/mm^2$). Small standard deviations indicate that the counting procedure was very repeatable. Diatoms were counted using SEM (n=5 tissue samples) and variance to mean ratios were much larger than one indicating a clumped distribution of diatoms on similarly positioned blades within the bed.

The summer of 1976 and 1977 were similar for both host kelps due to a high abundance of <u>C</u>. <u>scutellum</u> and low species diversity. Diatom assemblages in December 1976 and 1977 were similar with <u>C</u>. <u>scutellum</u>, <u>Achnanthes</u> no. 1, and <u>C</u>. <u>costata</u> var <u>pacificum</u> present on both kelps. The seasonal patterns of diversity (Fig. 18) were fairly similar for both kelps (r=0.74, $r_{(10,0.1)}=0.71$) with low diversity in late spring

32

and/or summer and increased diversity in other seasons. Equitability $\left(\begin{array}{c} H'\\ Log_2 N\end{array}\right)$ followed trends in diversity (H') indicating that increases in diversity mainly resulted from a more even distribution of species rather than an increase in numbers of species. However, an increase in species number was responsible for some of the increased diatom diversity during July, August and October 1977 where <u>Macrocystis</u> was the host.

Analysis indicated three clusters of diatom distributions on blades of <u>Macrocystis</u> 1.0 m from the frond meristem (Fig. 19a). June, July and August 1976 and June 1977 assemblages consisted of over 95% <u>C. scutelium</u> with 1 - 3% <u>S. tabulata</u>. Taxa in December 1976, 1977 and February 1977 formed a distinct cluster with a diverse assemblage: <u>C. costata</u> var pacificum (27 - 42%), <u>Achnanthes</u> no. 1 (22 - 45%), <u>C. scutelium</u> (8 - 26%) and <u>S. tabulata</u> (0.2 - 11%). July 1977, August 1977, April 1977 and October 1976 were joined with summer months of both years (when <u>C. scutelium</u> was abundant). This cluster had 20 -40% <u>C. scutelium</u> in common, but within this cluster associations were not close.

Taxa in June, July, and August 1976 formed a cluster on <u>Nereocystis</u>, all having 70 to 90% <u>Cocconeis scutellum</u> in common (Fig, 19b). April and February 1977 taxa were clustered together with 75 and 92% <u>C</u>. <u>costata</u> in common. December 1976 and 1977 taxa formed another cluster with similar abundances of <u>C</u>. <u>scutellum</u> (10 - 11%), <u>Achnanthes</u> no. 1 (26 - 52%), and <u>C</u>. <u>costata</u> var pacificum (23 - 44%).

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Table 2. Seasonal occurrence of diatom taxa on blades <u>Macrocystis</u> 0.1, 1.0, and 2.0 m from the frond meristem.

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TAXON	Niche ** Breadth	⊼ °/₀ ** ± SD	Jun 76 0.1 1.0 2.0	Jul 76 0.1 1.0 2.0	Aug 76 0.1 1.0 2.0	Oct 76 0.1 1.0 2.0	Dec 76 0.1 1.0 2.0	Feb 77 0.1 1.0 2.0*	Apr 77 0.1 1.0 2.0 *	Jun 77 * 0.1 1.0 2.0	Jui 77 0.1 1.0 2.0	Aug 77 0.1 1.0 2.0	Oet 77 0.1 1.0 2.0	Dec 77 0.1 1.0	2.0*
Achnanthes longissipes Ag.	3.22	0.2 ± 0.3	.0 0	000	0 0 0	0 0	0 0	0 0	0	0 03 0	0 1.0 0	2.7 0.7 1.8	0 0.5 0.2	0 0	.
Achnanthes no 1	5.15	9.2 ± 13.7	0 0 0	0 0 0	0 0.8 0	0.5 2.5 5.5	2.6 21.9 45.6	6.0 22.0	0 0.5	0 0 0.6	0 5.8 6.7	0 7.0 4.3	10.0 13.0 37.8	17.0 45.0	
Achnanthes no 2	1.00	0.2 ± 0.4	0 0 0	0	0 0 0	0 0 0	0 0 0	0 0.2	0 0	0 0 0	0 0 0	0 1.5 3.1	000	0	
Amphore spp	3.48	0.5 ± 1.1	0 0 0.1	0 0 0.7	0 0 0	000	0 0.1 0	0 0.9	0 0.2	0 0.2 0	0 0.6 1.6	2.7 3.9 3.1	5.0 0 0	0.3	
Asteromphalus no 1	1.00	0.1±0.2	0 0 0	0 0 0	0 0.2 0	0 0.7 0.4	000	0	0	0.00	0 0 0	0 0	000	0 0	
Biddulphia aurita (Lyng.) Breb.	2.80	0.2 ± 0.5	0 0 0	0 0 0	0 0 0.3	0 0.5 0.2	000	0	0	0 0 0.2	0 0.4 0	0 0 0	0 1.7 0.2	0 0	
Chaetoceros no 1	1.00	0.1 ± 0.3	0 0 0	0 0 0	000	000	0 0 0	0	0	0 0 0	0 1.0 1.1	0 0 0.6	0 0	0 0.1	
Cocconeis costata Gregory	1.95	3.9 ± 6.1	0 0 0	0 0.1 0	0 1.4 0	11.3 7.1 10.1	8.7 19.7 26.2	0 3.1	0 0.7	0 0.4 0.2	20.0 11.6 16.9	2.7 1.7 3.7	0 0.3 1.1	0 1.2	
Cocconeis costata var pacificum Grun.	4.48	14.2 ± 19.1	0 0.2 0.1	0 0 0.4	0 0 7.8	47.4 5.3 5.8	47.8 29.2 18.4	33.0 41.9	4.0 4.4	0.7 0.3 0.2	10.0 1.0 2.2	0 6.8 0	20.0 5.3 4.4	53.6 26.5	
Cocconeis scutellum Ehr.	8.59	50.8 ± 36.4	95.0 95.9 97.7	100 99.2 97.8	99.0 95.4 81.3	35.1 40.0 12.8	36.5 25.9 6.4	61.0 22.5	90.0 67.3	92.7 92.3 90.4	30.3 33.0 40.7	11.1 14.3 23.9	15.0 16.0 12.2	14.3 7.9	
Coscinodiscus spp	1.60	0.6 ± 1.9	0 0 0	0 0	0 0 0	000	0 0	0 0	0 0.2	0 0.1 0	0 0 0.1	0 6.8 0	000	0 6	34
Gomphonema no 1	5.47	1.8 ± 2.6	5.0 0.2 0.3	0 0 0.2	0 0 0.2	0.5 7.0 12.8	0 2.2 3.1	0 5.5	0 5.0	0.7 1.0 2.0	0 2.0 4.4	0 4.4 10.4	15.0 24.3 20.6	3.5 4.0	4ь
Grammatophora marina (Lyngb.) Kutz	3.85	4.6 ± 6.6	0 0.2 0.1	0 0 0	0 0 0.3	0.5 1.5 1.4	0 0 0	0 0.1	0 0.8	0 0.7 0.2	0 5,0 1.6	11.1 6.0 1.5	0 6.9 1.0	0 0.1	
Hyalodiscus subtilis Ehr.	3.49	0.1 ± 0.3	0 0.2 0	000	0 0 0.2	0 0 0	0 0 0	0	0 0	0 0	0 0 0	0 1.2 0	0 0.2 0.2	0 0	
Licmophore grecilis Grun.	1.00	0.1 ± 0.2	0	0 0 0	0 0 0	0 0 0	0 0 0	0 0.1	0 0	0 0	0 0 0	0 0 0	0 0 0	0 0	
Nevicule spp	3.71	3.4 ± 6.6	0 0 0.5	0 0.1 0.4	0 1.4 1.7	2.1 9.3 4.7	0 . 1	0	0 0.9	0 0.4 0	10.1 4.2 2.2	27.8 22.5 19.6	5.0 1.4 1.0	0 0.2	
Nitzschia longissima (Breb.) Ralfs	1.66	0.1 ± 0.5	0 0 0	0 0 0	0 0 0.3	0 0 0	0 0	0	0	0 0 0	10.0 0 0	0 1.7 0.6	000	0 0	
Nitzschie no 1	3.86	1.2 ± 2.1	0 0.2 0.2	0 0 0	0 0 0.3	0 2.3 3.9	0 0 0	0	0 0.4	0 0.5 0.5	0 4.2 0.4	5.5 6.5 0	0 0.7 0.2	0 0	
Nitzschia no 2	3.50	0.2 ± 0.5	0	0 0 0	0 0 0	0 0 0	0 0 0	0	0	0 0.2 0	0 0 0.2	0 1.7 2.5	0 0.5 0.2	0.4	
Meloseire nummuloides Ag.	2.87	0.5± 0.7	000	0 0 0.1	0 0 0	0 2.2 9.7	0 0 0	0	0 D	0 0 0	0 0.6 0	0 1.5 0	0 0.7 0.2	0 0.5	
Pleurosigme strigosum Smith	1.00	0.1±0.3	0 0 0	0 0 0	0	0 0 0	0 0 0	0	0 0.8	0	00.0	0 0.5 0	000	0	
Rhabdoneme arcuatum (Grun.) Hustedt	1.91	0.1 ± 0.1	0 0 0	0 0 0	0 0 0.2	0 0 0	0 0 0	0 0	0 0	0 0 0	0 0.2 0	0 0.3 0	0 0 0	0	
Rhoicosphenia curvata (Kutz.) Grun.	5.26	1.5±3.5	0 0 0.2	0 0.1 0	0 0.3	0 0.3 0.4	0 .1 .2	0 0	0 1.5	0.4 0.3 0.5	0 0.6 1.1	0 2.2 3.7	15.0 12.2 6.3	0	
Skeletonema costatum (Grev.) Cleve.	2.20	3.1±6.7	0 0 0.2	000	0 0 0	0 2.0 20.2	0 0 0	0	0 8.0	0 0 0	0 23.2 5.6	0 0.7 1.8	0 0.2 0.2	0 3.0	
Synedre camtschatice Grun.	1.0	0.1 ± 0.2	0	000	0 0 0	0 0 0	0 0 0	0	0	0 0 0	0 0 0	0 0	0 0 0.2	0 0	
Synedre tebulate (Ag.) Kutz	8.67	7.9 ± 7.6	, 0 2.9 0.4	0 0.7 0.9	1.0 1.0 1.3	2.5 16.6 16.7	· 0 0.2 0.5	0 2.5	6.0 10.2	5.1 3.4 3.6	20.0 5.6 14.2	33.3 11.8 18.4	25.0 18.0 14.2	0.11 01	
and an and a static set over a static static static set of the set			.*												

% OCCURRENCE OF TAXA ON BLADES 0.1, 1.0 and 2.0m FROM FROND MEHISTEM

* Blades were not available for sampling * * Calculated for blades 1.0 M from frond meristem.

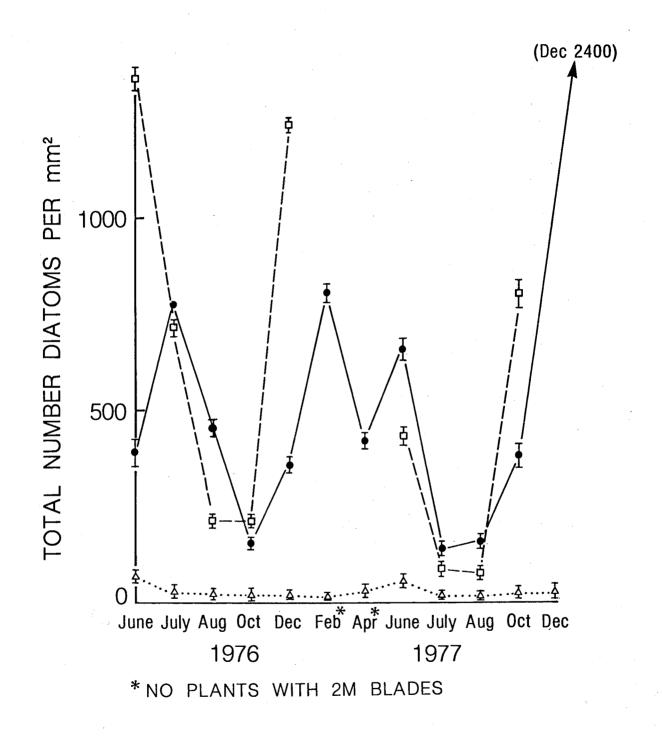
Table 3. Seasonal occurrence of diatom taxa on blades of Nereocystis 0.5 m from the float.

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					%		OCCURRENCE	ц (, 7					
TAXON	Niche Breadth	ž °/o ± SD	Jun 76	Jul 76	Aug 76	0	Dec 76	Feb 77	Apr 77	Jun 77	77 Inf	Oct 77	Dec 77
Achnanthes longissipes Ag-	1.0	0.2 ± 0.4	0	0	1.0	0	0	0	0	0	0	1.2	0
Achinanthes no 1	3.13	8.5 ± 16.4	0	0	0	0	51.6	11.6	1.4	0.6	0	1.86	26.0
Achnanthes no 2	2.75	1.1 ± 2.1	0	0	0.2	0	3.6	6.4	0	1,4	0	0	0
Amphora spp	1.0	0.03 ± 09	0	0	0	0	0	0	0	0	0.3	0	0
Chaetoceros no 1	1.0	0.02 ± 0.06	0	0	0	0	0	0	0	0	0.2	0	0
Cocconeis costata Gregory	3.63	23.6 <u>+</u> 34.8	0	0	0	58.4	0	74.4	92.2	27.8	1.6	0.8	4.4
Cocconeis costata var pacificum Grun.	2.61	6.7 ± 14.0	0	0.4	0.2	0.6	23.0	1.6	0	0	0.6	0.2	43.8
Cocconeis scutellum Ehr.	6.40	47.0 <u>+</u> 38.4	97.0	90.8	97.0	27.8	11.4	2.6	3.2	67.0	60.6	56.2	סרר 10.2
<i>Gomphonema</i> no 1	3.13	5.0 ± 9.6	1.6	0	0	0	1.6	0	0.4	0	21.6	26.6	3.6
Grammatophora marina (Lyngb.) Kutz.	4.81	0.6±0.9	0	0.4	0.2	0.2	0	0	0	0	1.4	2.8	1.2
Hyalodisus subtilis Ehr.	1.0	0.02 ± 0.06	0	0	0	0	0	0		0	0.2	0	0
Licmophora ehrenbergii (Kutz.) Grun.	1.0	0.1 ± 0.24	0	0	0	0.8	0	0	0	0	0	0	0
Licmophora gracilis Grun.	2.32	0.4 ± 0.8	0	0	0	2.2	0	0	0.2	0	0	1.6	0
Navicula spp	7.10	1.9 ± 1.8	0.4	5.0	0	0.4	1.6	0.4	• 0.8	1.7	4.5	3.8	1.8
<i>Nitzschia</i> no 1	1.0	0.05 ± 0.18	0	0	0	0.6	0	0	0	0	0	0	0
<i>Nitzschia</i> no 2	1.0	0.02 ± 0.0.6	0	0	0.2	0	0	0	0	0	0	0	0
Rhoicosphenia curvata (Kutz.) Grun.	5.31	0.6 ± 1.0	0	0	0	2.4	0	1.2	0	0.6	0.1	3.4	1.2
Synedra tabulata (Ag.) Kutz	7.32	4.1 <u>±</u> 4.1	0.8	2.0	0.6	12.2	3.2	1.4	0	1.4	7.6	9.2	7.0

35b

Figure 17. Seasonal variation in diatom densities on blades of <u>Macrocystis</u> 0.1 m (dotted line), 1.0 m (solid line), and 2.0 m (dashed line) from the frond meristem.



36b

Figure 18. Seasonal variation in diversity (H') of diatoms on blades of Macrocystis (solid line), I m from the frond meristem and on blades of Nereocystis (dashed line), 0.5 m from the float. Equitability ($\frac{H'}{Log_2S}$) is plotted above.

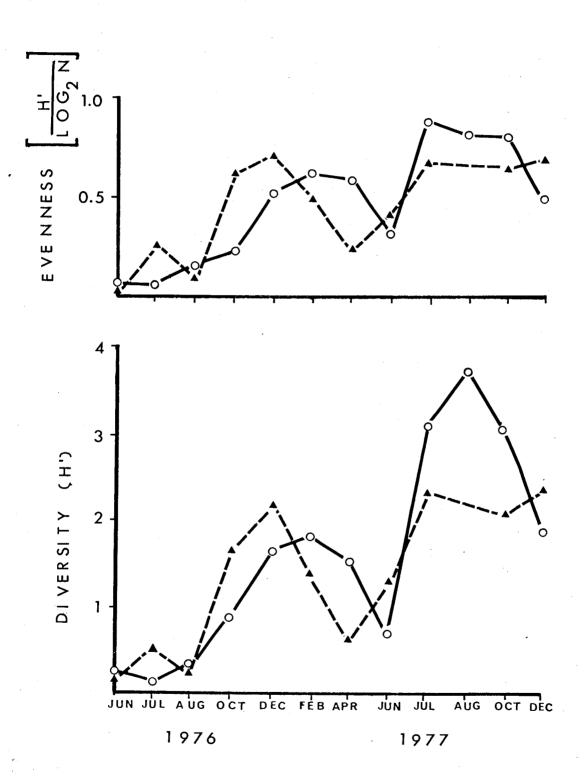
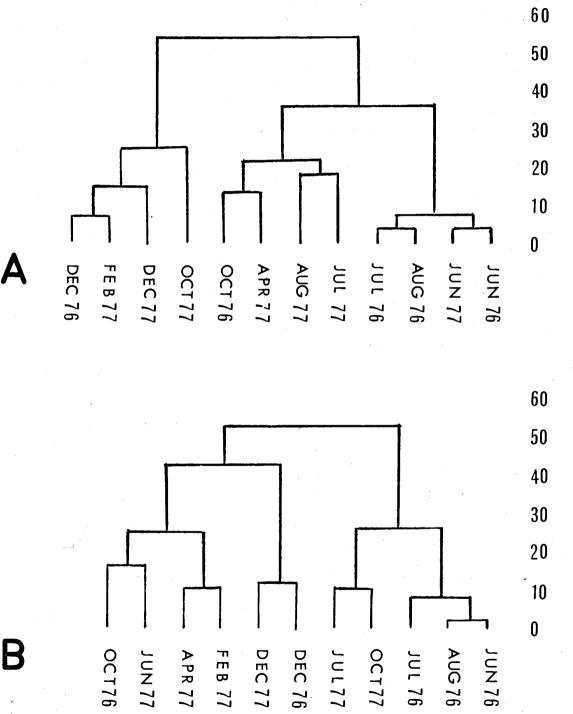


Figure 19. Dendrograms derived from cluster analysis of sampling dates for diatoms on A, Blades of <u>Macrocystis</u> I m from the frond meristem and B, on blades of <u>Nereocystis</u> 50 cm from the float.

38a



A MAL GAMATION DISTANCE

38b

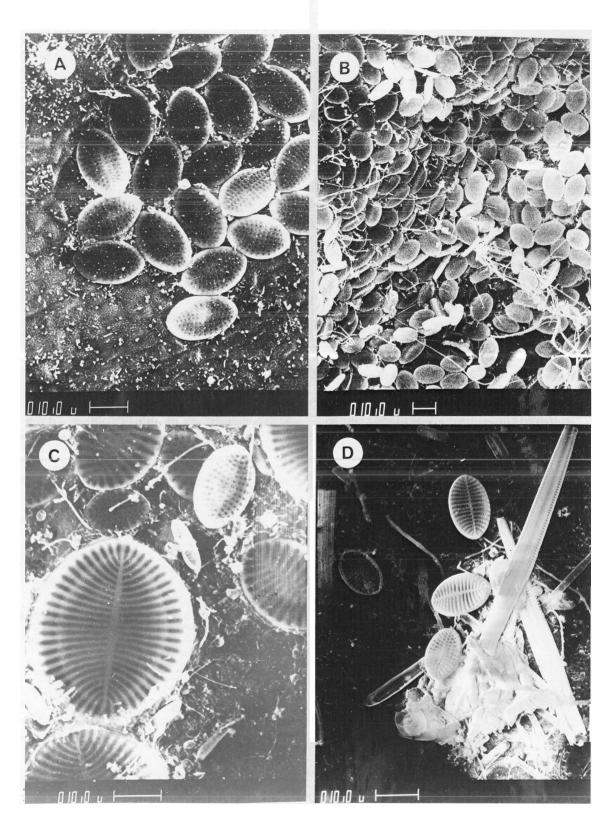
Figure 20. Diatoms on mature Macrocystis blades.

A. <u>Cocconeis scutellum</u> Ehr. in a cluster. Bacteria are visible.

B. Dense cover of Cocconeis scutellum.

- C. <u>Cocconeis costata var pacificum</u> Grun. (large and smaller diatoms with large costae), <u>C. scutellum</u> is in the upper right corner.
- D. Diatoms: <u>Cocconeis costata</u> Gregory, with punctate costae, (upper center), <u>C. scutellum</u> below, and <u>Synedra tabulata</u> (Ag.) Kutz. (long and narrow).

39a



39b

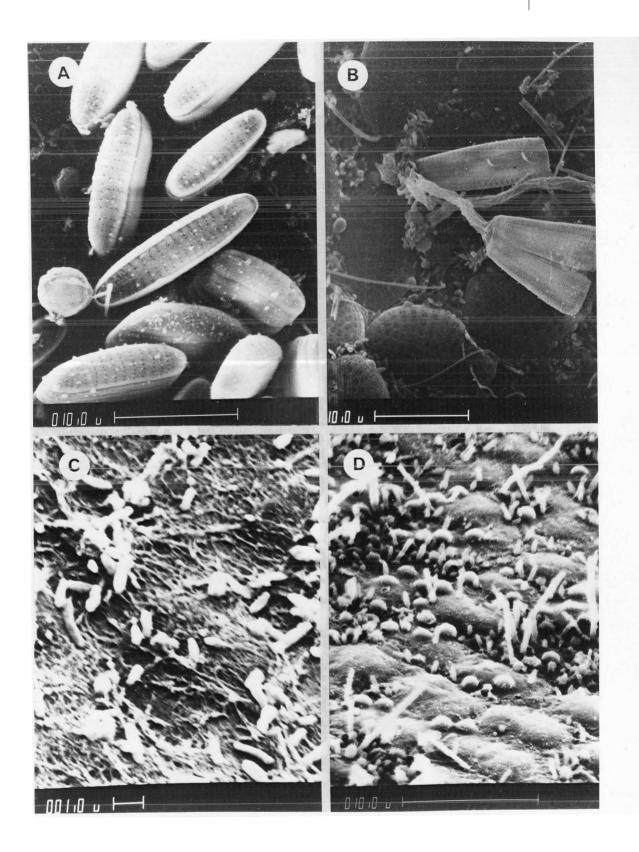
Figure 21. Diatoms and bacteria on Macrocystis.

A. Achnanthes no. I.

B. <u>Gomphonema</u> sp. (stalked).

C. Rod form bacteria on a growing <u>Macrocystis</u> blade.

D. Rod and coccoid form bacteria on a senescent Macrocystis blade.



40b

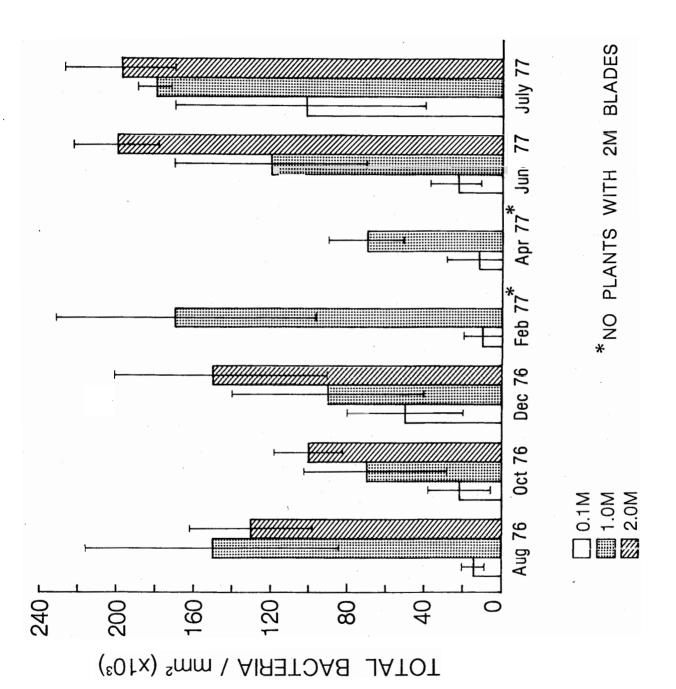
Bacteria on Macrocystis and Nereocystis Blades

Blades of Macrocystis 0.1 m from the frond meristem had an overall mean density of $32 \times 10^3 \pm 32 \times 10^3$ bacteria/mm² and approximately 10 times more rod than coccoid form bacteria (Fig. 21c, 22). Blades 1.0 m from the frond meristem had an overall density of 121 $\times 10^3 \pm 46 \times 10^3$ bacteria/mm² and an approximate 1:1 ratio of rod and coccoid forms. Blades 2.0 m from the frond meristem had an overall mean density of 156 $\times 10^3 \pm 43 \times 10^3$ bacteria/mm² and approximately a 0.6 rod to coccoid ratio (Fig. 21d). No seasonality in density or in rod to coccoid ratio was evident.

Blades of <u>Nereocystis</u> 0.1 and 0.5 m from the float had an overall mean of 9 X $10^3 \pm 7 \times 10^3_{s}$ and 81 X $10^3 \pm 54 \times 10^3_{s}$ bacteria/mm² respectively. Seasonal variations were not evident and the ratios of rod to coccoid form bacteria were approximately 1:1 for blade tissue at 0.1 and 0.5 m from the float.

Figure 22. Seasonal change in total numbers of bacteria/mm 2 on blades of Macrocystis 0.1, 1.0 and 2.0 m from the frond meristem $(\overline{X}\pm s.d.)$

42a



42b

Epiphytism of Artificial Substrates

Glass Slides

Colonization of glass slides (sampled with replacement each collection date) was seasonal and development could be effected by competition. Suctorians were particularly dense from April to June and <u>Obelia</u> was dense from June through October; however the encrusting ectoproct <u>Hippothoa hyalina</u> from June through August could totally cover the slides in the sampling interval forcing suctorian and <u>Obelia</u> populations to extinction. Slides bore very reduced densities of suctorians, <u>Obelia</u> and diatoms through the winter months. <u>Cocconeis</u> <u>scutellum</u>, as on blades of <u>Macrocystis</u> and <u>Nereocystis</u>, formed a near unialgal mat on the slides in summer 1976.

Plastic "Plants"

Bacteria and diatoms initially colonized the plastic. Suctorians were present in relatively low densities throughout the year. An ectocarpalean mat began forming in July 1976. Colonies of <u>Membranipora</u> were rare but <u>Obelia</u> was dense through the summer and fall of 1976, and in winter and spring the colonies disappeared leaving only a network of stolons on the plastic. <u>Ulva</u> was present from July 1976 and <u>Alaria</u> and <u>Porphyra nereocystis</u> occurred in the spring and summer 1977. By October 1977 mussels were common and colonial tunicates covered most of the blades, overgrowing all preceding biota.

DISCUSSION

The pattern of succession on blades, stipes and artificial substrate was similar and proceded in the following order: bacteria and diatoms, staiked ciliates and filamentous algae, hydroids and ectoprocts, structurally simple macro-algae, morphologically complex macro-algae. The population structure and degree of succession were subject to three major influences: I. The time of year was important to colonization. 2. Effects of the host influenced spatial and temporal patterns of epiphytes. 3. Tissue life span was critical to the degree of community development.

Epiphytism and endophytism of <u>Macrocystis</u> and <u>Nereocystis</u> were described for a particular kelp bed. I briefly surveyed other locations through the year over a range of wave exposures and found that the phenomena do vary. Also, a more frequent sampling program would have provided a more complete picture of the seasonal pattern of succession.

Colonization of Hosts

Immigration of epiphytes onto <u>Macrocystis</u> and <u>Nereocystis</u> was seasonal. A spring increase in settlement of <u>Membranipora</u>, suctorians, and <u>Obelia</u> followed by summer or early fall declines in density was observed on blades of <u>Macrocystis</u> and glass slides. Most macroalgae appeared on the stipes of both host species and on plastic plants during spring and early summer. In contrast, total diatom densities did not increase in spring and the number of species markedly declined.

The studies of epiphytism in more temperate waters generally indicate the importance of seasonality. The importance of seasonality to colonization of abiotic marine substrates in temperate climates

similarly is recognized (Scheer, 1945; Aleem, 1950; Lee, 1966; Castenholz, 1967; Sutherland, 1974; Shoener, 1974; Neushal, et al. 1976: Osman, 1977: Sutherland and Karlson, 1977; Emerson and Zedler, 1978: Osman, 1978). Lobban (1976) found that bryozoans and hydroids settled on Macrocystis in Barkley Sound, British Columbia, during the spring and disappeared by September. Markham (1969) reported little change in the epiphytic assemblage on stipes of Nereocystis after December in the vicinity of Friday Harbor, Washington; Laminarales, however, were not present until April. Mukai (1971) found high spring and summer abundances of epiphytes on Sargassum in Japan. Penhale (1977) discovered that epiphyte biomass on eelgrass (Zostera) in North Carolina remained fairly constant through the year. Humm (1964) found a spring decrease in epiphytism on the seagrass Thalassia in Florida and considered this a result of rapid plant growth which allowed less time for the epiphytic assemblage to develop. In the present study highest growth rate of Macrocystis (Lobban, 1977) was during the time of heaviest epiphytism. No correlation between Macrocystis growth rate (from Lobban, 1976) and diatom densities was observed (r=0.34).

The unialgal mat of diatoms in spring (<u>Cocconeis scutellum</u> on <u>Macrocystis</u> and <u>C. scutellum</u> or, <u>C. costata</u> on <u>Nereocystis</u>) was preceded by more diverse assemblages and may have resulted from the superior ability of <u>C. scutellum</u> and other dominant benthic and pelagic primary producers to out-compete other plants in an environment which remained stable long enough for competitive exclusion to occur. Hutchinson (1961) and Titman (1976) discussed such exclusion from competition but I do not know the resources that were in demand. I do not believe that competition for space on the blades determined assemblages since diatoms settling on

newly formed tissue were very similar to assemblages on older tissue. This unialgal population may have declined from over exploitation resources in a previously nutrient enriched environment (Rozenweig, 1971). Such enrichment occurs in temperate climates during spring from high light intensities and nutrient concentrations (O'Brien, 1974). The subsequent assemblages of higher diversity and distantly related species compositions may result from failure of any assemblage to competitively exclude other diatoms before the environment changed significantly (Hutchinson, 1961). The role of grazing in increasing diversity (Paine, 1966; O'Brien, 1971; Connell, 1972; Dayton, 1975; Menge, 1976; and Menge and Sutherland, 1976) seemed unimportant as areas of blade grazed by invertebrates (Druehl, <u>et al</u>, 1977) were recolonized by the same species of diatoms already present on the blades.

Variations in bacterial and endophyte densities on blades of <u>Macrocystis</u> were not seasonal. Although most dense in spring, <u>Cocconeis</u> <u>scutellum</u> had the greatest niche breadth value on both hosts, indicating that it was more evenly distributed than any other diatom throughout all seasons. McIntire and Overton (1971) considered diatoms with broad realized niches to be ecological generalists, and Levins (1968) suggested that a broad niche is optimal in an uncertain environment. Perhaps these organisms have a broad tolerance to environmental factors and/or readily exploit disturbed habitats.

Host Effects

Settlement of organisms onto seaweeds has been related to antibiotic properties of host tissue, (e.g. Sieburth, <u>et al</u>, 1974), age of tissue (Stebbing, 1972), the presence of attractive exudates (Crisp and Williams, 1960), loyality of larvae (Doyle, 1976), preference for certain

host species (Ryland, 1959), and morphology of the host (Ryland, 1959; Edsbagge, 1966).

The importance of the state of tissue development was demonstrated by the stipes of both plants. Nereocystis growing inside the bed of Macrocystis were apparently at a competitive disadvantage and were senescent by June and became heavily epiphytised with bryozoans and seaweeds. In contrast, plants outside the bed Macrocystis reached maturity by August and only subsequently became epiphytised. Stipes of Macrocystis were epiphytised only by microbiota and Membranipora until senescent, after which Obelia and seaweeds could appear. The abundance of slippery exudates on growing tissue of both species compared to older tissue suggests that sloughing or exudates containing antibiotics may help keep tissue clean. McLachlan and Craigie (1964), Sieburth et al (1974), and Hornsey and Hide (1976) found highest antibiotic activity in the growing tissue of the seaweed species examined. The state of tissue development may have been responsible for the high ratio of rod to coccoid form bacteria on growing blades Macrocystis and the high number of coccoid forms on old and senescent blades.

Settlement onto younger tissue, as occurs with <u>Spirobis</u> settling on <u>Laminaria</u> (Stebbing, 1972), may maximise the length of time for development of an organism but my observations did not indicate such a preference for <u>Membranipora serrilamella</u>. Bernstein (per. comm.) found that larvae of <u>Membranipora</u> were positively phototactic and tended to settle on the younger tissue of <u>Macrocystis pyrifera</u> in southern California. This effect may be removed for <u>M. integrifolia</u> because this species grows in waters about three to six times shallower than <u>M. pyrifera</u>.

Absence of <u>Obelia</u>, <u>Streblonema</u>, and several diatom species from <u>Nereocystis</u> may have indicated a preference of these organisms for <u>Macrocystis</u>. The presence of exudates (Crisp and Williams, 1960) or blade morphology may have effected settlement on the two host species. Suctorians, larvae of <u>Membranipora</u>, larvae and primary axes of <u>Obelia</u>, larvae of <u>Hippothoa</u>, larvae of <u>Tricellaria</u>, and <u>Spirorbis</u> were all located in the furrows of <u>Macrocystis</u> blades. Absence of such furrows on blades of <u>Macrocystis</u> may have been the cause of the comparatively less diverse and abundant fauna. Similarly the broyozoan, <u>Celleporelia</u>, was shown by Ryland (1959) to preferentially settle into concavities of the blade of Laminaria saccharina.

None of the epiphytes present on either species were obligate. epiphytes as all grew on glass slides or plastic. Harlin (1973a) found that the supposed obligate epiphyte <u>Smithora</u> would grow on plastic. The endophytes may have been obligate to plant tissue as no alternative morphology was known. Seasonality in occurrence cannot be exhibited by endophytes if no alternate epibenthic form, long lived planktonic phase, or other refuge exists. If plant tissue infected with these endophytes died and new tissue was not infected, extinction of the species would result. This is not the case and I suggest that endophytes have evolved as generalists in response to the short life span of seaweed substrate and to the predictability of formation of new substrate for colonization.

Possible antibiotic effects of host species were discussed earlier, but life history traits of hosts related to epiphytism were not mentioned. The rhizome production of <u>Macrocystis</u> provides a colonial growth strategy, and a single plant (colony of fronds) can prevent being overwhelmed with epiphytism by continually producing new fronds. The importance of

epiphytism in life histories is not known but this growth strategy may have evolved in response to epiphytism, thus allowing a perennial life history strategy. <u>Nereocystis</u> is solitary and becomes heavily epiphytised, thus epiphytism may have been important in the evolution of its annual life history strategy. This supports Jackson's (1977) conclusion that colonial organisms with continual indeterminant growth are less susceptable to fouling.

Host Life Span, Life Histories, and Succession

The life span of the host tissue was important in determining which organisms and assemblages of organisms developed. Abundant organisms on blades (life span of about 2 months) and glass slides were restricted to microbiota and small animals. Only on stipes (life span about 4 - 6 months for Macrocystis and 12 - 14 months for Nereocystis) and plastic did macroalgae become abundant. Osman (1977) discovered that as stability of rocks subject to turnover by waves increased, epifaunal assemblages changed from short lived species to assemblages with longer life spans and superior competitive tactics. Those organisms reproductively and competitively successful on blades and glass slides had more r-ish life history traits (Pianka, 1970) than certain major epiphytes on stipes. A life history trait associated with unstable environments where adult mortality is high is the large investment of energy into production of disseminules (Gadgil and Solbrig, 1972; Gaines, 1974; Giesel, 1976; Stearns, 1977). I did not directly measure energy allocation, however, the speed of development to reproductive maturity (a life history character used by McNaughton, 1975) was relatively rapid and within the host life span for all organisms except

Laminariales, Fucales, and mussels. Reproducing algal species had high surface to volume ratios, simple thalli, and indeterminate growth patterns. Animals were small at maturity and most were colonial. Littler and Murray (1975) reported similar r-type features for an algal assemblage subjected to a periodically toxic sewage outfall.

Mortality was mainly due to the death of the host, but could result from grazing. Diatoms on Macrocystis blades supported a year round population of harpacticoid copepods (Roland, 1978). Membranipora and Obelia were eaten by several species of nudibranchs (Druehl, et al, 1977), and both predator and prey occurred seasonally. Density dependent mortality resulted from the superior competitive ability of certain colonial animals to acquire space by overgrowing others. Solitary forms such as suctorians and Folliculina were overgrown by the colonial ectoprocts. The colonial hydroid Obelia was also overgrown by the ectoprocts Membranipora and Hippothoa but could produce primary axes from overgrown stolons at the interface of two of these encrusting colonies. Jackson (1977) concluded that the indeterminate growth of colonial animals allows continual substrate occupation eliminating the need for recruitment and making them less susceptable to fouling and overgrowth. For colonial animals in the epiphytic habitat recruitment is necessary only on young host tissue.

Studies of succession on marine substrates typically describe a microbial and diatom colonization which is succeeded by more complex organisms, (e.g. Wilson, 1925; Neushul, <u>et al</u>, 1976; Emerson and Zedler, 1978). I saw a similar progression in the epiphytic assemblages on <u>Macrocystis</u> and <u>Nereocystis</u> and Sieburth, <u>et al</u> (1974) noted the same trend in other seaweeds. A true succession where changes in

assemblages are dependent on the presence of other organisms (Connell, 1972) may occur in the epiphytic habitat. An assemblage of a limited biota not repulsed by antibiotic properties may form a crust nullifying antibiotic effects which may allow the colonization to proceed similar to that on inanimate objects (Sieburth, <u>et al</u>, 1974). I observed a succession from organisms with more r-type characteristics to organisms with more K-type life histories when I compared blades to senescent stipes.

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